20

**EXPRESS MAILING CERTIFICATE** 

"EXPRESS MAIL" Mailing Label No.: EL084652896US

Date of Deposit: January 31, 2001

I hereby certify that this paper or fee is being deposited

with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 on the date indicated above and is addressed to the Assistant Commissioner for Patents, Box

Patent Application, Washington, D.C. 20231

Typed or printed name of person signing this certificate:

Signed: Pat Simpson
PAT Simpson

**PATENT** 

### Detection of Methylated CpG Rich Sequences Diagnostic for Malignant Cells

This invention was conducted, at least in part, with government support under National Institutes of Health Grants No: P30 CA16058 and CA80912 awarded by the National Cancer Institute. The U.S. government has certain rights in the invention.

#### **Background of the Invention**

Diagnosis of cancer, classification of tumors, and cancer-patient prognosis all depend on detection of properties inherent to cancer, or malignant cells, that are absent in normal, nonmalignant cells. Since cancer is largely a genetic disease, resulting from and associated with changes in the DNA of cells, one important method of diagnosis is through detection of related changes within the DNA of cancer cells. Such changes can be of two types. The first type of change is a genetic change that occurs when the sequence of nucleotide bases within the DNA is changed. Base changes, deletions and insertions in the DNA are examples of such genetic changes. The second type of change in the DNA is an epigenetic change. Epigenetic changes do not result in nucleotide sequence changes, but rather, result in modification of nucleotide bases. The most common type of epigenetic change is DNA methylation.

In mammalian cells, DNA methylation consists exclusively of addition of a methyl group to the 5-carbon position of cytosine nucleotide bases. In the process, cytosine is changed to 5methylcytosine. Cellular enzymes carry out the methylation events. Only cytosines located 5' to

30

5

10

guanosines in CpG dinucleotides are methylated by the enzymes in mammalian cells. Such CpG dinucleotides are not distributed randomly throughout the genome. Instead, there are regions of mammalian genomes which contain many CpG dinucleotides, while other areas of the genome contain few CpG dinucleotides. Such CpG-rich areas of the genome are called "CpG islands." Most often, CpG islands are located in the transcriptional promoter regions of genes.

Not all CpG islands are methylated. However, the methylation status of CpG islands (i.e., whether the CpG dinucleotides within a particular CpG island are methylated or not) is relatively constant in cells. Nevertheless, the pattern of CpG island methylation can change and, when it does, often a new, relatively stable methylation pattern is established. Such changes in methylation of CpG islands can be either increases or decreases in methylation.

Methylation of CpG islands in the promoter region of a few specific genes has been observed in some types of human cancer. However, at present it is still uncertain whether the methylation status of multiple CpG islands in the genomic DNA of subjects suspected of having cancer can be used as a diagnostic tool for determining whether or not tissue obtained from such subjects contain malignant cells.

#### **Summary Of The Invention**

The present invention relates to methods for identifying CpG islands which are diagnostic of one or more cancers in a subject. The method employs restriction landmark genomic scanning (RLGS) techniques and comprises separately digesting genomic DNA which has been obtained from malignant cells derived from a particular tumor tissue and genomic DNA which has been obtained from control cells derived from healthy tissue with an infrequently cutting restriction enzyme that is not capable of cleaving methylated recognition sites to provide a first set of DNA restriction fragments from the tumor tissue, referred to hereinafter as "malignant cell restriction fragments", and a first set of DNA restriction fragments from the healthy tissue, referred to hereinafter as "control cell restriction fragments"; attaching a detectable label to the ends of the malignant and control cell restriction fragments; digesting the labeled malignant and control cell restriction fragments with a second restriction enzyme; separating each set of restriction fragments on a gel; digesting the restriction fragments in each of the gels with a third more frequently cutting restriction enzyme; electrophoresing each set of restriction fragments in a direction perpendicular to the first direction to provide a first pattern of detectable malignant cell

30

5

10

restriction fragments and a second pattern of detectable control cell restriction fragments; and comparing the second pattern to the first pattern to identify control cell restriction fragments, hereinafter referred to as "diagnostic fragments", which are absent, or exhibit an decreased intensity of label in the first pattern. Such fragments comprise CpG islands that are methylated in the malignant cells. Such patterns are useful for characterizing tissue which is suspected of containing malignant cells. Preferably, each of the diagnostic fragments is then isolated and sequenced, at least in part. In one preferred embodiment, the first restriction enzyme is NotI. In another preferred embodiment, the first restriction enzyme is AscI. Advantageously, the present method permits the detection of numerous methylation sites within the entire genome. In accordance with the present method, applicants have determined that particular CpG islands are preferentially methylated in DNA obtained from tumor tissues of subjects diagnosed as having breast cancer, glioma, acute myeloid leukemia, primitive neuroectodermal tumors of childhood, colon cancer, head and neck cancer, testicular cancer, and lung cancer.

The present invention also provides isolated polynucleotides, referred to hereinafter as "CpG diagnostic polynucleotides", and isolated oligonucleotides referred to hereinafter as "CpG diagnostic oligonucleotides", which are useful for characterizing tissue samples obtained from a subject suspected of having gliomas, acute myeloid leukemia, primitive neuroectodermal tumors of childhood, or cancer of the breast, colon, head and neck, testicle or lung. The CpG diagnostic polynucleotides and oligonucleotidess both comprise a sequence which contains CpG islands that have been shown to be preferentially methylated in DNA that has been obtained from malignant cells of subjects diagnosed as having breast cancer, glioma, acute myeloid leukemia, primitive neuroectodermal tumor of childhood, colon cancer, head and neck cancer, testicular cancer or lung cancer. The CpG diagnostic polynucleotides are from 35 to 3000, preferably, 35 to 100 nucleotides in length, and comprise from 15 to 34, preferably 18 to 25 of the consecutive nucleotides contained with the sequences depicted in the accompanying DNA sequence listing, or sequences which are complementary thereto. The CpG diagnostic polynucleotides comprise two or, preferably, more CpG dinucleotides or dinucleotides which are complementary thereto. The CpG diagnostic oligonucleotides are from 15 to 34 nucleotides in length and comprise from 15 to 34 consecutive nucleotides contained within the sequences depicted in the sequence listing, or sequences which are complementary thereto. The CpG oligonucleotides comprises two or more CpG dinucleotides, or dinucleotides which are complementary thereto.

30

5

10

The present invention also relates to methods which employ the CpG diagnostic polynucleotides and oligonucleotides of the present invention to characterize tissue from patients suspected of having cancer. Such methods are based on the methylation status of CpG islands that have been shown to be preferentially methylated in DNA that has been obtained from tumor tissues of subjects diagnosed as having breast cancer, glioma, acute myeloid leukemia, primitive neuroectodermal tumor of childhood, colon cancer, head and neck cancer, testicular cancer and lung cancer. In one method, DNA which is isolated from suspected tumor tissue from a subject is digested into smaller fragments and reacted with a CpG diagnostic polynucleotide under stringent hybridization conditions. The reaction products are then assayed to determine the size or the sequence of the DNA fragment with which the CpG diagnostic polynucleotide has hybridized. The size or the sequence of the DNA fragment to which the CpG diagnostic polynucleotide has hybridized, hereinafter referred to as the "target DNA fragment", indicates whether the target DNA fragment comprises methylated or non-methylated CpG islands. The presence of methylated CpG islands in the target DNA fragment indicates that the DNA has been obtained from a tumor or neoplasm for which the diagnostic CpG polynucleotide serves as a diagnostic marker.

In another method the DNA from the suspected tumor tissue is treated with a chemical compound which converts non-methylated cytosines to a different nucleotide base. An example of such a compound is sodium bisulfite which converts non-methylated cytosines to uracil. The DNA is then reacted with at CpG diagnostic oligonucleotides under conditions which permit the CpG diagnostic oligonucleotide to hybridize with a complementary sequence in the DNA, referred to hereinafter as the "target sequence". The DNA is also reacted with a modified CpG diagnostic oligonucleotide. The modified CpG diagnostic oligonucleotide comprises a sequence that is complementary to a modified target sequence, i.e., a sequence in which the non-methylated cytosines in the target sequence are converted to a different nucleotide base, e.g. uracil, when treated with a chemical compound. The reaction products are then assayed to determine whether the DNA contains sequences which have hybridized with the CpG diagnostic oligonucleotide or with the modified CpG diagnostic oligonucleotide. Hybridization of the sample DNA with the CpG diagnostic oligonucleotide, as opposed to the modified CpG diagnostic oligonucleotide, indicates that the cytosines in the target sequence are methylated and that the DNA sample has been obtained from a tumor or neoplasm for which the CpG

30

5

10

oligonucleotide has been shown to serve as a diagnostic marker.

The present invention also relates to a method of identifying genes whose expression is increased or decreased in cancer cells.

#### **Brief Description of the Figures**

Fig. 1. Methylation detection in restriction landmark genomic scanning (RLGS) profiles. A, Diagram of the RLGS procedure showing the quantitative nature of methylation detection on NotI fragments displayed on RLGS profiles. Methylation detection in RLGS profiles depends on the methylation sensitivity of the endonuclease activity of NotI. Differences in digestion are assessed by radiolabelling the DNA at cleaved NotI sites. Following further endonuclease digestion, two-dimensional electrophoretic separation and autoradiography, the intensity of a DNA fragment on the resultant RLGS profile quantitatively reflects the copy number and methylation status of the NotI fragment. A priori, this allows NotI fragments containing singlecopy CpG islands to be distinguished from the abundant NotI fragments present in repeat elements and rDNA sequences. B, A portion of an RLGS profile from normal peripheral blood lymphocyte DNA displaying nearly 2,000 single-copy NotI fragments and 15-20 high copynumber fragments. First-dimension separation of labeled Notl/EcoRV fragments extends from right to left horizontally. Following in-gel digestion with Hinfl, the fragments are separated vertically downward into a polyacrylamide gel and autoradiographed. To allow uniform comparisons of RLGS profiles from different samples and different laboratories, each fragment is given a three-variable designation (Y coordinate, X coordinate, fragment number). The central region of the RLGS profile used for all comparisons described in this invention has 28 sections (1-5 vertically and B-G horizontally; the 4G and 5G sections were excluded due to high density and lower resolution of fragments). C, Enlarged view of profile section 2D, showing the numbers assigned to each NotI fragment. D, Analysis of the GC content and CpG ratio {(number of CpGs)/(number of guanines)(number of cytosines)}(number of nucleotides analyzed) of 210 non-redundant Notl/EcoRV clones containing the Notl/HinfI fragments seen in B and in other portions of the RLGS profile. Of 210 clones, 184 clones were randomly chosen and 26 corresponded to fragments which were frequently lost from tumor profiles. CpG islands have a GC content of greater than 50% and a CpG value of 0.6 or greater, relative to bulk DNA (average CG content of 40% and CpG ratio of 0.2). Nucleotide sequences were determined with

30

5

10

greater than 99% accuracy overall. An average of 377nt/clone were analyzed (not indicative of actual CpG island size). The average NotI/EcoRV clone size was approximately 2 kb.

Fig. 2. Fragment loss from RLGS profiles is due to methylation. Top, portions of the RLGS profiles obtained from normal tissue and from two tumors having NotI fragments with either decreased intensity or no change in intensity. Bottom, Southern-blot analysis of EcoRV (NotI: -) and EvoRV/NotI (NotI: +) restriction digested DNAs from a larger number of samples, including the samples at top. In samples without methylation in the NotI site, the probe detects a smaller fragment on double digestion with NotI and EcoRV. The quantitation from multiple Southern blots using a phosphorimager allowed the determination of a lower limit of reliable detection in RLGS profiles of 30% decreased intensity of the diploid NotI/EcoRV fragments. Presence (+) or absence (-) of the corresponding NotI fragment is indicated. N, normal tissue DNA; T, tumor tissue DNA. A, CpG-island locus 3C1 methylation in low-grade gliomas. B, CpG island locus 2C40 methylation in leukemias. C, CpG-island locus 3E24 methylation in PNETs of childhood.

\*, EcoRV fragment of approximately 13 kb with homology to the probe. BLAST searches using the NotI-EcoRV clone sequence identified a homologous BAC clone sequence lacking an internal NotI site, which accounts for the 13-kb fragment on the Southern blot.

Fig. 3. Heterogeneity in CpG-island methylation across tumors. RLGS profiles were generated from 98 primary human tumors and compared with profiles of either matched normal DNA (58 of 98 cases) or to multiple profiles of tissue type-matched normal DNA from unrelated individuals. Loss or decreased intensity of single-copy fragments in the tumors, relative to several neighboring unaltered NotI fragments, were detected by visual inspection of overlaid autoradiographs and confirmed in many cases by independent profiles of the same DNA samples. For each tumor type, the dot plots display the total number of methylated CpG islands (of 1,184 CpG islands analyzed) observed in each tumor. Under the assumption that the tumors are drawn from a homogeneous distribution, with all tumors having the same frequency of methylation, the loss distributions should be approximately Poisson. The colored curve represents the expected distribution. BRE, breast tumors; CLN, colon tumors; GLI, gliomas; HN, head and neck tumors; LEU, acute myeloid leukemias; PNET, primitive neuroectodermal tumors of childhood; TST, testicular tumors.

30

5

10

Fig. 4. Subsets of CpG islands are preferentially methylated. For each tumor type, the histograms display the number of tumors in which the particular CpG islands were methylated. Most of the 1,184 CpG islands were not methylated in any of the tumors (histogram bar at 0 is not shown), but several CpG islands were methylated in multiple tumors. The black line shows the expected distribution under the null hypothesis that the CpG islands have equal frequencies of methylation. Most of the tumor types show significant preferential methylation.

#### **Detailed Description Of The Invention**

In one aspect, the present invention relates to methods for identifying clones within a DNA library that can be used for cancer diagnosis and tumor classification, based on the methylation status of CpG dinucleotides contained within or closely adjacent to the specific clones. Such method employs methylation-sensitive restriction endonucleases (MSREs) and restriction landmark genomic scanning (RLGS) gels to identify new, differentially-methylated CpG islands within malignant cells obtained from patients diagnosed as having cancer. In accordance with the present invention, Applicants have identified 93 clones which can be used to determine whether a tumor biopsy from a patient contains benign or malignant cells.

To carry out such method, tissue (referred to hereinafter as "tumor tissue") which contains a tumor or neoplasm is obtained from a patient known to have a cancer. In some cases, the tumor tissue is obtained from a particular type of solid tumor which has bee surgically removed from the patient. In some cases, the tumor tissue is obtained from the hematopoietic system, such as for example, bone marrow or blood, of the patient. The tumor tissue will have been determined to be from either a benign or malignant tumor or neoplasm.

Separately, tissue (referred to hereinafter as "healthy tissue") which does not contain a tumor or neoplasm is obtained from a subject. The healthy tissue, may be obtained by surgically removing normal tissue from the patient or by surgically removing normal tissue from a healthy control subject who does not have cancer. The healthy tissue may also come from the hematopoietic system, such as for example, bone marrow or blood, of a healthy control subject. The healthy tissue will have been determined to be non-tumorigenic or non-neoplastic.

DNA is then isolated from both the tumor tissue and healthy tissue. If the tumor tissue is a solid tissue sample, such procedure may first comprise separating the individual cells contained

30

5

10

within the tissue from each other. For example, if the tissue samples were frozen after surgical removal from a patient, cells may be separated from one another by grinding the frozen tissue with a mortar and pestle. DNA is then isolated from the individual cells using procedures well known to those skilled in the art. Commonly, such DNA isolation procedures comprise lysis of the individual cells using detergents, for example. After cell lysis, proteins are commonly removed from the DNA using various proteases. The DNA is then commonly extracted with phenol, precipitated in alcohol and dissolved in an aqueous solution.

In the procedures which follow, the DNA obtained from the tumor tissue is treated separately from the DNA obtained from healthy tissue (i.e., the two DNAs are not mixed). The DNAs are separately analyzed using a method called restriction landmark genomic scanning (RLGS). The purpose is to analyze both DNAs separately. The two analyses are then compared in order to identify CpG islands that distinguish cancer cells from normal cells.

Both DNA samples are treated with restriction enzymes and the free ends that result from the restriction enzyme cleavage are labeled. However, since the isolated DNA is in linear pieces, there are free ends that exist before the DNA is cleaved with the restriction enzymes. To prevent these ends from being labeled, the ends, preferably, are blocked before restriction enzyme treatment. Such blocking can be done by addition of dideoxynucleotides and sulfur-substituted nucleotides to the free ends before treatment with restriction enzymes. Subsequently, when the DNA is cleaved by restriction enzymes and labeled, only the ends resulting from the restriction enzyme cleavage will be labeled.

After the reaction to block free ends, the DNA samples are cleaved with a first restriction enzyme that can be characterized as an infrequently cleaving, methylation-sensitive restriction enzyme. Examples of suitable first restriction enzymes are NotI, AscI, BssHII and EagI. As used herein the term "infrequently cleaving" refers to a restriction enzyme that is expected to cleave genomic DNA at intervals greater than 10 kilobases. For example, NotI is an infrequently cleaving restriction enzyme. NotI recognizes a nucleotide sequence of 8 base pairs (bp) in the genome (i.e., 5'GCGGCCGC3') and cleaves the DNA at this site. There are an estimated 4000-5000 of such NotI recognition sequences within the human genome. It is estimated that such recognition sequences are spaced at approximately 1 megabase (Mb) intervals within the genome. In contrast, a frequently cleaving restriction enzyme is expected to cleave the human genome at from 5-10 kb intervals. Such an enzyme will have approximately 100-times more

30

5

10

cleavage sites within the human genome than infrequently-cleaving enzymes. Such frequently cleaving enzymes usually recognize a nucleotide sequence of less than 8 bp in the genome and cleave the DNA at that site. However, not all restriction enzymes that have nucleotide recognition sequences of less than 8 bp are frequently cleaving enzymes. BssHII and EagI both have 6 bp recognition sequences but the recognition sequences for these two enzymes are spaced at intervals within the genome that are greater than 10 kb. "Methylation sensitive" as used herein refers to any enzyme that is unable to cleave DNA at its normal restriction site if one or more nucleotides within the recognition sequence is methylated. For example, the restriction enzyme NotI will cleave the 5'GCGGCCGC3' recognition sequence if the sequence does not contain a 5-methylcytosine. However, the NotI enzyme will not cleave this sequence if any of the cytosines have been methylated to become 5-methylcytosine.

Following digestion of the DNA with the first restriction enzyme, the ends of the DNA fragments are labeled. This can be done, for example, by attachment of nucleotides carrying a detectable label, such as a radiolabel, to the ends of the DNA sample. Typically, attachment is accomplished by filling in the recessed DNA ends left by cleavage with the first restriction enzyme such that the ends become blunt (i.e., non-recessed). Such end-filling reaction may employ deoxynucleoside triphosphates having a radiolabeled phosphate at the  $\alpha$  phosphate position. Such labeled phosphate is preferably  $^{32}$ P.

The labeled fragments from each sample are then cleaved with a second restriction enzyme. Such second restriction enzyme preferably cleaves human DNA at average intervals of between 5-10 kb. Such enzymes normally have a 6 bp recognition sequence. Preferably, the second restriction enzyme is not methylation sensitive. Examples of suitable second restriction enzymes are PstI, PvuI, EcoRV or BamHI. Cleavage of the DNA fragments with the second restriction enzyme provides a second set of fragments, labeled at the ends left by cleavage with the first enzyme. Many of such second fragments are smaller than the fragments resulting from cleavage with the first restriction enzyme.

The DNA fragments are then separated from one another. Preferably this separation is based on size. Preferably this separation is performed by first-dimension electrophoresis through an agarose tube-shaped gel of approximately 60 cm in length.

After electrophoresis through the tube-shaped gel, the DNA is digested within the gel with a third restriction enzyme. Such third restriction enzymes preferably have recognition

30

5

10

sequences of 4 or 6 bp. Such third restriction enzymes also have the property of being able to cleave DNA which is embedded within agarose. One such enzyme is HinfI.

After cleavage by the third restriction enzyme, the DNA is again separated based on size, preferably by electrophoresis through a polyacrylamide gel. Subsequently, the separated DNA fragments are detected based on the labeled ends of the DNA fragments. In those cases where the fragments are radiolabeld, detection is by autoradiography of the two-dimensional gel. Such autoradiography provides a pattern of DNA fragments or "spots." Such pattern is called an RLGS profile.

Each fragment on the RLGS profile obtained from using the DNA from healthy tissues is uniquely identified by its location on the autoradiograph (Y coordinate, X coordinate, fragment number). For each fragment location on the RLGS profile obtained from healthy tissue DNA, the identical location is observed on the RLGS profile obtained from tumor tissue DNA.

In a fragment by fragment comparison of RLGS profiles obtained from tumor tissue DNA with healthy tissue DNA, three different patterns are possible. First, for a given fragment on the healthy tissue RLGS profile, there may be a corresponding fragment at the same location, and of the same intensity, on the tumor tissue RLGS profile. This indicates that the first restriction enzyme cleaved both DNAs at the same sequences (Fig. 1A). This indicates that there were no differences in methylation of the NotI nucleotide recognition sequence of that fragment between the tumor tissue DNA and the healthy tissue DNA.

Second, for a given fragment on the healthy tissue RLGS profile, there may be no fragment at the same location on the tumor tissue RLGS profile. Such a pattern indicates that the first restriction enzyme did not cleave the tumor tissue DNA at the recognition sequence required to produce that specific fragment, but did cleave at such sequence within the healthy tissue DNA (Fig. 1A). This indicates that there was methylation within the NotI recognition sequence in the tumor tissue DNA but not in the healthy tissue DNA.

Third, for a given fragment on the healthy tissue RLGS profile, there may be a corresponding fragment at the same location on the tumor tissue RLGS profile, but the intensity of the fragment may be of decreased intensity. Such a pattern indicates that the first restriction enzyme cleaved one of two copies (i.e., the genome is diploid) of the tumor tissue DNA at the recognition sequence required to produce that specific fragment (Fig. 1A). In healthy tissue DNA, the first restriction enzyme cleaved both copies of the recognition sequence. This

30

5

10

indicates that there was methylation within one of two NotI recogniton sequences in the tumor tissue DNA.

Through comparisons of RLGS profiles obtained from healthy tissue DNA with profiles obtained from a large number of different tumor tissue DNAs, loss of specific fragments in multiple tumors can be associated with a specific type of cancer. Loss of such fragments from RLGS profiles, therefore, can be diagnostic for cancer in a subject. For example, loss of a specific fragment (i.e., methylation of the first restriction enzyme site at the end of said fragment) in a high percentage of tumor tissue DNAs from women known to have breast cancer can be diagnostic for breast cancer in subjects suspected of having the disease. To perform such a diagnostic analysis, DNA isolated from a patient suspected of having breast cancer would be analyzed by RLGS, as described above, to determine whether there was loss of one or more fragments in RLGS profiles that are known to be lost at high frequency in women known to have breast cancer. Similarly, loss of other specific fragments can be diagnostic for other cancers, such as for example, colon cancer, head and neck cancer, lung cancer, testicular cancer, neuroectodermal cancer, gliomas, acute myeloid leukemias, and others.

Loss of a specific fragment in RLGS profiles from multiple tumors can also be diagnostic of several types of cancer, rather than a single type of cancer. For example, loss of a specific fragment can occur in a high percentage of tumor tissue DNAs obtained from individuals with either breast, colon or lung cancer. Loss of such a spot from RLGS profiles using DNA obtained from a patient suspected of having cancer would be diagnostic for either breast, colon or lung cancer in that patient.

#### Isolated Polynucleotides and Oligonucleotides Diagnostic for Cancer

Individual DNA clones that contain the DNA present in each spot or fragment that makes up an RLGS profile can be obtained. This is done by constructing a DNA library of healthy tissue DNA that has been cleaved with the same first and second enzymes used to perform the RLGS gel analysis. Such DNA library will contain individual clones, each clone comprising DNA that is present in a single spot of the RLGS profile. The totality of clones within the library is representative of the combined DNA spots in the RLGS profile.

Individual clones within the library can be identified that contain the DNA of each spot on the RLGS profile. This can be done by taking DNA from one or a few individual clones of the DNA library and mixing it with healthy tissue DNA, before RLGS analysis is begun. When

30

5

10

this mixture of DNAs is used to produce an RLGS profile, the intensity of the spots that contain the same DNA as the individual clones added to the mixture will be increased. By performing multiple analyses of this type, each spot on an RLGS profile can be matched up with a DNA clone within the library. The result of such an analysis is an ordered human genomic library of restriction fragments containing the same subset of genomic fragments as those displayed on RLGS profiles. In such ordered genomic libraries, an individual library clone corresponding to any spot or fragment in an RLGS profile can be rapidly located.

To design diagnostic CpG polynucleotides and oligonucleotides, the sequence of the DNA within each clone (referred to hereinafter as a "diagnostic clone") that corresponds to a spot that is absent or exhibits decreased intensity on the RLGS profile of the DNA from malignant tumor tissue is sequenced using standard techniques. Once sequence information is obtained, regions comprising multiple CpG dinucleotides are located. Such regions serve as the target sequence for the CpG polynucleotides and oligonucleotides.

The CpG polynucleotides are from 35 to 3000, preferably from 35 to 1500 nucleotides in length and comprise two or, preferably, more CpG dinucleotides or dinucleotides which are complementary thereto. The CpG diagnostic oligonucleotides are from 15 to 34 nucleotides, preferably from 18 to 25 nucleotides, in length and comprise at least two CpG dinucleotides or dinucleotides which are complementary thereto. The CpG diagnostic polynucleotides and oligonucleotides each comprise a sequence which is substantially complementary to target sequences containing CpG islands that are known to be preferentially methylated in the DNA from one or more types of cancer cells. "Substantially complementary" means that there is enough complementarity between the CpG diagnostic polynucleotides or oligonucleotides and oligonucleotides under stringent conditions, preferably under highly stringent conditions. Such assays include hybridization assays, such as for example Southern analysis, where the sample DNA is reacted with the CpG diagnostic polynucleotide under stringent hybridization conditions.

The term "stringent conditions, as used herein, is the "stringency" which occurs within a range from about Tm-5 (5 below the melting temperature of the probe) to about 20 C below Tm. "Highly Stringent hybridization conditions" refers to an overnight incubation at 42 degree C in a solution comprising 50% formamide, 5x SSC (750 mM NaCl, 75 mM sodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 g/ml denatured,

5

10

sheared salmon sperm DNA, followed by washing the filters in 0.2x SSC at about 65 degree C. As recognized in the art, stringency conditions can be attained by varying a number of factors such as the length and nature, i.e., DNA or RNA, of the probe; the length and nature of the target sequence, the concentration of the salts and other components, such as formamide, dextran sulfate, and polyethylene glycol, of the hybridization solution. All of these factors may be varied to generate conditions of stringency which are equivalent to the conditions listed above.

Changes in the stringency of hybridization and signal detection are primarily accomplished through the manipulation of formamide concentration (lower percentages of formamide result in lower stringency); salt conditions, or temperature. For example, moderately high stringency conditions include an overnight incubation at 37 degree C in a solution comprising 6X SSPE (20X SSPE = 3M NaCl; 0.2 M NaH<sub>2</sub>PO<sub>4</sub>; 0.02M EDTA, pH 7.4), 0.5% SDS, 30% formamide, 100 ug/ml salmon sperm blocking DNA; followed by washes at 50 degree C with 1XSSPE, 0.1% SDS. In addition, to achieve even lower stringency, washes performed following stringent hybridization can be done at higher salt concentrations (e.g. 5X SSC).

Note that variations in the above conditions may be accomplished through the inclusion and/or substitution of alternate blocking reagents used to suppress background in hybridization experiments. Typical blocking reagents include Denhardt's reagent, BLOTTO, heparin, denatured salmon sperm DNA, and commercially available proprietary formulations. The inclusion of specific blocking reag ents may require modification of the hybridization conditions described above, due to problems with compatibility.

Such assays also include polymerase chain reactions (PCR) where the sample DNA and the diagnostic CpG oligonucleotides are reacted, preferably under conditions which result in the synthesis of a single PCR product. Computer programs, such as for example, the "Primer3" program that can be accessed at "http://genome.wi.mit.edu/cgi-bin/primer/primer\_3www.cgi" can be used to determine the size and sequence of the CpG diagnostic oligonucleotides. Optimum conditions are determined empirically.

The CpG diagnostic polynucleotides and oligonucleotides are made using standard techniques. For example, these polynucleotides and oligonucleotides may be made using commercially available synthesizers.

#### 30 Diagnostic Methods

In another aspect, the present invention relates to methods which use the CpG diagnostic

30

5

10

polynucleotides and oligonucleotides to characterize tissue samples from a subject suspected of having cancer, referred to hereinafter as test sample DNA. To do this, DNA is isolated from the cells of the tissue sample of the patient. Preferably, DNA that serves as a control is also obtained from healthy tissue of the test subject or a control subject as described previously. The diagnostic methods comprise reacting the test sample DNA with the diagnostic CpG polynucleotide or oligonucleotide and assaying the products that are formed as the result of the reaction. In some cases, the sample DNA is digested into smaller fragments prior to reaction with the CpG diagnostic polynucleotides or oligonucleotides. In some cases, a portion of the test sample DNA is first reacted with a chemical compound, such as for example sodium bisulfite, which converts methylated cytosines to a different nucleotide base.

#### Southern Blot Analysis

One such method for diagnosing cancer in a patient involves cleavage of the test sample DNA with a methylation sensitive enzyme, then Southern blot analysis of said cleaved DNA using a CpG diagnostic polyncleotide or oligonucleotide as a probe. For example, the DNA from the patient and the control, healthy tissue DNA are separately cleaved with a methylationsensitive restriction endonuclease, such nuclease being the same first restriction enzyme used to identify the diagnostic spot in the RLGS profile that corresponds to the CpG diagnostic polynucleotide or oligonucleotide. After cleavage, the test sample and control DNAs are electrophoretically separated by size in different lanes of the same agarose gel and blotted to a membrane that can be used in hybridization, such as for example, nitrocellulose or nylon. The membrane is then used in a hybridization reaction with a labeled CpG diagnostic polynucleotide The labeled CpG diagnostic polynucleotide or oligoneucleotide will or oligonucleotide. hybridize to complementary DNA sequences on the membrane. After hybridization, the location on the membrane where the probe hybridized to the control and patient DNAs is visualized. Such locations will identify DNA fragments or bands within the control and patient DNAs containing the same sequence as the CpG diagnostic polynucleotide or oligonucleotide. Hybridization of the probe to a fragment within the patient DNA that is of higher molecular weight than that of the fragment within the control DNA to which the probe hybridized, indicates that a restriction endonuclease cleavage site flanking the target sequence of the CpG diagnostic polynucleotide or oligonucleotide was not cleaved due to methylation. Such result indicates that the tissue is from a cancer for which the CpG diagnostic polynucleotide or oligonucleotide serves

30

5

10

as a diagnostic tool.

A second method for diagnosing cancer in a patient involves cleavage of patient DNA with a methylation-sensitive restriction endonuclease, such nuclease being the same first restriction enzyme used to identify the diagnostic spot in the RLGS profile that corresponds to the fragment. Such nuclease will cleave the patient DNA at the diagnostic recognition sequence only if the DNA is unmethylated. Using nucleotide information derived from sequencing of the library clone corresponding to the diagnostic spot on the RLGS gel, primers for PCR are selected that span the diagnostic recognition sequence. Using the primers, PCR is performed on the DNA. PCR amplification of the sequences will be successful only if the diagnostic nucleotide sequence in the patient DNA had been methylated and was not cleaved by the enzyme. Successful PCR amplification, therefore, is indicative of cancer in the patient.

### Methods Employing a Chemically-Modified DNA Test Sample

Another group of methods for diagnosing cancer in a patient using CpG diagnostic polynucleotides and oligonucleotides are based on treatment of patient DNA with sodium bisulfite which converts all cytosines, but not methylated cytosines, to uracil. The bisulfite converted patient DNA can then be analyzed in a number of different ways. One method of analysis is direct sequencing of the DNA to determine whether the sequence contains cytosine or uracil. Such DNA sequencing requires primers adjacent to the sequenced region to be made. Such primers would be based on DNA sequence information obtained from the diagnostic RLGS spots.

Another method of analyzing bisulfite converted patient DNA is a method called "methylation sensitive PCR" (MSR). In MSR, primers are designed to comprise a sequence which is substantially complementary to the the CpG islands which are known to be preferentially methylated in DNA of cells found in one or more type of tumor tissues. Two sets of PCR primers are made to emcompass this region. One set of primers is designed to be complementary to the sequence that was changed by bisulfite (i.e., cytosines that were originally unmethylated and changed to uracil). As discussed above, these are the modified CpG diagnostic oligonucleotides. A second set of primers is designed to be complementary to the same sequence that was not changed by bisulfite (i.e., cytosines that were methylated and not changed to uracil). As discussed above these are the unmodified CpG diagnostic oligonucleotides, i.e the oligonucleotides which containe at least two CpG dinucleotides or

30

5

10

dinucleotides which are complementary thereto. Two sets of PCR reactions are then run, one reaction with each set of primers, using DNA from the subject as the template. In the case where cytosines within the target sequence of the subject DNA are not methylated, the target sequence will be modified by the chemical reaction and the primers complementary to the modified sequence, i.e., the modified CpG diagnostic oligonucleotides, will produce a PCR reaction product while the primers complementary to the methylated sequence, i.e., the unmodified CpG diagnostic oligonucleotides, will not produce a PCR product. In the case where cytosines within the target sequence of the subject DNA are methylated, the target sequence will not be altered by the reaction with the sodium bisulfite, and the primers complementary to the unaltered sequence, i.e., the unmodified CpG diagnostic oligonucleotides, will produce a PCR reaction product while the modified CpG diagnostic oligonucleotides, which are complementary to the modified target sequence (i.e., unmethylated sequence) will not produce a PCR product

A modification of MSR is bisulfite treatment of patient DNA and PCR amplification of said DNA using primers designed to amplify either methylated or unmethylated sequences. The PCR product is then digested with a restriction enzyme that will cleave or not depending on whether said product contains uracil (rather, thymidine, the complement of uracil; found in PCR product if original patient DNA contained unmethylated cytosine) or cytosine (found in PCR product if original patient DNA contained methylated cytosine).

Another technique referred to as MS-SnuPE, uses bisulfite/PCR followed by primer extension, where incorporation of C (vs. T) denotes methylation.

#### Methods of Identifying Genes

In another aspect of the invention, the CpG diagnostic polynucleotides and oligonucleotides can be used as probes to to identify genes whose expression is increased or decreased in cancerous tissues. To do this, CpG diagnostic polynuceotides are reacted with individual clones of the DNA library. The clones which hybridize with the CpG diagnostic polynucleotide can then be analyzed to determine if they contain an open reading frames that could encode proteins. To determine if the CpG diagnostic polynucleotide hybridizes with the promoter region of a known gene, the open reading frame sequence is analyzed by searching existing DNA databases. For example, GenBank databases can be searched using the BLAST algorithm. If no known genes that correspond to a library clone is found, the sequence can be

10

searched for open reading frames that could encode a protein. Such searching can be performed using commercially available sequence analysis programs commonly known to those skilled in the art. GCG is an example of one such program.

Sequences from clones of the DNA library that contain either known genes or open reading frames can be used as probes to determine whether genes encoded by the sequences are expressed in tumor tissues as compared to control, healthy tissues. To do this, RNA, preferably messenger RNA (mRNA) is isolated from healthy tissue and from tumor tissue from which it is desired to test expression. Such RNA is examined for the presence of expressed transcripts encoded by the sequences obtained from the library. Examination for the presence of expressed transcripts can be performed using a number of methods. One method is Northern blotting where the isolated RNA is separated by size using gel electrophoresis and then blotted to a hybridization membrane. A fragment, polynucleotide or oligonucleotide from the sequence obtained from a library clone is labeled and then used to probe the hybridization membrane containing the size-separated RNA. Detection of hybridization of the probe to the membrane indicates presence of a transcript encoded by the sequence and indicates expression of the gene encoded by that sequence.

Another method to examine isolated RNA for the presence of expressed transcripts is to use RT-PCR analysis. In such analysis, primers are designed and made that span a region of the gene whose expression is to be tested. The isolated RNA is reverse transcribed into DNA using reverse transcriptase. Such DNA is then amplified with the designed primers using PCR. PCR products are visualized after electrophoresis. The presence of PCR products on the gel indicates that the gene encompassed by the designed primers was expressing RNA transcripts. Such analysis can identify and determine genes whose expression is changed in cancer cells as compared to normal, non-cancerous cells.

25

The following examples are for purposes of illustration only and are not intended to limit the scope of the invention as defined in the claims which are appended hereto.

30

5

10

#### **Examples**

# Example 1. Identification of diagnostic markers using NotI and RLGS

### A. Isolation and enzymatic processing of genomic DNA

Tissue from solid tumors was obtained as surgical tissue samples. Where possible, surrounding non-tumor tissue was taken and used as a control. Where it was not possible to obtain patient-matched normal tissue, normal tissue from multiple patients was used. Tissue samples from patients with acute myelogenous leukemis (AML) consisted of either bone marrow aspirates or peripheral blood. Normal samples were obtained from the same patients who were in remission after chemotherapy.

The surgically removed tissues were quickly frozen in liquid nitrogen and stored at -80°C prior to isolation of DNA. When DNA was ready to be isolated, 2 ml of lysis buffer (10 mM Tris, pH 8.0; 150 mM EDTA, 1% sarkosyl) was added to 100-300 mg of tissue in a 50 ml Falcon tube and frozen in liquid nitrogen. The frozen mixture was then removed from the tube, wrapped in aluminum foil, and quickly broken into pieces with a hammer. The broken pieces of cells were transferred to a chilled mortar and ground to a powder with a chilled pestle. For peripheral blood samples, cells were separated on a sterile Histopaque-1077 (SIGMA) gradient and stored at -80°C before DNA isolation.

Cells were transferred to a 50 ml tube and 15-25 ml of lysis buffer containing 0.1 mg proteinase K per ml of lysis buffer was added and mixed using a glass rod. The mixture was incubated at 55°C for 20 min with gentle mixing every 5 min. The mixture was then placed on ice for 10 min. Subsequently, an equal volume of PCI (phenol:chloroform:isoamylalcohol in a ratio of 50:49:1) was added and the tubes containing the mixture were gently rotated for 30-60 min. The tubes were then centrifuged for 30 min at 2500 rpm and the separated, aqueous phase was transferred to a new 50 ml tube using a wide-bore pipette. The PCI extraction was repeated one time. The collected aqueous phase containing the DNA was transferred to dialysis tubing and dialyzed against 4 L of 10 mM Tris, pH 8 for 2 hr. The dialysis tubing was then transferred into fresh 10 mM Tris and dialyzed overnight at room temperature. One additional dialysis was performed in fresh 10 mM Tris for an additional 2 hr. The DNA was then transferred from the dialysis tubing to 50 ml tubes and RNase A was added to a final concentration of 1 μg/ml. The mixture was incubated at 37°C for 2 hr. Subsequently, 2.5 volumes of 100% ethanol were added

30

5

10

to the DNA and the mixture was gently rotated. The insoluble DNA was transferred to a microfuge tube, centrifuged briefly, and the remaining alcohol removed. The pellet was briefly dried in air. The DNA in the pellet was resuspended to a final concentration of 1  $\mu$ g/ $\mu$ l. Such isolated DNA had an average size of 200-300 kb.

The isolated genomic DNA was blocked at ends where the DNA had been sheared. Blocking was done by addition of dideoxynucleotides and sulfur-substituted nucleotides. In a 1.5 ml tube, 7  $\mu$ l of genomic DNA solution was added along with 2.5  $\mu$ l of blocking buffer (1 $\mu$ l 10X buffer 1, 0.1 $\mu$ l 1 M DTT, 0.4  $\mu$ l each of 10  $\mu$ M dGTP $\alpha$ S, 10  $\mu$ M ddATP, 10  $\mu$ M ddTTP, and 0.2 µl 10 µM dCTPaS; buffer 1 consists of 500 mM Tris, pH 7.4, 100 mM MgCl<sub>2</sub>, 1 M NaCl, 10mM DTT) and 0.5 µl DNA polymerase I. The mixture was mixed thoroughly and incubated at 37°C for 20 min. The mixture was then incubated at 65°C for 30 min to inactivate the polymerase. The reaction was then cooled on ice for 2 min. The DNA was digested with NotI by adding to the sample, 8µl of 2.5X buffer 2 (20X buffer 2 is 3 M NaCl, 0.2% Triton X-100, 0.2%BSA) and 2μl (10 U/μl) of NotI. The sample was incubated at 37°C for 2 hr. The DNA was then radioactively labeled. This was done by adding to the sample 0.3  $\mu$ l 1 M DTT, 1  $\mu$ l [ $\alpha$ - $^{32}P$ ]-GTP, 1 µl [ $\alpha$ - $^{32}P$ ]-dCTP and 0.1 1 µl[ $\alpha$ - $^{32}P$ ]-GTP Sequenase ver 2.0 (13 U/µl). The mixture was incubated at 37°C for 30 min. The DNA was then digested with EcoRV by adding to the sample 7.6 µl second enzyme digestion buffer (1 µl 1 mM ddGTP, 1 ul 1 mM ddCTP, 4.4 µl ddH<sub>2</sub>O, 1.2 μl 100 mM MgCl<sub>2</sub>) and 2 μl EcoRV (10 U/μl). The mixture was incubated at 37°C for 1 hr. Then, 7 µl of 6X first-dimension loading dye (0.25% Bromophenol Blue, 0.25% Xylene Cyanol, 15% Ficoll type 400) was added.

### B. First dimension gel set-up and electrophoresis

To make the 60 cm long agarose tube-shaped gel, a gel holder was made. To do this, a sharp razor was used to cut one end of PFA-grade teflon tubing (PFA 11 thin wall, natural; American Plastic, Columbus, Ohio) at an angle to make a bevel. The beveled end of the tubing was fed into glass tubes (4 mm inner diameter, 5 mm outer diameter, 60 cm long). Using a hemostat, the beveled end was pulled up through the tapered end of the glass rod until it protruded 2 to 4 cm. The tubing was cut horizontally at the same end, leaving a 2 mm protrusion (this is the top of the gel holder). The opposite end was cut horizontally, leaving a 5 to 6 cm protrusion from the glass tube. The gel holder was inverted and the top protruding end was

30

5

10

pressed firmly against a hot metal surface (metal spatula heated by a Bunsen burner) to fold the edges of the teflon outward onto the rim of the glass support. A rubber stopper with cored center was pulled over the top end of the gel holder until it was just past the taper of the glass rod. A two-way stopcock was attached to a 10 ml syringe and then to the gel holder via 2 to 3 cm of flexible tubing. The stopcock valve was adjusted to the open position.

Then, to a clean 200 ml glass bottle was added, 60 ml 2X Boyer's buffer (20X is 1 M Tris, 360 mM NaCl, 400 mM sodium acetate, 40 mM EDTA) and 0.48 g Seakem GTG agarose (0.8%). The mixture was heated in a microwave oven until the agarose was dissolved. The mixture was then equilibrated to 55°C in a water bath. With the stopcock valve in the open position, the protruding teflon tube was lowered into the molten agarose solution. The gel solution was suctioned into the gel holder until the gel solution reached 1-2 cm from the top of the gel holder. The stopcock valve was then closed. Keeping the gel upright, the gel was suspended from a ring stand. The gel was allowed to solidify for 20 min.

The stopcock valve was then opened and the syringe and connecting tubes were removed from each gel. After adding 2X Boyer's buffer to the bottom of the first dimension gel apparatus (C.B.S. Scientific), the gels were lowered into the first dimension gel apparatus, seating the rubber stopper firmly into the appropriate holes in the top portion of the apparatus. The top chamber was filled with 2X Boyer's buffer.

Between 1.0-1.5  $\mu g$  of DNA was loaded onto each gel. The sample was electrophoresed at 110 V for 2 hr, and then 230 V for 24 hr.

#### C. In-gel digest

After the DNA was electrophoresed in the first dimension in the agarose tube gel, the DNA was further digested with an additional restriction endonuclease so it could be electrophoresed in the second dimension. In order to perform this additional endonuclease digestion, the buffer and gel holders were removed from the first dimension apparatus. The gel was extruded into a pan containing 1X buffer K (10X buffer K is 200 mM Tris, pH 7.4, 100 mM MgCl2, 1 M NaCl) by forcing the gel out through the bottom of the gel holder. This was accomplished using a 1 ml syringe fitted with a pipet tip and filled with buffer K. The tip was firmly inserted into the top of the gel holder and the plunger depressed until the gel began to come out through the bottom of the gel holder. The 1 ml syringe was replaced with a 5 ml

30

5

10

syringe, and the plunger was depressed until the entire gel was expelled. With a razor, a bevel was cut in the low molecular weight end of the gel and a horizontal cut was made at the high molecular weight end so that the gel was approximately 43 cm in length. The gel length was now the same as the width of the second dimension gel.

The gel was placed into a separate 50 ml tube containing 40 ml of 1X buffer K. The tube was incubated for 10 min at room temperature. The buffer was poured off and the gel incubated in 1X buffer K for an additional 10 min. The buffer K and gel was poured into a pan containing fresh buffer K. Using a 10 ml syringe attached to restriction digest tubing (PFA grade teflon, 9, thin wall, natural; 2.7 mm inner diameter and approximately 3.3 mm outer diameter; American Plastic, Columbus, Ohio), via a 1 to 2 cm segment of flexible tubing, the gel was suctioned into the digest tubing, low molecular weight (beveled) end first. The gel was suctioned into the digest tubing by placing the end of the tubing in line with the beveled end of the gel and pulling the syringe plunger. The tubing was positioned vertically, with the syringe at the bottom and remaining buffer from the tubing was suctioned into the syringe.

In a clean tube, a 1.6 ml mix of 1X HinfI restriction enzyme buffer (50 mM NaCl, 10 mM Tris pH 7.9, 1 mM DTT), 0.1 % BSA, and 750 U of HinfI restriction enzyme was made. The open end of the digest tubing was placed into the tube containing restriction digestion solution. Holding the syringe end up, suction was applied until a small amount of digestion solution appeared in the syringe. The digest tubing was removed and both ends were oriented upward in a U-shape. The syringe was removed and the two ends of the tubing were attached to form a closed circle. This was placed in a moist chamber and incubated at 37°C for 2 hr.

### D. Second dimension electrophoresis

The digested DNA was now run in the second dimension using a 5% non-denaturing acrylamide gel with a 0.8% agarose spacer. To do this, the second dimension gel apparatus (C.B.S. Scientific) was first assembled. All glass plates were cleaned thoroughly and the non-beveled face of each plate was coated with Gelslick or Sigmacote (only once every 10 uses). The back half of the apparatus was laid horizontally on a table top with the upper buffer chamber hanging over the table edge. The two small clear plastic blocks were inserted at the bottom corners of each apparatus. A glass plate was placed in the apparatus, beveled edge facing upward and near the upper buffer chamber, followed by two spacers, one along each side. Glass

30

5

10

plates and spacers were added in this manner until the fifth plate had been added. After the third plate, flexible Tygon tubing was slid down the side channel of the apparatus, with a bevel cut in the leading end of the tubing. The other end was cut, leaving approximately 10 cm protruding from the apparatus. The Plexiglas "filler" sheet was placed over the fifth glass plate. The front half of the apparatus was positioned by aligning the screw holes of the front and back half. These were secured with the teflon screws. The oblong oval "windows" at the lower, front face were sealed with Plastic tape (Scotch brand). The apparatus was stood upright in the lower buffer chamber.

Using a three-way stopcock, the gel apparatus tubing was attached in series with a 2 L reservoir and a 60 ml syringe was attached to the remaining stopcock outlet. The tubing was attached to the 2 L reservoir through a bottom drain (a 2 L graduated cylinder was used). The reservoir was secured above the gel apparatus to allow for gravity flow. The stopcock valve was adjusted to allow liquid to flow between the 2 L reservoir and the 60 ml syringe. Once the TEMED was added, the acrylamide solution (1X TBE, pH 8.3, 96.9 g acrylamide, 3.3 g bisacrylamide, 1.3 g ammonium persulfate and 700 µl TEMED in a total volume of 2 L) was poured into the 2 L reservoir. The syringe plunger was pulled down to the 50 ml mark. The plunger was depressed to push the air out of the upper tubing. Once all air was removed, the valve was adjusted so that all three ports were open. Acrylamide flowed into the apparatus, filling all four gels simultaneously from the bottom upward. The flow was stopped when the level reached 3 mm from the top edge of the glass plates. The solution was allowed to settle for 2 to 3 minutes. After the valve leading to the gel apparatus had been closed, the syringe and reservoir were detached.

The ends of the in-gel digest digest tubing were separated and the first dimension gel was extruded into a pan containing 1X TBE, pH 8.3. The gel was transferred to a 50 ml tube containing 40 ml 1X TBE, pH 8.3. This was incubated for 10 min at room temperature, replaced with fresh TBE, and incubated for an additional 10 min. The first dimension gel was placed in a horizontal position across the beveled edge of each glass plate. Once all gels were in place, the space between the agarose gel and the top of each polyacrylamide gel was filled with molten 0.8% agarose (equilibrated to 55°C). This connecting agarose was allowed to solidify for 10 to 15 min and then 250 µl second dimension loading dye (1X TE, pH 8.3, 0.25% Bromophenol Blue, 0.25% Xylene Cyanol) was added along the length of each gel. Then 1X TBE, pH 8.3 was

5

10

15

added to the upper and lower buffer chambers and electrophoresis was carried out at 100 V for 2 hr and then at 150 V for approximately 24 hr.

Buffers were then removed and the apparatus was disassembled. Each gel was lifted from the plates by overlaying with Whatmann paper cut to size for autoradiographic or phosphorimager cassettes. The perimeter of the paper was traced with the edge of a plastic ruler, removing any excess gel. The Whatmann paper and gel were lifted and placed, gel side up, on a second piece of Whatmann paper. This was overlaid with saran wrap and a third piece of Whatmann paper was added to the top and saran wrap was folded over the top of the Whatmann paper. This was placed in a gel drier, in the same orientation, for 1 hr at 80°C while applying a vacuum. The lower and upper Whatman paper was then removed, saran wrap folded under the remaining paper and exposed to X-ray film (BioMax MS).

## E. RLGS spots resulting from methylation-sensitive restriction enzymes identify CpG islands

Using this methodology, an RLGS profile of DNA from human cells produces a pattern displaying approximately 2,000 spots. Fig. 1B, for example, shows such an RLGS profile from normal peripheral blood lymphocyte DNA. First-dimension separation of labeled NotI/EcoRV fragments extends from right to left horizontally. Following in-gel digestion with HinfI, the fragments were separated vertically downward into a polyacrylamide gel and autoradiographed. To allow uniform comparisons of RLGS profiles, spots were defined based on their location in the gel by assigning each spot a three-variable designation (Y coordinate, X coordinate, fragment number). This can be more easily seen in the enlarged portion of section 2D of the RLGS profile (Fig. 1C) showing the numbers assigned to each spot.

From a set of 1,567 NotI spots comprising the central portion of the RLGS profile of normal DNA, 392 spots were eliminated from all analyses on the basis of having more than diploid intensity, less than diploid intensity, or a degree of positional overlap with neighboring fragments. In addition, a small fraction of loci in individual tumor profiles was not able to be analyzed due to poor local gel quality. In normal DNA profiles, the less-than-diploid copynumber intensities can result from polymorphism, partial methylation or spots derived from sex chromosomes. Thus, the analyzed spots were of diploid copy number in most samples. Tumor tissue and healthy tissue DNA profiles were compared by visual inspection of overlaid autoradiographs. In those cases in which matched normal tissue was not available, tumor

30

5

10

profiles were compared with profiles matched for tissue type of four to five unrelated individuals. Each CpG island was defined as unmethylated or methylated (a visually apparent decrease in intensity on the RLGS profile, which, through corroboration with Southern-blot data for 26 CpG island loci and more that 100 loss events, corresponded to a 30% or greater level of methylation).

To determine if the NotI restriction sites which produced the RLGS spots, had characteristics of authentic CpG islands, DNA from 210 of the NotI/EcoRV RLGS spots was partially sequenced. This was possible because each spot on the human NotI/EcoRV RLGS profile had previously been assigned to a clone from a NotI/EcoRV genomic plasmid library (see description earlier in the specification). Of the 210 spots, 184 were randomly chosen. Another 26 spots were chosen because they were frequently lost from RLGS profiles from human tumors, suggesting that cytosine nucleotides within the NotI sequence of that spot were methylated in the tumor. From the sequences derived from these clones, the GC content (%GC) was plotted against the CpG ratio for each clone (Fig. 1D; CpG ratio = [(number of CpGs)/(number of guanines)(number of cytosines)(number of nucleotides analyzed)]). CpG islands have a GC content of greater than 50% and a CpG value of at least 0.6. Fig. 1D shows that, of 210 clones sequenced, 197 (94%) had sequence characteristics consistent with CpG-island DNA.

### F. Tumor tissue samples analyzed

DNA used to perform the RLGS analyses was obtained from 98 primary human tumors and, where possible, matched normal samples. These samples were from 8 broad tumor types; breast, colon, gliomas, head and neck, acute myeloid leukemias, primitive neuroectodermal tumors (PNETs) and testicular.

Fourteen breast cancers included 2 adenocarcinomas, 2 lobular carcinomas and 10 ductal carcinomas. The samples were from obtained the Cooperative Human Tissue Network (CHTN). All tumors were from females, 38-89 years of age (average of 54 years). Breast tissue adjacent to the tumor was available for 6 of 14 cases, and 8 tumor profiles were compared with 4 breast samples from the matched sets.

Colon tumors were obtained from Roswell Park Cancer Institute and classified according the American Joint Committee on Cancer staging manual. The 8 primary tumors included 1 stage I tumor, 2 stage II tumors, 2 stage III tumors and 3 stage IV tumors. Patient ages ranged

30

5

10

from 49 to 77 years (average of 63 years). Normal adjacent colon mucosa samples were obtained for all tumors.

Fourteen gliomas, including 12 World Health Organization (WHO) grade II astrocytomas and 2 WHO grade III anaplastic astrocytomas, from Saitama Medical School, the University of Tokyo, Teikyo University School of Medicine, Komagome Metropolitan Hospital and the University of Washington, Seattle. Patients included 10 females and 4 males with an age range of 7-57 years (average of 34 years). Brain tissue adjacent to the tumor was also obtained for 1 WHO grade II and 1 WHO grade III tumor. Twelve cases were compared with 3 unmatched normal brain samples and with the 2 brain samples from the matched sets.

Fourteen head and neck squamous cell carcinomas were obtained through the CHTN. Tumors were from 11 males and 3 females. Patients were 42-77 years of age (average of 57 years). Tissue adjacent to the tumor was available for 12 of 14 cases, and 2 tumors were compared with 4 samples from the matched sets.

Nineteen acute myelogenous leukemia samples (3 bone marrow aspirates and 14 peripheral blood) from the Cancer and Leukemia Group B Tissue Bank. Samples were classified according to the French-American-British system. Samples were obtained from patients at the time of initial diagnosis with AML and again at complete remission (24-154 days, average 45 days) after induction chemotherapy. Samples were from 14 males and 3 females. Patients were 22-61 years of age (average 40 years). All cases were compared with matched samples (either peripheral blood lymphocytes or bone marrow, but always matched with the origin of the cancer sample) obtained at remission.

Twenty-two PNETs, including 17 medulloblastomas and 5 supratentorial PNETs, through the CHTN, Pediatric Division. Tumors were from 15 males and 7 females. Patients were 2-26 years of age, with peak ages between 3 and 6 years. All tumors were WHO grade IV. Matched peripheral blood lymphocytes were available for 6 of 22 cases, and 18 samples were compared with unmatched normal cerebellum DNA.

Nine testicular tumors included 6 seminomas and three nonseminomas. Samples were obtained from the Norwegian Radium Hospital and from the Helsinki University Central Hospital. Patients were 21-77 years (average of 41 years). Adjacent testicular tissue was available for 7 of 9 cases, and 2 samples were compared with 4 samples of testicular DNA used in the matched sets.

30

5

10

### G. Loss of spots from RLGS profiles is due to methylation

In comparing RLGS profiles of DNAs from different tumors with control, healthy tissue DNAs, loss of a fragment or spot from an RLGS profile (Fig. 1A) was frequently detected. Loss of such a spot could be due to either methylation of DNA sequences at the NotI site giving rise to that spot, or to deletion of DNA surrounding that NotI site from the genome of the tumor. The relative contribution of each mechanism was assessed by using clones from the NotI/EcoRV genomic library, specific for lost spots, as probes in Southern blotting studies. In Fig. 2A, a section of an RLGS profile, from normal, healthy tissue was compared with tumor tissue from two gliomas, J7 and J16. This RLGS section contains spot 3C1. In tumor J16, spot 3C1 is absent from the RLGS profile. If there was a deletion of DNA surrounding the NotI site, however, the expected result in the Southern blot would be either no hybridization of the probe to the J16 genomic DNA or hybridization to a band of a size different from those detected in the lane containing normal, healthy tissue DNA digested with NotI plus EcoRV, and tumor tissue DNA digested with EcoRV alone. This result is not seen. These results show, therefore, that DNA corresponding to a missing 3C1 spot in J16 glioma DNA is present in the genome, as shown by the Southern hybridization result.

Likewise, DNA corresponding to specific RLGS spots missing in certain leukemias (Fig. 2B) and neuroectodermal tumors of childhood (Fig. 2C) are found to be present when these DNA are analyzed by Southern blotting. Overall, in 26 tumors where specific spots in RLGS profiles were missing DNA corresponding to the spot, was found to be present in the genome by Southern blotting. These results show that loss of spots on RLGS profiles is due to methylation of the corresponding NotI site and not deletion from the genome of DNA representing that spot. Therefore, methylation is the predominant mechanism underlying loss of spots from RLGS profiles.

## H. Heterogeneity in CpG-island methylation across tumors.

To compare the overall pattern of methylated CpG islands among different tumors of the same tumor type, 1,184 spots in each of 98 tumors (and their non-tumorigenic controls) were analyzed by RLGS. The analysis was performed by determining the number of RLGS spots lost, or of decreased intensity, as compared to the controls. Each lost spot or spot of decreased

5

10

intensity is equivalent to one methylated CpG island. For each tumor type, the number of methylated CpG islands in each individual tumor, as compared to controls, was plotted (Fig. 3). These data showed that breast, head and neck, and testicular tumors had relatively low levels of methylation, with many such tumors showing no methylation. Colon tumors, gliomas, acute myeloid leukemias and primitive neuroectodermal tumors (PNETs) had a much higher frequency of methylation. Nonparametric comparison (Kruskal-Wallis procedure) of the methylation frequencies of the various tumor types showed significant differences between them ( $\chi_6^2$ =56.9, P<0.0001).

Within a tumor type, the range of methylated CpG islands in individual tumors was variable. The data (Fig. 3) are not consistent with chance variation between tumors because, in the absence of heterogeneity, the variance of the methylation frequency would not be expected to be greater than the mean<sup>1</sup>. A formal test of this overdispersion was performed for each tumor type and the results are shown in Fig. 3 as a superimposition of the expected Poisson distribution on the dot plots. These data showed that aberrant methylation of CpG islands can be quantitatively different in individual tumors within a tumor type and more pronounced overall in particular tumor types.

### I. Subsets of CpG islands were preferentially methylated in tumors

Through analysis of the RLGS spots lost in different tumors, it was determined that certain spots on the RLGS gels were lost in multiple tumors. This means that specific CpG dinucleotides were methylated in more than one tumor. This is shown in Fig. 4 where the number of tumors within a specific tumor type that had a particular CpG island methylated are shown.

To test the hypothesis that methylation of these common CpG islands was not random, a standard goodness-of-fit test was used.<sup>2</sup> This can be seen in the plots of Fig. 4 where the black

<sup>&</sup>lt;sup>1</sup>Heterogeneity of methylation frequencies across samples was assessed within each tumor type by a standard test for evidence that the variance in methylation frequency exceeds the mean. This test is motivated by the Poisson approximation, which applies even if the frequencies of methylation vary across CpG islands. Moreover, a simple result from the binomial distribution shows that the test is conservative, because under homogeneity the population variance cannot exceed the mean.

<sup>&</sup>lt;sup>2</sup>Under the null hypothesis of equal methylation frequencies for each CpG island, a goodness-offit test ( $\chi^2$ ) was applied to the observed versus expected frequencies of islands exhibiting

30

5

10

line of each plot shows the expected distributions if methylation of specific CpG islands in multiple tumors was random. It can be seen from Fig. 4 that for breast tumors, colon tumors, gliomas, acute myeloid leukemias and childhood PNETs, the actual distributions were significantly different (P<0.0001) from the theoretical distributions indicative of randomness. Similarly, the results for head and neck tumors were significant (P<0.025). The results for testicular tumors (P=0.365) were not significant. However, tumors of this type have a low overall methylation frequency and larger sample sizes are needed. Overall, the data indicate that the patterns of CpG island methylation in tumors is not random.

## J. Frequencies of aberrant CpG-island methylation of shared and tumor-type-specific targets

Because the data have shown that they are methylated in a nonrandom fashion, CpG islands that are methylated at a high frequency in one or more tumor types can be used for diagnosis of tumors. From analysis of 98 tumors using Notl/EcoRV RLGS analysis, a number of spots that are absent or of decreased intensity, as compared to control healthy tissue DNA, have been found. Table I lists these spots. Each fragment (CpG island) is identified in three ways in the table. First, the location of each CpG island is designated as the distance (in cm) migrated during electrophoresis, from the gel origin, in both the first dimension and the second dimension. Second, each CpG island is given a three-variable designation (Y coordinate, X coordinate, fragment number). The X coordinate indicates horizontal direction on the two-dimensional RLGS profile and is a letter from B-G. The Y coordinate indicates vertical direction and is a number from 1-5. Together, an X and Y designation divide the RLGS profile into 28 sections. Within each section, the spots/fragments are given a number. Such a profile is available at: http://pandora.med.ohio-state.edu/masterRLGS/. Third, the partial DNA sequence of individual spots has been determined by sequencing of library clones corresponding to each spot. These sequences are shown in the attached Sequence Listing and have been assigned SEQ ID NOS. from 1 to 82.

The diagnostic NotI/EcoRV spots are of two types (Fig. 1). The first type of spot is absent or of decreased intensity in a single tumor type. For example, the NotI site that is part of the CpG island designated 2.B.53, is methylated only in head and neck tumors. Similarly, the NotI site of CpG island 2.F.2 is methylated only in breast tumors.

The second type of spot is absent or of decreased intensity in more than one type of tumor. For example, the NotI/EcoRV spot designated 2.C.24 is missing in gliomas and AMLs. Similarly, the NotI/EcoRV spot designated 3.B.55 is methylated in breast, colon and PNETs.

### 5 Table I. Diagnostic CpG islands in tumors.

CpG	1st-D	2nd-D	Type <sup>3</sup>	Methylated
Island <sup>1</sup>	(cm) <sup>2</sup>	(cm) <sup>2</sup>		In <sup>4</sup> :
2.B.53	36.85	9.25	t	HN
2.C.24	30.3	5.32	s	Abt/Leu
2.C.29	27.8	5.45	S	Leu/Hn
2.C.35	29.45	6.9	S	Abt/Bre/Cln/Leu/Pbt
2.C.54	32.38	9.42	S	Leu/Hn
2.C.57	30.9	8.5	ND	Tst
2.C.58	31.2	9.2	S	Abt/Leu
2.C.59	30.4	9.35	ND	Hn
2.D.10	27.55	5.3	S	Leu/Pbt
2.D.14	24.25	4.47	t	Leu
2.D.20	26.3	5.3	t	Cln
2.D.25	27.15	6.4	ND	Bre
2.D.27	25.65	5.82	ND	Hn
2.D.34	23.62	6.6	S	Leu/Pbt
2.D.40	23.95	7.25	ND	Pbt
2.D.48	26.1	8.1	ND	Leu
2.D.55	24.2	8.3	S	Cln/Leu
2.D.74	23.95	9.35	S	Abt/Bre/Cln/Leu
2.E.20	20.6	5.95	ND	Pbt
2.E.24	19.35	5.7	S	Abt/Leu
2.E.25	18.27	5.65	t	Bre
2.E.30	20.35	6.4	S	Abt/Bre/Leu
2.E.37	21.42	7.1	ND	Bre
2.E.4	21.1	4.48	S	Leu/Pbt
2.E.40	NA	NA	ND	Tst
2.E.61	19.4	8.08	S	Abt/Pbt
2.E.64	20.5	8.35	S	Abt/Cln
2.F.2	17.27	4.72	t	Bre
2.F.41	NA	NA	t	Tst
2.F.50	15.23	7	S	Abt/Leu
2.F.59	17.49	8	ND	Bre
2.F.70	15.88	13.3	S	Pbt/Tst
2.G.10	10.29	4.49	S	Leu/Tst
2.G.108	7.68	7.44	ND	Bre
3.B.30	35.4	12.55	ND	Tst

3.B.36	34.2	11.8	s	Abt/Cln/Leu/Pbt
3.B.55	NA	NA NA	s	Bre/Cln/Pbt
3.C.01	31.6	9.7	s	Abt/Cln/Leu
3.C.16	27.9	11.8	t	Pbt
3.C.16 3.C.17	29.2	10.57	t	Cln
3.C.17 3.C.30	31.61	10.37	t	Bre
3.C.35	31.6	11.5	t	Pbt
	29.1	14.05	ND	Bre
3.C.64	24.2	10.75	t	Leu
3.D.21	23.2	11.03	S	Abt/Leu
3.D.24	26.1	11.65	S S	Abt/Cln/Leu/Pbt
3.D.35	23.4	12.26	S	Abt/Cln/Leu
3.D.40	24.45	12.20	t	Leu
3.D.44		12.62		Abt/Cln/Leu
3.D.60	27.2	14.2	S	Hn/Pbt
3.E.04	20.4		S	Hn/Tst
3.E.50	20.55	10.7	S	Cln/Leu
3.E.55	18.78	10.55	<u>s</u>	Cln/Hn
3.E.57	18.09	10.9	<u>s</u>	Abt/Tst
3.E.59	18.4	9.72	S	Leu
3.F.16	16.6	9.75	ND	Leu/Tst
3.F.2	16.73	9.35	S	Cln/Leu/Tst
3.F.50	16.25	11.6	S	Leu
3.F.72	16.9	13.7	t	Abt/Cln/Leu
3.F.82	13.8	13.12	S	
3.G.46	9.88	11.5	ND	Bre
3.G.78	10	12.93	ND_	Leu/Pbt
4.B.44	33.7	18.53	S	Cln/Hn
4.B.56	33.2	19.45	S	Bre/Leu
4.C.05	30	14.9	ND	Bre
4.C.25	28.62	17	ND	Bre Tst
4.C.42	NA	NA_	ND	
4.C.9	30.3	15.3	ND	Bre T/T-4
4.D.07	22.9	14.5	S	Leu/Tst
4.D.08	23.5	15	S	Abt/Tst
4.D.12	25	14.85	S	Abt/Leu/Tst
4.D.13	24.95	15.3	S	Abt/Bre
4.D.47	27.6	18.25	S	Abt/Leu/Pbt
4.E.53	19.39	18.43	t	Leu
4.F.15	13.25	15.45	t	Cln
4.F.17	14.1	15.6	S	Abt/Bre/Cln
4.F.22	17.56	16.2	S	Cln/Hn/Leu
4.F.6	14.85	14.59	ND	Bre
4.F.69	12.58	18.86	t	Abt
5.D.9	25.17	23.4	t	Hn

5

5.E.2	20.58	19.5	t	Bre
5.E.25	18.7	21.3	t	Cln
5.E.4	18.45	19.75	S	Abt/Bre/Leu
Y coordinat	e, X coordinat	e, fragment r	number	
<sup>2</sup> NA, spots to	oo close to ana	lyze.		
3	• • •	· · · · · · · · · · · · · · · · · · ·	1.11.	1 towast of mothylation

<sup>&</sup>lt;sup>3</sup>T, tumor-type specific target of methyaltion; s, shared target of methylation; ND, not determined.

# Example 2. Identification of diagnostic markers for lung cancer using AscI and RLGS

Tissue from lung tumors was obtained as surgical tissue samples. Where possible, surrounding non-tumor tissue from the same patient was obtained and used as a control. DNA was isolated from the tissue as described in Example 1. In preparation for RLGS analysis, the ends of the DNA were blocked as described in Example 1. The DNA was then digested with AscI followed by digestion with EcoRV. The AscI restriction enzyme recognizes the sequence 5'GGCGCGCC3' and does not cleave said sequence if cytosines within the sequence are methylated. First dimension gel electrophoresis, in-gel digestion with HinfI, second dimension gel electrophoresis and autoradiography were performed as described in Example 1.

RLGS profiles from lung tumor DNA were compared with RLGS profiles obtained from healthy, non-tumor tissue DNA. Spots which were lost or present at reduced intensity in tumor tissue RLGS profiles as compared to profiles obtained from healthy tissue were noted. Eight spots were lost or altered in the RLGS profiles from multiple lung tumor samples. A compilation of such spots is shown in Table II (lung tumors).

DNA sequence information was obtained from the lung cancer-specific spots. This was done by sequencing individual clones of an AscI/EcoRV library that was made from DNA from healthy tissue. Individual clones of this library that corresponded to spots on the AscI/EcoRV RLGS profile were identified by overloading an RLGS gel with DNA from various groups of library clones, as was described earlier in the specification of this application for the NotI/EcoRV library. After individual clones were matched with spots in the AscI/EcoRV profile, the DNA from the spots that were missing in profiles from the lung tumor DNAs were sequenced. Such sequence information is shown in the attached DNA Sequence Listing.

<sup>&</sup>lt;sup>4</sup>Types of tumor in which CpG island is methylated: Abt, gliomas; Bre, breast; Cln, colon; Hn, head and neck; Leu, acute myeloid leukemia; Pbt, pediatric brain tumors; Tst, testicular germ cell tumors.

Table II. Diagnostic CpG islands grouped by tumor type.

Library	Tumor type	Tumor type specific (+), shared (-), or not determined (ND) <sup>1</sup>	CpG island designation`
NotI/EcoRV	Breast	+	2.E.25, 2.F.2, 3.C.30, 5.E.2
		-	3.B.55, 4.B.56, 4.D.13, 4.F.17, 2.D.74, 2.C.35, 2.E.30, 5.E.4
		ND	2.D.25, 2.E.37, 2.F.59, 2.G.108, 3.C.64, 3.G.46, 4.C.05, 4.C.25, 4.C.9, 4.F.6
NotI/EcoRV	Colon	+	2.D.20, 3.C.17, 4.F.15, 5.E.25
		-	3.E.57, 4.B.44, 4.F.22, 2.D.55, 3.E.55, 3.F.50, 3.B.55, 4.F.17, 2.D.74, 2.C.35, 2.E.64, 3.C.01, 3.D.40, 3.D.60, 3.F.82, 3.B.36, 3.D.35
		ND	
NotI/EcoRV	Glioma	+	4.F.69
		-	4.D.13, 4.F.17, 2.D.74, 2.C.35, 2.E.30, 5.E.4, 2.E.64, 3.C.01, 3.D.40, 3.D.60, 3.F.82, 3.B.36, 3.D.35, 2.C.24, 2.C.58, 2.E.24, 2.F.50, 3.D.24, 4.D.47, 4.D.12, 2.E.61, 3.E.59, 4.D.08
		ND	
NotI/EcoRV	Head & neck	+	2.B.53, 5.D.9
		-	2.C.29, 2.C.54, 3.E.04, 3.E.50, 3.E.57, 4.B.44, 4.F.22
		ND	2.C.59, 2.D.27
NotI/EcoRV	Acute myelogenous Leukemia	+	2.D.14, 3.D.21, 3.D.44, 3.F.72, 4.E.53, 2.C.29, 2.C.54

10

		-	2.D.10, 2.D.34, 2.E.4,
			2.G.10, 3.F.2, 4.D.07,
			4.F.22, 2.D.55, 3.E.55,
			3.F.50, 2.E.64, 3.C.01,
			3.D.40, 3.D.60, 3.F.82,
			3.B.36, 3.D.35, 3.C.01,
			3.D.40, 3.D.60, 3.F.82,
			3.B.36, 3.D.35, 2.C.24,
			2.C.58, 2.E.24, 2.F.50,
ļ			3.D.24, 4.D.47, 4.D.12
		ND	2.D.48, 3.F.16, 3.G.78,
}			4.B.56
NotI/EcoRV	Pediatric	+	3.C.16, 3.C.35, 3.E.04
- 1	neuroectoder		
	mal tumor of		
	childhood		
		_	2.D.10, 2.D.34, 2.E.4,
			3.B.55, 2.C.35, 3.B.36,
			3.D.35, 4.D.47, 2.E.61
		ND	2.D.40, 2.E.20, 3.G.78
NotI/EcoRV	Testicular	+	2.F.41
		-	2.G.10, 3.F.2, 4.D.07,
			3.E.50, 3.F.50, 4.D.12,
			3.E.59, 4.D.08
		ND	2.C.57, 2.E.40, 3.B.30,
			4.C.42
AscI/EcoRV	Lung	+	
		-	
		ND	A.2.F.45, A.2.F.50,
			A.2.F.67, A.3.F.38,
			A.4.D.30, A.4.D.36,
			A.4.E.32, A.5.E.28 <sup>2</sup>

<sup>&</sup>lt;sup>1</sup>ND, not determined. Indicates that the designated CpG island was methylated in the indicated tumor type but its methylation in other tumor types was not determined.

### Example 3. Design of primers for cancer diagnosis

Primers are designed for diagnosis of cancer using methylation-specific PCR (MSR). The primers are designed to amplify regions of the human genome whose sequences are contained within the library clones disclosed in this application. Two sets of primers are needed for each library clone whose DNA sequence is to be used for diagnosis of cancer. Each primer

<sup>&</sup>lt;sup>2</sup>The "A" preceding the X, Y, number designation for the CpG islands indicates that these islands are from the AscI/EcoRV RLGS profile.

30

5

10

set is designed to amplify the same region of the genome, said region beginning at the end of a library clone containing the methylation-sensitive restriction enzyme recognition site (i.e., the NotI site for the library described in Example 1; the AscI site for the library described in

Example 2) and ending at a region contained within the clone up to 200 nucleotides from the

methylation-sensitive restriction enzyme recognition site.

The first set of primers is designed to amplify template genome DNA whose cytosine residues are not methylated and, after bisulfite treatment, the cytosines of said genome DNA are converted to uracil. The second set of primers is designed to amplify template genome DNA which is methylated on cytosines that comprise CpG dinucleotides. Such methylated cytosines are unaffected by bisulfite treatment. Therefore, by using two sets of primers, one set that will amplify only unmethylated DNA and another set that will amplify only methylated DNA, methylation state of the template DNA can be determined. Such methylation state can be diagnostic for cancer.

The primers used for MSR are designed to be from 15 to 34 nucleotides in length and contain within their sequence either CpG dinucleotides or dinucleotides complementary to CpG dinucleotides that have been treated with bisulfite. It is preferred that the 3' ends of primers used to amplify unmethylated DNA are CpA dinucleotides. It is preferred that the 3' ends of primers used to amplify methylated DNA are CpG dinucleotides.

For each library clone to be used diagnostically, the first set of primers are designed to amplify genome DNA that is not methylated. After treatment of such genome DNA with bisulfite, all such unmethylated cytosines are converted to uracil. PCR primers that will use such DNA as a template and amplify it, will have adenine residues which are complimentary to these uracils.

For the first set of primers, the 5' end of one of the primers begins at the end of the library clone containing the methylation-sensitive restriction enzyme recognition site. The sequence of this primer is identical in sequence to the strand of the template which has its 5' end as part of the methylation-sensitive restriction enzyme site, except that guanine residues are replaced with adenine residues. The adenines allow the primer to hybridize with the template strand in which cytosines have been converted to uracils by bisulfite. This primer extends to a length of between 15 and 32 total nucleotides. Preferably, the 3' end of said primer ends with a

CpA dinucleotide, the adenine of said dinucleotide hybridizing to a uracil which, before bisulfite treatment, had been a cytosine that comprised a CpG dinucleotide.

The diagram below shows implementation of these rules to select a primer that can be used to amplify clone 2.B.53 of the Notl/EcoRV library (Table I and attached sequence listing). In the diagram, I shows the end of the 2.B.53 clone containing the methylation-sensitive NotI site (NotI recognition sequence is shown in bold letters). CpG dinucleotides are shaded. To amplify a region of this clone rightward of the NotI site, the first primer is identical to the top strand of the duplex shown in I. However, since bisulfite treatment of the DNA in I converts cytosines to uracils, guanines within the PCR primer must be replaced with adenines. II shows the sequence of the bottom strand of I after bisulfite treatment converts cytosines to uracils. A primer complementary to the bisulfite-treated bottom strand has the sequence shown in III.



5

10

ב ייל

ngen green eigen gerig 181

[] \_\_25

30

III shows the entire sequence of one of the two primers used to amplify unmethylated This primer encompasses 5 CpG genome DNA corresponding to library clone 2.B.53. dinuceotides, as shown by the shading in I above. Encompassment of 2 or more such CpG dinucleotides is preferred so that this primer will not hybridize to a bisulfite-treated template which contains methylated cytosines. The 3'end of the primer shown in III ends in a CpA dinucleotide. This is also preferred in order to provide maximal discrimination of the primer between methylated and unmethylated template DNA in MSR. The primer shown in III has a length of 31 nucleotides.

The second primer is designed to work with the first primer in PCR amplification such that a fragment of less than about 200 base pairs is amplified. Therefore, this primer is made to a sequence rightward of the sequence shown in I. The sequence of this primer is complementary in sequence to the strand of the template which has its 5' end as part of the methylation-sensitive restriction enzyme site, except that guanine residues are replaced with adenine residues. This primer is preferably between 15 and 32 nucleotides in length. This primer is also designed to preferably encompass 2 or more CpG dinucleotides. Preferably, the 3' end of said primer ends with a CpA dinucleotide.

The diagram below shows implementation of these rules to select a primer that can be used to amplify unmethylated genome DNA corresponding to clone 2.B.53 of the Notl/EcoRV library. IV shows a region of the 2.B.53 clone about 70 nucleotides rightward of the sequence in I of the earlier diagram. The CpG dinucleotides within the sequence are shaded. To amplify a region leftward of this region, this second primer must be complementary to the top strand of the duplex shown in IV. However, bisulfite treatment of the DNA in IV converts cytosines to uracils. A primer complementary to this bisulfite-treated top strand has the sequence shown in VI.

```
IV
5'----GGAGTGGCGTCGCGGGAGGCTGCGCGCCACCGA----3'
3'----CCTCAGCGCCAGCGCCCTCCGACGGGGGGGGGGGTGGCT----5'

V
5'----GGAGTUGUGGTUGUGGGAGGUTGUGUUGUGUAUUGA----3'
VI
3' ACACCAACACCCTCCAACACACACATAACT 5'
```

5

10

<u>1</u> ≥ 25

30

VI shows the entire sequence of the second primer used to amplify unmethylated genome DNA corresponding to library clone 2.B.53. This primer encompasses 8 CpG dinucleotides, as shown by the shading in IV. Encompassment of 2 or more such CpG dinucleotides is preferred. The 3'end of the primer shown in VI ends in a CpA dinucleotide. This is also preferred. The primer shown in VI has a length of 31 nucleotides. Together, the first and second primers amplify a PCR fragment of 128 base pairs in length.

The above primers amplify genome DNA that does not contain 5-methylcytosine. The above primers will not amplify genome DNA containing 5-methylcytosines because 5-methylcytosines are not converted to uracils by bisulfite treatement. The two primers already described (III and VI), therefore, will not be complementary to bisulfite-treated genome DNA which is methylated.

30

5

10

Therefore, a second set of primers is designed to amplify genome DNA that is methylated. Methylation in human cells occurs at cytosines that are part of CpG residues. Such methylated cytosines are not converted to uracil by bisulfite treatment. Cytosines that are not part of CpG residues are not methylated and, therefore, are converted to uracil by bisulfite. The primers of the second set are designed to amplify the same region of a library clone as did the first set of primers. But, because the genome DNA contains both cytosines that are methylated and cytosines that are not methylated, the sequences of primers used to amplify such DNA are different than the sequences of the first primer set. Like the first set of primers, however, the primers of the second set are preferably between 15 and 32 nucleotides in length. Preferably the 3' ends of such primers contain CpG dinucleotides.

The diagram below shows implementation of these rules to select the first of two primers that can be used to amplify methylated genomic DNA corresponding to clone 2.B.53 of the NotI/EcoRV library. In the diagram below, VII shows the end of the 2.B.53 clone containing the NotI site (NotI recognition sequence is bolded). CpG dinucleotides are shaded. Cytosines within said CpG dinucleotides are methylated and are underlined in VII to indicate methylation to 5-methylcytosine. Treatment of the DNA in VII with bisulfite produces a bottom strand with the sequence shown in VIII. In VIII, only unmethylated cytosines are converted to uracil by bisulfite.

## 

#### VIII

3'UGCUGGCGCUAATUGAAGAGGAUAGGCTTGCGTUUU-----

#### IX

5'ACGACCGCGATTAACTTCTCCTATCCGAACG 3'

A primer complementary to the bisulfite-treated bottom strand shown in VIII is shown in IX. Said primer will prime PCR amplification of sequences rightward of those shown in VII. The primer shown in IX encompasses 5 CpG dinucleotides. Encompassment of 2 or more such CpG dinucleotides is preferred. The 3' end of the primer shown in IX ends in CpG. This is also preferred. The primer shown in IX has a length of 31 nucleotides.

A second primer is designed to work with the primer shown in IX to amplify methylated genome template DNA. Design of such a primer is shown below. In the diagram, X shows the same region of clone 2.B.53 (approximately 70 nucleotides rightward of the sequences shown in VII) that is shown in IV. Treatment of the DNA in X with bisulfite produces a top strand with the sequence shown in XI. In XI, only unmethylated cytosines are converted to uracil by bisulfite.

5

The man diese limb to the limb the man term to the limb t

į. į.

<u>2</u>25

30

A primer complementary to the bisulfite-treated top strand (XI) has the sequence shown in XII. Said primer will prime PCR amplification of sequences leftward of those shown in X. The primer shown in XII encompasses 8 CpG dinucleotides. Encompassment of 2 or more such CpG dinucleotides is preferred. The 3' end of the primer shown in XII ends in a CpG dinucleotide. This is also preferred. The primer shown in XII has a length of 31 nucleotides. Together, the first (IX) and second primers (XII) of the second set amplify a PCR fragment of 128 base pairs in length.

# Example 4. Use of oligonucleotides to diagnose cancer

The library clones, and DNA sequences within, can be used to detect DNA methylation in a genome at the specific sequences identified by the sequences within the clone. Such detection can be diagnostic for cancer. Various methods can be used for such diagnosis.

A. Diagnosis of cancer using methylation-sensitive restriction enzymes followed by Southern blot

Cleavage or lack of cleavage by a methylation-sensitive restriction enzyme at a specific restriction enzyme recognition site can be detected by a probe for the specific recognition site,

30

5

10

using Southern blotting. Genomic DNAs were isolated (as described in Example 1) from tumor tissue from a patient with acute myelogenous leukemia (AML). Cells from the same patient after chemotherapy and remission of the disease served as a source of control, healthy tissue DNA. The AML and control DNAs were designated as 26T and 26N, respectively. The DNAs were digested with NotI and EcoRV for 4 hours and then electrophoresed through a 0.8% agarose gel. DNA within the gel was depurinated by soaking the gel in 0.2 N HCl for 10 min. The gel was equlibrated in transfer solution (0.5 N NaOH, 1 M NaCl) for 10 min. and then blotted to Zeta Bind-GT nylon membranes (Bio-Rad). Blots were crosslinked with UV light, baked in a vacuum oven and then prehybridized for 1 hour at 65°C in a solution of 7% SDS, 500 mM sodium phosphate buffer (pH 7.2) and 1 mM EDTA. The blot was hybridized overnight at 65°C in prehybridization solution with 10 ng of  $\alpha^{-32}$ P-labeled probe at a specific activity of  $10^8$ - $10^9$ dpm/µg. The DNA probe used was the 2.C.40 clone from the NotI/EcoRV 2.C.40library. The purified NotI/EcoRV fragment (50 ng) was labeled with [α-32P]dCTP by random priming using the Prime-It II random-prime labeling kit (Stratagene). The blot was washed with two quick rinses at 65°C in wash solution 1 (100 mM sodium phosphate buffer, pH 7.2, 0.1% SDS), followed by one 30 min. wash at 65°C in wash solution 1. The blot was next washed for 30 min. at 65°C in wash solution 2 (40 mM sodium phosphate buffer, pH 7.2, 0.1% SDS). Bands were visualized by autoradiography using Kodak X-OMAT AR film.

Fig. 2B shows the data. The first 2 lanes of the autoradiograph are relevant. The first lane, labeled 26N is the normal, healthy tissue DNA cleaved with both NotI and EcoRV. The 26N lane shows a band near the bottom of the autoradiograph labeled "NotI/EcoRV." This is fragment resulting when the NotI site present in the 2.C.40 clone is unmethylated. The adjacent lane, labeled "26T," is the tumor tissue DNA cleaved with both NotI and EcoRV. It is seen that this band, labeled "EcoRV," does not migrate as fast as did the 26N band. The reason is that the NotI site present in the 2.C.40 clone is methylated and the NotI enzyme was unable to cleave at this site.

### B. Diagnosis of cancer using methylation-specific PCR (MSR)

MSR is a technique whereby DNA is amplified by PCR dependent upon the methylation state of the DNA. In this example, the specific areas of the genome whose methylation status is to be determined are the regions at the ends of the CpG islands that are demarcated by the

30

5

10

methylation-sensitive restriction enzyme recognition sequence. In the case of the NotI/EcoRV RLGS profiles, this is the NotI site. In the case of the AscI/EcoRV RLGS profiles, this is the AscI site, at the end of each clone.

For the purposes of this example, the methylation status of genomic sequences corresponding to the NotI site of clone 2.B.53 of the NotI/EcoRV library is examined. Genomic DNA is first isolated from normal tissue and from tumor tissue, as described in Example 1. This DNA is then treated with bisulfite. This is done by taking 1 µg of genomic DNA in a volume of 50 µl and denaturing said DNA in a final concentration of 0.2 M NaOH. Thirty microliters of 10 mM hydroquinone and 520 µl of 3 M sodium bisulfite, at pH 5.0, are added, mixed and incubated under mineral oil at 50°C for 16 hours. The modified DNA is then purified using the Wizard DNA purification resin (Promega) and eluted into 50 µl of water. Modification is completed by NaOH (final concentration, 0.3 M) treatment for 5 min. at room temperature, followed by ethanol precipitation. DNA is resuspended in water.

Each genomic DNA is then used in two PCR reactions. One PCR reaction will amplify DNA that is not methylated and has, therefore, been modified by bisulfite. The second PCR reaction will amplify DNA that is methylated. Separate primers are used for each reaction. To determine the methylation status of the NotI site in the genomic DNA which corresponds to the 2.B.53 clone, the two sets of primers described in Example 3 are used. Each PCR reaction contains 1X PCR buffer (16.6 mM ammonium sulfate, 67 mM Tris, pH 8.8, 6.7 mM MgCl<sub>2</sub>, 10 mM 2-mercaptoethanol), dNTPs (each at 1.25 mM), primers (300 ng each per reaction), and 50 ng bisulfite-modified DNA in a final volume of 50 µl. Separate control reactions are run which contain DNA that has not been modified by bisulfite. Reactions are hot-started at 95°C for 5 min. before the addition of 1.25 units of *Taq* polymerase. Amplification is carried out for 35 cycles (30 sec at 95°C, 30 sec at the annealing temperature, and 30 sec at 72°C), followed by a final 4 min. extension at 72°C. Each PCR reaction is directly loaded onto nondenaturing 6-8% polyacrylamide gels and electrophoresed. Gels are stained with ethidium bromide and visualized under UV illumination.

If input genomic DNA is not methylated at cytosines within CpG dinuceotides at the NotI site corresponding to the end of the 2.B.53 CpG island clone, the PCR reaction using the primers specific for nonmethylated DNA (primers III and VI in Example 3) will produce an amplification product of 128 base pairs in length. Using the same input genomic DNA, the PCR reaction using

5

10

the primers specific for methylated DNA (primers IX and XII in Example 3) will not produce an amplification product.

If input genomic DNA is methylated at cytosines within CpG dinucleotides at the NotI site corresponding to the end of the 2.B.53 CpG island clone, the PCR reaction using the nonmethylation-specific primers will not produce an amplification product. Using the same input genomic DNA, the PCR reaction using the methylation-specific primers will produce an amplification product of 128 base pairs in length.

### Example 5. Detection of gene expression

The library clones (Tables I and II) and DNA sequences (attached sequence listing) are useful for determining whether genes encoded within said clones are being transcribed in tumor tissue or cultured cells. To determine transcription, RNA was isolated from five different human glioma cell lines (U87MG, U178, T98G, U251 and LN235) using Trizol (Gibco BRL). Such RNA isolation reagent is known to those skilled in the art. RNAs were quantified using a spectrophotometer and then treated with amplification grade Dnase I (Gibco). The RNA (2 μg) was reverse transcribed by incubation with oligo-dT and random primers in a 20 μl reaction, heated to 70°C for 10 min. and placed on ice. A mix containing 1X reaction buffer (Gibco), DTT (10 mM), dNTPs (0.5 mM each), and RNAsin (80 U, Promega) was added to each sample. The samples were divided into two tubes, each containing 19 μl, and incubated at 37°C for 2 min. M-MLV reverse transcriptase (RT, 200 U) was added to one of the two tubes and each was incubated at 37°C for 1 h. DEPC-treated water (30 μl) was added to each sample and heated in boiling water for 5 min.

PCR amplification of the reverse transcribed RNA was then performed. In this study, transcripts encoded by sequences within the 2.C.24 library clone (Table I) were looked for. A computer search using the BLAST program had identified an open reading frame within the sequence of this library clone. PCR primers were made to this region. Primer 1 was 5' TGGTGCTGAAGTCGGTGAA 3'. Primer 2 was 5' GGGCCATCTTCACCATCTG 3'.

These primers (10 pmol of each) were used in 10  $\mu$ l PCR reactions which contained 1.5  $\mu$ l of the reverse transcription reaction, 1X reaction buffer, Taq polymerase (0.5 U, Boehringer),

10

and dNTPs (250  $\mu$ M each). For each gene, separate amplification reactions were carried out using RT-positive and RT-negative reactions as template. Amplification was not detected from the RT-negative reactions. The PCR reactions were carried out by heating the samples to 94°C for 5 min and then amplifying for 35 cycles, each cycle consisting of 94°C for 30 sec., a 30 sec. annealing step at 56°C, and 72°C for 45 sec. The reactions were then incubated at 72°C for 7 min and cooled to 4. The sample was then electrophoresed through an agarose gel containing ethidium bromide and PCR products were visualized using an Eagle Eye gel documentation system (Stratagene). The correct identity of the PCR products was confirmed by nucleotide sequencing of both strands.

The data showed that no transcripts encoded by this region of the 2.C.24 clone were found in any of the 5 glioma cell lines. Such expressed transcripts are present in RNA obtained from human fetal brain and adult brain.

In addition to examination of cell lines, tumor tissue obtained from patient samples can be similarly tested for the presence of transcripts by one skilled in the art. Other techniques to detect transcripts can also be used. Such techniques include, for example, Northern blot hybridization, RNase protection and primer extension assays.

30

5

10

### **CLAIMS**

What is claimed is:

- 1. A method of identifying CpG islands which are preferentially methylated in malignant cells contained within a tumor or neoplasm, comprising:
- a) digesting genomic DNA obtained from the malignant cells with an infrequentlycutting, methylation-sensitive, restriction enzyme to provide a set of malignant cell restriction fragments;
- b) digesting genomic DNA obtained from non-malignant, control cells with an infrequently-cutting, methylation-sensitive, restriction enzyme to provide a set of control cell restriction fragments;
- c) attaching a detectable label to the ends of the malignant cell restriction fragments and the control restriction fragments;
- d) digesting the labeled malignant cell and control cell restriction fragments with a second restriction enzyme;
- e) separating the labeled malignant cell restriction fragments and the labeled control cell restriction fragments, wherein the malignant cell restriction fragments and the control cell restriction fragments are separated by electrophoresis on two different gels;
- f) digesting the restriction fragments in each of said gels with a third restriction enzyme;
- g) electrophoresing the restriction fragments in each of said gels in a direction perpendicular to the first direction to provide a first pattern of detectable malignant cell restriction fragments and a second pattern of detectable control cell restriction fragments; and
- h) comparing the first pattern to the second pattern to identify diagnostic control cell restriction fragments in said second pattern which are absent or exhibit a decreased intensity in the first pattern, wherein said diagnostic control cell restriction fragments comprise a CpG island that is unmethylated in the DNA of the control cells and methylated in the DNA of the malignant cells.
- 2. The method of claim 1 further comprising the step of determining the sequence of at least a portion of a diagnostic control cell restriction fragment, wherein said portion is located at or near an end of the fragment.
- 3. The method of claim 1 further comprising the step of obtaining a clone from a DNA

30

5

library which comprises a diagnostic control cell restriction fragment.

4. A method of preparing a polynucleotide or oligonucleotide for characterizing tissue obtained from a subject suspected of having cancer, comprising:

synthesizing a polynucleotide or oligonucleotide which comprises a sequence which is identical to or substantially complementary to a target sequence on one of the strands of a diagnostic control fragment identified according to the method of claim 2, wherein said target sequence comprises at least two CpG dinucleotides, wherein said oligonucleotide is from 15 to 34 nucleotides in length, and wherein said polynucleotide is from 35 to 2000 nucleotides in length.

- 10 5. The method of claim 4 wherein said target sequence is located at or near the control restriction fragment end which was cleaved by the methylation-sensitive, restriction enzyme.
  - 6. The method of claim 4 wherein the target sequence is located from about 100 nucleotides to about 500 nucleotides downstream of the control restriction fragment end that was cleaved by the methylation-sensitive, restriction enzyme.
  - The method of claim 4 wherein the control restriction fragment comprises a sequence 7. selected from the group consisting of SEQ. ID. NO.:1, SEQ. ID. NO.:2, SEQ. ID. NO.:3, SEQ. ID. NO.:4, SEQ. ID. NO.:5, SEQ. ID. NO.:6, SEQ. ID. NO.:7, SEQ. ID. NO.:8, SEQ. ID. NO.:9, SEQ. ID. NO.:10, SEQ. ID. NO:11, SEQ. ID. NO.: 12, SEQ. ID. NO.:13, SEQ. ID. NO.:14, SEQ. ID. NO.:15, SEQ. ID. NO.: 16, SEQ. ID. NO.:17, SEQ. ID. NO.:18, SEQ. ID. NO.: 19, SEQ. ID. NO.:20, SEQ. ID. NO.:21, SEQ. ID. NO.:22, SEQ. ID. NO.:23, SEQ. ID. NO.:24, SEQ. ID. NO.:25, SEQ. ID. NO.:26, SEQ. ID. NO.:27, SEQ. ID. NO.:28, SEQ. ID. NO.:29, SEQ. ID. NO.:30, SEQ. ID. NO.:31, SEQ. ID. NO.:32, SEQ. ID. NO.:33, SEQ. ID. NO.:34, SEQ. ID. NO.:35, SEQ. ID. NO.:36, SEQ. ID. NO.:37, SEQ. ID. NO.:38, SEQ. ID. NO.:39, SEQ. ID. NO.:40, SEQ. ID. NO.:41, SEQ. ID. NO.:42, SEQ. ID. NO.:43, SEQ. ID. NO.:44, SEQ. ID. NO.:45, SEQ. ID. NO.:46, SEQ. ID. NO.:47, SEQ. ID. NO.:48, SEQ. ID. NO.:49, SEQ. ID. NO.:50, SEQ. ID. NO.:51, SEQ. ID. NO.:52, SEQ. ID. NO.:53, SEQ. ID. NO.:54, SEQ. ID. NO.:55, SEQ. ID. NO.:56, SEQ. ID. NO.:57, SEQ. ID. NO.:58, SEQ. ID. NO.:59, SEQ. ID. NO.:60, SEQ. ID. NO.:61, SEQ. ID. NO.:62, , SEQ. ID. NO.:63, SEQ. ID. NO.:64, SEQ. ID. NO.:65, SEQ. ID. NO.:66, SEQ. ID. NO.:67, SEQ. ID. NO.:68, SEQ. ID. NO.:69, SEQ. ID. NO.:70, SEQ. ID. NO.:71, SEQ. ID. NO.:72, SEQ. ID. NO.:73, SEQ. ID. NO.:74, SEQ. ID. NO.:75, SEQ. ID. NO.:76, SEQ. ID. NO.:77, SEQ. ID. NO.:78, SEQ. ID. NO.:79,

30

5

10

SEQ. ID. NO.:80, SEQ. ID. NO.:81, SEQ. ID. NO.:82, SEQ ID NO: 83, SEQ ID NO. 84, SEQ ID. NO. 85, SEQ. ID. NO. 86, SEQ. ID. NO. 87, SEQ. ID. NO. 88, SEQ. ID. NO. 89, SEQ. ID. NO. 90, SEQ. ID. NO. 91, SEQ ID. NO. 92, and SEQ. ID. NO. 93.

- 8. An isolated polynucleotide or oligonucleotide for characterizing cells that are obtained from a subject suspected of having a cancer which is associated with methylation of one or a plurality of CpG islands in the genomic DNA of malignant cells, wherein said polynucleotide or oligonucleotide comprises a sequence which is identical to or complementary to a target sequence on one of the strands of a diagnostic control fragment identified according to the method of claim 2, wherein said target sequence comprises at least two CpG dinucleotides, wherein said oligonucleotide is from 15 to 34 nucleotides in length; and wherein said polynucleotide is from 35 to 3000 nucleotides in length.
- 9. The isolated polynucleotide or oligonucleotide of claim 8 wherein said target sequence is located at or near the control restriction fragment end which was cleaved by the methylation sensitive restriction enzyme.
- 10. The isolated polynucleotide or oligonucleotide of claim 8 wherein the target sequence is located from about 100 nucleotides to about 500 nucleotides downstream of the control restriction fragment end that was cleaved by the methylation-sensitive, restriction enzyme.
- 11. An isolated polynucleotide or oligonucleotide for characterizing cells which are obtained from a subject suspected of having a cancer which is associated with methylation of one or a plurality of CpG islands in the genomic DNA of malignant cells, wherein said polynucleotide or oligonucleotide comprises a sequence which is identical to or complementary to a modified target sequence on one of the strands of a diagnostic control fragment identified according to the method of claim 2, wherein said modified target sequence is derived from a target sequence that has been modified by treatment with sodium bisulfite, wherein said modified target sequence lacks cytosines and comprises at least two UpG dinucleotides, wherein said oligonucleotide is from 15 to 34 nucleotides in length; and wherein said polynucleotide is from 35 to 3000 nucleotides in length.
- 12. The isolated polynucleotide or oligonucleotide of claim 11 wherein the modified target sequence is derived from a target sequence located at or near the control restriction fragment end that was cleaved by the methylation sensitive restriction enzyme.
- 13. The isolated polynucleotide or oligonucleotide of claim11 wherein the modifed target

30

15.

5

10

sequence is derived from a target sequence that is located from about 100 nucleotides to about 500 nucleotides downstream of the control restriction fragment end that was cleaved by the methylation-sensitive restriction enzyme.

An isolated polynucleotide for characterizing cells which are obtained from a subject 14. suspected of having a cancer selected from the group consisting of glioma, acute myeloid leukemia, primitive neuroectodermal tumors of childhood, breast cancer, colon cancer, head and neck cancer, testiclular cancer and lung cancer; wherein said polynucleotide is from 35 to 3000 nucleotides in length and comprises at least two CpG dinucleotides, and wherein said polynucleotide comprise a sequence which is identical to or complementary to a target sequence located within a sequence selected from the group consisting of SEQ. ID. NO.:1, SEQ. ID. NO.:2, SEQ. ID. NO.:3, SEQ. ID. NO.:4, SEQ. ID. NO.:5, SEQ. ID. NO.:6, SEQ. ID. NO.:7, SEQ. ID. NO.:8, SEQ. ID. NO.:9, SEQ. ID. NO.:10, SEQ. ID. NO:11, SEQ. ID. NO.: 12, SEQ. ID. NO.:13, SEQ. ID. NO.:14, SEQ. ID. NO.:15, SEQ. ID. NO.: 16, SEQ. ID. NO.:17, SEQ. ID. NO.:18, SEQ. ID. NO.: 19, SEQ. ID. NO.:20, SEQ. ID. NO.:21, SEQ. ID. NO.:22, SEQ. ID. NO.:23, SEQ. ID. NO.:24, SEQ. ID. NO.:25, SEQ. ID. NO.:26, SEQ. ID. NO.:27, SEQ. ID. NO.:28, SEQ. ID. NO.:29, SEQ. ID. NO.:30, SEQ. ID. NO.:31, SEQ. ID. NO.:32, SEQ. ID. NO.:33, SEQ. ID. NO.:34, SEQ. ID. NO.:35, SEQ. ID. NO.:36, SEQ. ID. NO.:37, SEQ. ID. NO.:38, SEQ. ID. NO.:39, SEQ. ID. NO.:40, SEQ. ID. NO.:41, SEQ. ID. NO.:42, SEQ. ID. NO.:43, SEQ. ID. NO.:44, SEQ. ID. NO.:45, SEQ. ID. NO.:46, SEQ. ID. NO.:47, SEQ. ID. NO.:48, SEQ. ID. NO.:49, SEQ. ID. NO.:50, SEQ. ID. NO.:51, SEQ. ID. NO.:52, SEQ. ID. NO.:53, SEQ. ID. NO.:54, SEQ. ID. NO.:55, SEQ. ID. NO.:56, SEQ. ID. NO.:57, SEQ. ID. NO.:58, SEQ. ID. NO.:59, SEQ. ID. NO.:60, SEQ. ID. NO.:61, SEQ. ID. NO.:62, , SEQ. ID. NO.:63, SEQ. ID. NO.:64, SEQ. ID. NO.:65, SEQ. ID. NO.:66, SEQ. ID. NO.:67, SEQ. ID. NO.:68, SEQ. ID. NO.:69, SEQ. ID. NO.:70, SEQ. ID. NO.:71, SEQ. ID. NO.:72, SEQ. ID. NO.:73, SEQ. ID. NO.:74, SEQ. ID. NO.:75, SEQ. ID. NO.:76, SEQ. ID. NO.:77, SEQ. ID. NO.:78, SEQ. ID. NO.:79, SEQ. ID. NO.:80, SEQ. ID. NO.:81, SEQ. ID. NO.:82, SEQ ID NO: 83, SEQ ID NO. 84, SEQ ID. NO. 85, SEQ. ID. NO. 86, SEQ. ID. NO. 87, SEQ. ID. NO. 88, SEQ. ID. NO. 89, SEQ. ID. NO. 90, SEQ. ID. NO. 91, SEQ ID. NO. 92, and SEQ. ID. NO. 93.

glioma and said polynucleotide comprises a sequence which is identical to or complementary to

The isolated polynucleotide of claim 14 wherein said subject is suspected of having

30

5

10

a target sequence located within SEQ. ID.NO. 78, SEQ ID NO:71, SEQ ID NO:75, SEQ ID NO:18, SEQ ID NO:44, SEQ ID NO:22, SEQ ID NO:82, SEQ ID NO:27, SEQ ID NO:38, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:59, SEQ ID NO:36, SEQ ID NO:46, SEQ ID NO:2, SEQ ID NO:7, SEQ ID NO:20, SEQ ID NO:30, SEQ ID NO:45, SEQ ID NO:72, SEQ ID NO:70, SEQ ID NO:26, SEQ ID NO:54, or SEQ ID NO:69.

- The isolated polynucleotide of claim 14 wherein said subject is suspected of having acute myeloid leukemia and said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located within SEQ ID NO:10, SEQ ID NO:44, SEQ ID NO:48, SEQ ID NO:58, SEQ ID NO:73, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:9, SEQ ID NO:14, SEQ ID NO:24, SEQ ID NO:33, SEQ ID NO:56, SEQ ID NO:68, SEQ ID NO:76, SEQ ID NO:17, SEQ ID NO:52, SEQ ID NO:57, SEQ ID NO:26, SEQ ID NO:38, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:59, SEQ ID NO:36, SEQ ID NO:46, SEQ ID NO:38, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:59, SEQ ID NO:36, SEQ ID NO:46, SEQ ID NO:2, SEQ ID NO:7, SEQ ID NO:20, SEQ ID NO:30, SEQ ID NO:45, SEQ ID NO:72, SEQ ID NO:16, SEQ ID NO:55, SEQ ID NO:61, SEQ ID NO:63, or SEQ ID NO:70.
- The isolated polynucleotide of claim 14 wherein said subject is suspected of having a primitive neuroectodermal tumor of childhood and said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located within SEQ ID NO:39, SEQ ID NO:42, SEQ ID NO:50, SEQ ID NO:9, SEQ ID NO:14, SEQ ID NO:24, SEQ ID NO:37, SEQ ID NO:4, SEQ ID NO:36, SEQ ID NO:46, SEQ ID NO:72, SEQ ID NO:26, SEQ ID NO:15, SEQ ID NO:19, or SEQ ID NO:61.
- The isolated polynucleotide of claim 14 wherein said subject is suspected of having breast cancer and said polynucleotide comprises a sequence identical or complementary to a target sequence which is located within SEQ ID NO:21, SEQ ID NO:28, SEQ ID NO:41, SEQ ID NO:80, SEQ ID NO:37, SEQ ID NO:63, SEQ ID NO:71, SEQ ID NO:75, SEQ ID NO:18, SEQ ID NO:4, SEQ ID NO:22, SEQ ID NO:82, SEQ ID NO:12, SEQ ID NO:23, SEQ ID NO:31, SEQ ID NO:34, SEQ ID NO:43, SEQ ID NO:60, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:77.
- 19. The isolated polynucleotide of claim 14 wherein said subject is suspected of having colon cancer and said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located within SEQ ID NO:11, SEQ ID NO:40, SEQ ID NO:74, SEQ ID NO:81,

30

5

SEQ ID NO:53, SEQ ID NO:62, SEQ ID NO:76, SEQ ID NO:17, SEQ ID NO:52, SEQ ID NO:57, SEQ ID NO:37, SEQ ID NO:75, SEQ ID NO:18, SEQ ID NO:4, SEQ ID NO:27, SEQ ID NO:38, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:59, SEQ ID NO:36, or SEQ ID NO:46.

- 20. The isolated polynucleotide of claim 14 wherein said subject is suspected of head and neck cancer and said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located within in SEQ ID NO:1, SEQ ID NO. 79, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:62, SEQ ID NO:76, SEQ ID NO:8, or SEQ ID NO:13.
- 21. The isolated polynucleotide of claim 14 wherein said subject is suspected of testiclular cancer and said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located within in SEQ ID NO. 29, SEQ ID NO:33, SEQ ID NO:56, SEQ ID NO:68, SEQ ID NO:51, SEQ ID NO:57, SEQ ID NO:70, SEQ ID NO:54, or SEQ ID NO:69
  - 22. The isolated polynucleotide of claim 14 wherein said subject is suspected of having a lung cancer and said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located within SEQ ID NO: 83, SEQ ID NO. 84, SEQ ID. NO. 85, SEQ. ID. NO. 86, SEQ. ID. NO. 87, SEQ. ID. NO. 88, SEQ. ID. NO. 89, SEQ. ID. NO. 90, SEQ. ID. NO. 91, SEQ ID. NO. 92, and SEQ. ID. NO. 93.
  - An isolated CpG diagnostic oligonucleotide for characterizing cells which are obtained from a subject suspected of having a cancer selected from the group consisting of glioma, acute myeloid leukemia, primitive neuroectodermal tumors of childhood, breast cancer, colon cancer, head and neck cancer, testicular cancer and lung cancer; wherein said oligonucleotide is from 15 to 34 nucleotides in length and comprises at least two CpG dinucleotides, and wherein said oligonucleotide comprise a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 of a sequence selected from the group consisting of SEQ. ID. NO.:1, SEQ. ID. NO.:2, SEQ. ID. NO.:3, SEQ. ID. NO.:4, SEQ. ID. NO.:5, SEQ. ID. NO.:6, SEQ. ID. NO.:7, SEQ. ID. NO.:8, SEQ. ID. NO.:9, SEQ. ID. NO.:10, SEQ. ID. NO.:11, SEQ. ID. NO.:12, SEQ. ID. NO.:13, SEQ. ID. NO.:14, SEQ. ID. NO.:15, SEQ. ID. NO.:16, SEQ. ID. NO.:17, SEQ. ID. NO.:18, SEQ. ID. NO.:19, SEQ. ID. NO.:20, SEQ. ID. NO.:21, SEQ. ID. NO.:22, SEQ. ID. NO.:23, SEQ. ID. NO.:24, SEQ. ID. NO.:25, SEQ. ID. NO.:26, SEQ. ID. NO.:27, SEQ. ID. NO.:28, SEQ. ID. NO.:29, SEQ. ID. NO.:25, SEQ. ID. NO.:26, SEQ. ID. NO.:27, SEQ. ID. NO.:28, SEQ. ID. NO.:29, SEQ. ID. NO.:30,

30

5

SEQ. ID. NO.:31, SEQ. ID. NO.:32, SEQ. ID. NO.:33, SEQ. ID. NO.:34, SEQ. ID. NO.:35, SEQ. ID. NO.:36, SEQ. ID. NO.:37, SEQ. ID. NO.:38, SEQ. ID. NO.:39, SEQ. ID. NO.:40, SEQ. ID. NO.:41, SEQ. ID. NO.:42, SEQ. ID. NO.:43, SEQ. ID. NO.:44, SEQ. ID. NO.:45, SEQ. ID. NO.:46, SEQ. ID. NO.:47, SEQ. ID. NO.:48, SEQ. ID. NO.:49, SEQ. ID. NO.:50, SEQ. ID. NO.:51, SEQ. ID. NO.:52, SEQ. ID. NO.:53, SEQ. ID. NO.:54, SEQ. ID. NO.:55, SEQ. ID. NO.:56, SEQ. ID. NO.:57, SEQ. ID. NO.:58, SEQ. ID. NO.:59, SEQ. ID. NO.:60, SEQ. ID. NO.:61, SEQ. ID. NO.:62, , SEQ. ID. NO.:63, SEQ. ID. NO.:64, SEQ. ID. NO.:65, SEQ. ID. NO.:66, SEQ. ID. NO.:67, SEQ. ID. NO.:68, SEQ. ID. NO.:69, SEQ. ID. NO.:70, SEQ. ID. NO.:71, SEQ. ID. NO.:72, SEQ. ID. NO.:73, SEQ. ID. NO.:74, SEQ. ID. NO.:75, SEQ. ID. NO.:76, SEQ. ID. NO.:77, SEQ. ID. NO.:78, SEQ. ID. NO.:79, SEQ. ID. NO.:80, 10 SEQ. ID. NO.:81, SEQ. ID. NO.:82; SEQ ID NO: 83, SEQ ID NO. 84, SEQ ID. NO. 85, SEQ. ID. NO. 86, SEQ. ID. NO. 87, SEQ. ID. NO. 88, SEQ. ID. NO. 89, SEQ. ID. NO. 90, SEQ. ID. NO. 91, SEQ ID. NO. 92, and SEQ. ID. NO. 93; or a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.

- The isolated oligonucleotide of claim 23 wherein said subject is suspected of having 24. glioma and said oligonucleotide comprises a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 in SEQ ID 78, SEQ ID NO:71, SEQ ID NO:75, SEQ ID NO:18, SEQ ID NO:4, SEQ ID NO:22, SEQ ID NO:82, SEQ ID NO:27, SEQ ID NO:38, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:59, SEQ ID NO:36, SEQ ID NO:46, SEQ ID NO:2, SEQ ID NO:7, SEQ ID NO:20, SEQ ID NO:30, SEQ ID NO:45, SEQ ID NO:72, SEQ ID NO:70, SEQ ID NO:26, SEQ ID NO:54, or SEQ ID NO:69, or said oligonucleotide comprises a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.
- The isolated oligonucleotide of claim 23 wherein the subject is suspected of having acute 25. myeloid leukemia; and said oligonucleotide comprises a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 SEQ ID NO:10, SEQ ID NO:44, SEQ ID NO:48, SEQ ID NO:58, SEQ ID NO:73, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:9, SEQ ID NO:14, SEQ ID NO:24, SEQ ID NO:33, SEQ ID NO:56, SEQ ID NO:68, SEQ ID NO:76, SEQ ID NO:17, SEQ ID NO:52, SEQ ID NO:57, SEQ ID NO:26,

30

5

10

SEQ ID NO:38, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:59, SEQ ID NO:36, SEQ ID NO:46, SEQ ID NO:38, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:59, SEQ ID NO:36, SEQ ID NO:46, SEQ ID NO:2, SEQ ID NO:7, SEQ ID NO:20, SEQ ID NO:30, SEQ ID NO:45, SEQ ID NO:72, SEQ ID NO:16, SEQ ID NO:55, SEQ ID NO:61, SEQ ID NO:63, or SEQ ID NO:70, or said oligonucleotide comprises a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.

- 26. The isolated oligonucleotide of claim 23 wherein said subject is suspected of having primitive neuroectodermal tumors of childhood; and said oligonucleotide comprises a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 of SEQ ID NO:39, SEQ ID NO:42, SEQ ID NO:50, SEQ ID NO:9, SEQ ID NO:14, SEQ ID NO:24, SEQ ID NO:37, SEQ ID NO:4, SEQ ID NO:36, SEQ ID NO:46, SEQ ID NO:72, SEQ ID NO:26, SEQ ID NO:15, SEQ ID NO:19, or SEQ ID NO:61, or said nucleotide comprises a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.
- 27. The isolated oligonucleotide of claim 23 wherein said subject is suspected of having breast cancer; and said oligonucleotide comprises a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 of SEQ ID NO:21, SEQ ID NO:28, SEQ ID NO:41, SEQ ID NO:80, SEQ ID NO:37, SEQ ID NO:63, SEQ ID NO:71, SEQ ID NO:75, SEQ ID NO:18, SEQ ID NO:4, SEQ ID NO:22, SEQ ID NO:82, SEQ ID NO:12, SEQ ID NO:23, SEQ ID NO:31, SEQ ID NO:34, SEQ ID NO:43, SEQ ID NO:60, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:77, or said oligonucleotide comprises a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.
- The isolated oligonucleotide of claim 23 wherein said subject is suspected of having colon cancer; and said oligonucleotide comprises a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 of SEQ ID NO:11, SEQ ID NO:40, SEQ ID NO:74, SEQ ID NO:81, SEQ ID NO:53, SEQ ID NO:62, SEQ ID NO:76, SEQ ID NO:17, SEQ ID NO:52, SEQ ID NO:57, SEQ ID NO:37, SEQ ID NO:75, SEQ ID NO:4, SEQ ID NO:4, SEQ ID NO:27, SEQ ID NO:38, SEQ ID NO:47, SEQ ID N

30

5

10

NO:49, SEQ ID NO:59, SEQ ID NO:36, SEQ ID NO:46 or said oligonucleotide comprises a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.

- 29. The isolated oligonucleotide of claim 23 wherein said subject is suspected of having head and neck cancer; and said oligonucleotide comprises a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 in SEQ ID NO:1, SEQ ID NO.79, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:62, SEQ ID NO:76, SEQ ID NO:8, or SEQ ID NO:13., or said nucleotide a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.
- 30. The isolated oligonucleotide of claim 23 wherein said subject is suspected of having testiclular cancer; and said oligonucleotide comprises a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 in SEQ ID NO. 29, SEQ ID NO:33, SEQ ID NO:56, SEQ ID NO:68, SEQ ID NO:51, SEQ ID NO:57, SEQ ID NO:70, SEQ ID NO:54, or SEQ ID NO:69, or said oligonucleotide comprises a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.
- The isolated oligonucleotide of claim 23 wherein said subject is suspected of having lung cancer; and said oligonucleotide comprises a sequence which is identical to a target sequence located within a region extending from nucleotide 1 through nucleotide 99 of SEQ ID NO SEQ ID NO: 83, SEQ ID NO. 84, SEQ ID. NO. 85, SEQ. ID. NO. 86, SEQ. ID. NO. 87, SEQ. ID. NO. 88, SEQ. ID. NO. 89, SEQ. ID. NO. 90, SEQ. ID. NO. 91, SEQ ID. NO. 92, and SEQ. ID. NO. 93, or said oligonucleotide comprises a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.
- 32. A method for determining whether cells obtained from a subject suspected of having a cancer are malignant or non-malignant, comprising:
- a) digesting DNA which has been isolated from the cells with a methylationsensitive restriction enzyme to provide a set of restriction fragments;
- b) hybridizing said restriction fragments with a CpG diagnostic polynucleotide comprising a sequence which is identical to or complementary to a target sequence on one of the

5

10

strands of a diagnostic control fragment identified according to the method of claim 2, wherein said method employed said methylation-sensitive restriction enzyme, wherein said target sequence comprises at least two CpG dinucleotides, wherein said polynucleotide is from 35 to 3000 nucleotides in length, and wherein said reaction is conducted under stringent hybridization conditions; and

- c) assaying the reaction products of step b to determine the size or the sequence of the restriction fragment to which the CpG diagnostic polynucleotide has hybridized.
- 33. The method of claim 32 wherein said subject is suspected of having a cancer selected from the group consisting of glioma, acute myeloid leukemia, primitive neuroectodermal tumors of childhood, breast cancer, colon cancer, head and neck cancer, and testiclular cancer;

wherein said DNA sample is digested with NotI; and wherein said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located in SEQ. ID. NO.:1, SEQ. ID. NO.:2, SEQ. ID. NO.:3, SEQ. ID. NO.:4, SEQ. ID. NO.:5, SEQ. ID. NO.:6, SEQ. ID. NO.:7, SEQ. ID. NO.:8, SEQ. ID. NO.:9, SEQ. ID. NO.:10, SEQ. ID. NO:11, SEQ. ID. NO.: 12, SEQ. ID. NO.:13, SEQ. ID. NO.:14, SEQ. ID. NO.:15, SEQ. ID. NO.: 16, SEQ. ID. NO.:17, SEQ. ID. NO.:18, SEQ. ID. NO.: 19, SEQ. ID. NO.:20, SEQ. ID. NO.:21, SEQ. ID. NO.:22, SEQ. ID. NO.:23, SEQ. ID. NO.:24, SEQ. ID. NO.:25, SEQ. ID. NO.:26, SEQ. ID. NO.:27, SEQ. ID. NO.:28, SEQ. ID. NO.:29, SEQ. ID. NO.:30, SEQ. ID. NO.:31, SEQ. ID. NO.:32, SEQ. ID. NO.:33, SEQ. ID. NO.:34, SEQ. ID. NO.:35, SEQ. ID. NO.:36, SEQ. ID. NO.:37, SEQ. ID. NO.:38, SEQ. ID. NO.:39, SEQ. ID. NO.:40, SEQ. ID. NO.:41, SEQ. ID. NO.:42, SEQ. ID. NO.:43, SEQ. ID. NO.:44, SEQ. ID. NO.:45, SEQ. ID. NO.:46, SEQ. ID. NO.:47, SEQ. ID. NO.:48, SEQ. ID. NO.:49, SEQ. ID. NO.:50, SEQ. ID. NO.:51, SEQ. ID. NO.:52, SEQ. ID. NO.:53, SEQ. ID. NO.:54, SEQ. ID. NO.:55, SEQ. ID. NO.:56, SEQ. ID. NO.:57, SEQ. ID. NO.:58, SEQ. ID. NO.:59, SEQ. ID. NO.:60, SEQ. ID. NO.:61, SEQ. ID. NO.:62, , SEQ. ID. NO.:63, SEQ. ID. NO.:64, SEQ. ID. NO.:65, SEQ. ID. NO.:66, SEQ. ID. NO.:67, SEQ. ID. NO.:68, SEQ. ID. NO.:69, SEQ. ID. NO.:70, SEQ. ID. NO.:71, SEQ. ID. NO.:72, SEQ. ID. NO.:73, SEQ. ID. NO.:74, SEQ. ID. NO.:75, SEQ. ID. NO.:76, SEQ. ID. NO.:77, SEQ. ID. NO.:78, SEQ. ID. NO.:79, SEQ. ID. NO.:80, SEQ. ID. NO.:81, and SEQ. ID. NO.:82.

30 34. The method of claim 32 wherein the subject is suspected of having lung cancer; wherein the DNA is digested with AscI;

30

and

10

and wherein said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located in , SEQ ID NO: 83, SEQ ID NO. 84, SEQ ID. NO. 85, SEQ. ID. NO. 86, SEQ. ID. NO. 87, SEQ. ID. NO. 88, SEQ. ID. NO. 89, SEQ. ID. NO. 90, SEQ. ID. NO. 91, SEQ ID. NO. 92, and SEQ. ID. NO. 93.

- 5 35. A method of determining whether cells contained within a tissue sample obtained from a subject suspected of having cancer are malignant, comprising:
  - a) treating DNA isolated from the tissue sample with a compound which converts non-methylated cytosines to a different nucleotide base;
  - b) reacting a portion of the treated DNA with a CpG diagnostic oligonucleotide which is complementary to a target sequence which comprises CpG islands that are preferentially methylated in malignant cells of subjects known to have said cancer;
  - c) reacting a portion of the treated DNA with a modified CpG diagnostic oligonucleotide which is complementary to a modified target sequence in which the cytosines in said target sequence are replaced with the different nucleotide base; and
  - d) assaying the reaction products of step b and step c to determine whether the treated DNA has hybridized with the CpG diagnostic oligonucleotide or the modified CpG diagnostic oligonucleotide; wherein hybridization of the treated DNA with the CpG diagnostic oligonucleotide as opposed to the modified CpG diagnostic oligonucleotide indicates that the DNA has been obtained from malignant cells.
  - 36. The method of claim 35 wherein the chemical compound is sodium bisulfite and the non-methylated cytosines are converted to uracil.
  - 37. The method of claim 35 wherein the assay is a polymerase chain reaction,

wherein a portion of the treated DNA is reacted with a first primer set which comprises two diagnostic CpG olignonucleotides; and

- wherein a portion of the treated DNA is reacted with a second primer set which comprises two modified diagnostic CpG oligonucleotides.
- 38. The method of claim 35 wherein the subject is suspected of having a cancer selected from the group consisting of glioma, acute myeloid leukemia, primitive neuroectodermal tumors of childhood, breast cancer, colon cancer, head and neck cancer, testiclular cancer and lung cancer;
- wherein the CpG diagnostic oligonucleotide comprises a sequence which is identical to a target

30

sequence located between nucleotide 1 and 100 in a sequece selected from the group consisting said polynucleotide comprises a sequence which is identical to or complementary to a target sequence located in SEQ. ID. NO.:1, SEQ. ID. NO.:2, SEQ. ID. NO.:3, SEQ. ID. NO.:4, SEQ. ID. NO.:5, SEQ. ID. NO.:6, SEQ. ID. NO.:7, SEQ. ID. NO.:8, SEQ. ID. NO.:9, SEQ. ID. NO.:10, SEQ. ID. NO:11, SEQ. ID. NO.: 12, SEQ. ID. NO.:13, SEQ. ID. NO.:14, SEQ. ID. 5 NO.:15, SEQ. ID. NO.: 16, SEQ. ID. NO.:17, SEQ. ID. NO.:18, SEQ. ID. NO.: 19, SEQ. ID. NO.:20, SEQ. ID. NO.:21, SEQ. ID. NO.:22, SEQ. ID. NO.:23, SEQ. ID. NO.:24, SEQ. ID. NO.:25, SEQ. ID. NO.:26, SEQ. ID. NO.:27, SEQ. ID. NO.:28, SEQ. ID. NO.:29, SEQ. ID. NO.:30, SEQ. ID. NO.:31, SEQ. ID. NO.:32, SEQ. ID. NO.:33, SEQ. ID. NO.:34, SEQ. ID. NO.:35, SEQ. ID. NO.:36, SEQ. ID. NO.:37, SEQ. ID. NO.:38, SEQ. ID. NO.:39, SEQ. ID. 10 NO.:40, SEQ. ID. NO.:41, SEQ. ID. NO.:42, SEQ. ID. NO.:43, SEQ. ID. NO.:44, SEQ. ID. NO.:45, SEQ. ID. NO.:46, SEQ. ID. NO.:47, SEQ. ID. NO.:48, SEQ. ID. NO.:49, SEQ. ID. NO.:50, SEQ. ID. NO.:51, SEQ. ID. NO.:52, SEQ. ID. NO.:53, SEQ. ID. NO.:54, SEQ. ID. NO.:55, SEQ. ID. NO.:56, SEQ. ID. NO.:57, SEQ. ID. NO.:58, SEQ. ID. NO.:59, SEQ. ID. NO.:60, SEO. ID. NO.:61, SEQ. ID. NO.:62, , SEQ. ID. NO.:63, SEQ. ID. NO.:64, SEQ. ID. NO.:65, SEQ. ID. NO.:66, SEQ. ID. NO.:67, SEQ. ID. NO.:68, SEQ. ID. NO.:69, SEQ. ID. NO.:70, SEQ. ID. NO.:71, SEQ. ID. NO.:72, SEQ. ID. NO.:73, SEQ. ID. NO.:74, SEQ. ID. NO.:75, SEQ. ID. NO.:76, SEQ. ID. NO.:77, SEQ. ID. NO.:78, SEQ. ID. NO.:79, SEQ. ID. NO.:80, SEQ. ID. NO.:81, SEQ. ID. NO.:82, SEQ ID NO: 83, SEQ ID NO. 84, SEQ ID. NO. 85, SEQ. ID. NO. 86, SEQ. ID. NO. 87, SEQ. ID. NO. 88, SEQ. ID. NO. 89, SEQ. ID. NO. 90, SEQ. ID. NO. 91, SEQ ID. NO. 92, and SEQ. ID. NO. 93; or a sequence which is the reverse complement of a target sequence located in a region extending from about nucleotide 100 through nucleotide 500 in said SEQ ID NO.

An isolated polynucleotide for characterizing cells which are obtained from a subject 39. suspected of having a cancer selected from the group consisting of glioma, acute myeloid leukemia, primitive neuroectodermal tumors of childhood, breast cancer, colon cancer, head and neck cancer, testiclular cancer and lung cancer; wherein said polynucleotide is from 35 to 1000 nucleotides in length and comprises at least two CpG dinucleotides, and wherein said polynucleotide comprise a sequence which is identical to or complementary to a target sequence which originates between nucleotide 1 and nucleotide 15 of SEQ. ID. NO.:1, SEQ. ID. NO.:2, SEQ. ID. NO.:3, SEQ. ID. NO.:5, SEQ. ID. NO.:6, SEQ. ID. NO.:7, SEQ. ID. NO.:8, SEQ. ID.

10

NO.:9, SEQ. ID. NO.:10, SEQ. ID. NO:11, SEQ. ID. NO.: 12, SEQ. ID. NO.:13, SEQ. ID. NO.:14, SEQ. ID. NO.:17, SEQ. ID. NO.:18, SEQ. ID. NO.: 19, SEQ. ID. NO.:20, SEQ. ID. NO.:21, SEQ. ID. NO.:22, SEQ. ID. NO.:23, SEQ. ID. NO.:24, SEQ. ID. NO.:25, SEQ. ID. NO.:27, SEQ. ID. NO.:28, SEQ. ID. NO.:29, SEQ. ID. NO.:30, SEQ. ID. NO.:31, SEQ. ID. NO.:32, SEQ. ID. NO.:33, SEQ. ID. NO.:34, SEQ. ID. NO.:35, SEQ. ID. NO.:36, SEQ. ID. NO.:37, SEQ. ID. NO.:38, SEQ. ID. NO.:40, SEQ. ID. NO.:41, SEQ. ID. NO.:42, SEQ. ID. NO.:43, SEQ. ID. NO.:44, SEQ. ID. NO.:45, SEQ. ID. NO.:47, SEQ. ID. NO.:48, SEQ. ID. NO.:49, SEQ. ID. NO.:50, SEQ. ID. NO.:51, SEQ. ID. NO.:52, SEQ. ID. NO.:53, SEQ. ID. NO.:54, SEQ. ID. NO.:55, SEQ. ID. NO.:56, SEQ. ID. NO.:58, SEQ. ID. NO.:60, SEQ. ID. NO.:62, , SEQ. ID. NO.:63, SEQ. ID. NO.:64, SEQ. ID. NO.:65, SEQ. ID. NO.:66, SEQ. ID. NO.:67, SEQ. ID. NO.:69, SEQ. ID. NO.:70, SEQ. ID. NO.:72, SEQ. ID. NO.:73, , SEQ. ID. NO.:75, SEQ. ID. NO.:76, SEQ. ID. NO.:77, SEQ. ID. NO.:78, SEQ. ID. NO.:79, SEQ. ID. NO.:80, SEQ. ID. NO.:81, SEQ. ID. NO.:82, SEQ. ID. NO.:83, SEQ. ID. NO. 86, SEQ. ID. NO. 87, SEQ. ID. NO. 88, SEQ. ID. NO. 89, SEQ. ID. NO. 90, SEQ. ID. NO. 91, SEQ ID. NO. 92, and SEQ. ID. NO. 93.



### SEQUENCE LISTING

<110> Ohio State Research Foundtaion Plass, Christoph

<120> Detection of Methylated CpG Rich Sequences Diagnostic for Malignant Cells

<130> 22727/04075

<160> 90

<170> PatentIn version 3.0

<210> 1

<211> 677

<212> DNA

<213> Homo sapiens 2.B.53

<220>

<221> n

<222> (1)..(677)

<223> a or g or c or t

<400> 1 geggeegegg ttagettete etgteegaac geagggttte aetggggege egetaeggtt 60 cctatggcaa cgcggctcct cgacgcagcc caggagtcgc ggtcgcggga ggctgcgccg 120 cgcaccgagc tettecetgt ggccgccgca gccgccagcc tettectgct catgetttte 180 ctcatcttca tctcggtctg agtgggctct ggacctctcc accagcctct gccccagaac 240 tgttaactgc gggggggaaa aaaggaattt gtcgtcgcaa cgcgcgttcc gatggagccg 300 cacgccacaa aggaagactc atgctgcacc ccgcggggca gatgcggcga cactggacat 360 cgctgcacag ctgggtctgc ccgtttccag agctgcttag cgccgacgcc cataaatgag 420 gaggactccc tgtgtattaa aagggggatc cgcagggttt aatttgataa ggattatagc 480 cttcataaag gcatttttaa caaaaagatg taggtggcat ggtaatcgag tattatttac 540 gcatctctcc gcacacgcac tcatacctga aaacgttntg gcaggcacaa aatgattttt 600 ttgtgtataa aagaatgtgt gtaactcgtg gatggtgggg ttcagcagga caagatagtg 660 677 acattagata aattaca

<210> 2

<211> 380

<212> DNA

<213> Homo sapiens 2.C.24

<220>

<221> n

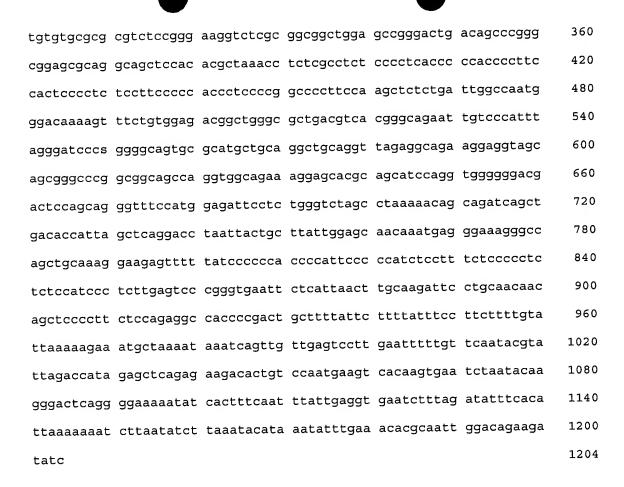
<222> (1)..(380)

<223> a or g or c or t

<400> 2 gcggccgcct	tgaaggcgct	ggacgggatg	gtgctgaagt	cggtgaagga	gccccggcag	60
gtgagctcgc	ggcccgccag	cccgctgccc	acgcagtagt	ggaagaggcc	gaagtagcca	120
ggcttggggg	tgctcacgct	gtcgcccacc	cagtagggct	ggatgaagac	caccacgttg	180
atgatggcga	agcagatggt	gaagatggcc	cacagcacgc	cgatggcccg	cgagttccgc	240
atgtantgct	cgtggtagag	cttggaacct	cctgcgaggg	cagcatggtg	cccggangcg	300
gggccggcgg	cggctgtngc	tggcngnggc	cgtcggcccg	ggacngacgc	ctggctgccg	360
ggcgggaact	ggggactcac					380
	o sapiens 2	.C.29				
<400> 3 gcggccgccg	cgctagtgac	tacttcctcc	tactccttct	cctcctgctc	cggcctcctg	60
gcgccctgct	ccaggctctc	cggcgccctg	ctccaggctc	teeggegeee	tccagccagg	120
caccggccga	accgggtagt	gccgcaaggt	gtaattactg	ctttgaaact	ttaaaggcat	180
ttggaaagaa	actacgggtt	atgcttactt	tttttgtttt	tgattattat	tttgtaggag	240
acacaaagtt	taaaaataga	aagcaaaaag	tgtgacacat	ttaaagagtt	aaaggaaata	300
aacgtttcca	atttacctta	taacatgatt	ttcatacact	ggatttgttt	aaaacagact	360
gactacatgg	ataacttttc	taggaattgt	tcttaactct	gatagctggc	tcaactgatg	420
taggcattaa	aataacgtca	tattaccatc	tttcctccac	gaattgatga	tatttgacta	480
tagctttgtc	agggttatgt	ccaacțattg	tataatatgt	gtcagtttcc	tattgctacc	540
gtaacaaatt	accccaaatt	tactgg				566
<210> 4 <211> 129 <212> DNA <213> Hom <220> <221> n		.C.35				
<223> a c	(1297) or g or c or	t				
<400> 4 gttcacttct	cgctgcgccg	cgggttctgt	agaagcgcaa	ı gaatggggct	gattattccg	60
gtgcccacat	geegeeeea	cacgccccca	ccccgtccg	gcgcaagact	tecettggee	120
aaaagaggc	g tttaattagt	tctggggccg	g cggagagcca	ı gegtggeega	caaagcccgg	180

		aaataaaaaa	aaaaaaaaaa	acacacaaaa	ccadaacacc	240
	aacccgggtt					
	tgcgcgctcc					300
ccccgcaggc	caaaggctca	tctgccgggc	ttgggtggcc	cgggccagcg	ccgcctgcgg	360
tccccgagtg	cggctggctc	taaggccggc	gccctctccc	cggctttcag	tgctcagagc	420
caggccagcg	ggaaagaagg	cagcatggtc	cgcaaaagac	aggtggcagt	ggcagtcttg	480
catgatactt	gtccttcttc	cctgttcccc	attttgggga	aacactggaa	acacttttct	540
ctttatgcgc	attcgcgtct	cagcaccgag	tgctccaagc	cctgcgcgca	gcgccgggct	600
tggaaggcgg	cgaatggctg	cctagccgcc	gcccctacta	gtgacactcg	gccgccagcc	660
cccgcccagg	atgtgcacat	ctgctggcag	cactggcccg	ggtggcagtc	accgggccac	720
ccactccaca	ggtacaaccg	cacccaatcc	aacctggaac	tcggagggct	gtgcgcgccg	780
agctgggatc	gcgccccaac	gagccgggcc	tttggctgcg	ccagggccca	ggccgagtca	840
tcccccgct	cgcgtcgccg	cgaggcggga	caccgtgtaa	tacctttgcc	gtgggctggg	900
cgtcggccgc	gggccggaga	gegggtgtee	cacctcgcct	catcatttga	tttccgccag	960
cgtctgagga	cggcgcaccc	aattcgttcc	actcgctgcg	ctctgtgaac	cagcggcggg	1020
cagggcgggg	gaggccgggc	tggggnaggg	cagggtggtc	ccaatccccc	geceegece	1080
cgccggcctc	gcggagcaca	agtgttggga	ttcccacggg	caggcgtgct	ctgcggctgg	1140
aggcccgagc	gcccagggcc	caggagacgt	ggcggacaca	gaggggtttg	taggcacggt	1200
gacctccgtg	ctcctgctct	gaaagggcct	gaaaggagcg	gtttatggtg	cattaccagt	1260
caagggctca	ggtaccagcg	cctgtgtcgg	gaacccg			1297
		.C.54				
<400> 5 gcggccgccg	cgggggacgc	tcagatctcg	cgagaagagg	gcgagcgcgc	tgcccccctg	60
gtgggcgggg	cgaagcccgg	gagagggtgg	gcgccaccgg	aggggaggag	gggaacaggg	120
aactgaagga	agtgggaggg	gccggcgggg	cggggaagcg	gaaagggggc	gtggctgagg	180
gcgggaggat	taagctgcct	ttttgaaagt	ggagcgccag	gtcccgggtt	ctgggtggag	240
gtggttgctg	attggtggag	ctcggagcgg	cggttgggag	ggtcctggtc	acatggtggg	300
gagtgggagg	ggggaagttc	ggagagcggg	agcgggatgg	, tagtgggctg	ggccccactg	360
ggctgggaca	ı gcaggaggat	agtcttgagg	aggagcgtgg	g ggtgctagat	gtgtaactac	420
gtcccgaact	ggttcctgtg	tttttctagg	gcatgtggad	tagggatggg	, tacttgagta	48

gaagcctgca	acttgaagag	tttgtgcagg	agttagctgc	agtgtcggaa	attagtgtcc	540
tgtatgctca	acaaggtatt	cggactgggt	gtgcacacca	cagctctcag	gactggaagg	600
tggaaattta	atctacgaag	ttcccttaaa	ctgcataagc	ttcgggacct	С	651
	o sapiens 2.	.C.57				
	(710) r g or c or	t				
<400> 6 gcggccgcac	ggagttgaag	acactaaccc	agctaagcca	catacagacc	ctcacggccg	60
cctggtctac	acaggccgcc	acagctacac	aggctcaggc	ctcagcctgg	tcacaatggt	120
cacacccaca	ctctcgggtc	ccacagtttt	gcgggagcgg	tgacacacac	ccgctcccaa	180
ctgaccacgc	ccacacacgc	tggcttcagc	cgcacacgca	cacagtagcc	acgccccctt	240
atgctccagc	cttgccagca	cccgccctcg	ccacgctggt	cacgcccaca	cacacacaca	300
cacacacaca	cacacgcacg	caggcctggg	gcacgcccct	ccccacacg	caggcgtgcg	360
gcacgccttc	ccatacacac	acacacgcgc	gcgggcctgg	ggcacgccct	ccacacacat	420
gcaggggtag	ggcacgcccc	cacacacaca	cacgccggcc	tggggcacgc	tegegegeae	480
atgcacacac	atacacacgc	acaggcctgg	ggcaggcccc	acccccacac	acgcaggcct	540
ggggcacgcc	ccccacaca	tgcaggcctg	gagcatgcgc	acactcgcag	gccttgggca	600
cacgcgcaca	cactcatgca	cagacacgca	cgcacacatc	gagccccgcc	cncggaagca	660
catgagaggc	acttgctttc	actgactgan	ggcanggctt	tgggcccgcn		710
		.C.58				
<400> 7 geggeegete	ctctttattc	tactctcacc	cgaggcccgc	gcccgtcccg	gggagcggct	60
ctgccaggaa	aacggcccga	ccagtgcccg	gcgcctgggc	: tgcgtccgag	cccaccttct	120
tccctcgtcg	tegtetecca	gactaaatcc	cggaaaggga	aagcgggatg	tttgcgccca	180
ccgcgctgta	gctggtcctg	acacttgcaa	aatggtcagt	ggctcctgct	cggccaggct	240
gagtgtgtg	gtgtgtgtg	gcaagggagc	gagggtgtgc	ggtgtgcagg	gggtgcgctg	300



```
<210>
       687
<211>
<212>
       DNA
      Homo sapiens 2.C.59
<213>
<220>
<221>
       n
      (1)..(687)
<222>
       a or g or c or t
```

<400> geggeegeae aagegeaeae geaeaegtee agggeggagg aacaetaeta gtaacaeeeg 60 cctccttcta gcctccctat cccaaagtta tggtgccgat tttgtccgcg gcaggggctc 120 180 caggggcaca ctcataaatt cggtgcggag gaacacaact agcagcacca cacccccgcc 240 actgccagaa ccaaagtgac ggtgccgaca cccctccgca agcgcaaggc cgacttccat aagtaattag ccagagcacc gtcccgttcc tgtcagcacc gagccccagc caggacaccg 300 gtattcccag caccatacaa gaactacttt ttcgatgaag caacccaaaa gctgcgagcg 360 gttcccggtg aggccgccca ctcacctggc cggcgcagac aagctccgtg cgtcaagaca 420 taacagcgta agtgtacgac gttgcgcagc gacgcggggg ccttcgggaa atgtagtcta 480

caactggaaa	ccggccggat	cgtgtctgcg	caggcccagc	agctaagatc	gggtccggcg	540
ctccagaaca	gaacgatccc	tgaggctccc	ttgctcgaac	tgtgggactt	accctactat	600
ggtccgagcc	taccctattt	cattatactc	aagtaacgcc	ccagaaattn	cagagaatct	660
acacaaagag	gttgagtctt	gccgtgg				687
<210> 9 <211> 1520 <212> DNA <213> Homo	sapiens 2	D.10				
<400> 9 gcggccgcga	ggacagctcg	gacgggggag	agaaaggagg	tttccagtaa	aaataataac	60
gccagagaga	aaaccgtaac	tcgcgtgaca	cagacagaaa	tttccagtaa	taatcatcag	120
gtgatagaga	aggaaggctt	ccaaaatgaa	gaacaagtga	aataaaggtt	ttagtcatga	180
attacagcac	gtgcgatgga	tgagtggtga	tttctcatca	taaatggtaa	ctcgggagat	240
agagaaacgt	gtccagccct	aaactacaac	agggtttggt	ttgaaagaga	ggtgctgtca	300
taaagcggaa	ctcaggggat	ggggaagacg	gcctccgtcc	caaatgacaa	ctcaatgaca	360
gagaacaaaa	gatccaaact	aaagtgatgg	agaaaaaggg	tttccaacca	ccacacaaat	420
gaagagaaag	actgatcaca	taatgaagta	ttcagtcatt	aatacatgat	aaacccggtg	480
atagagaaag	aggcttagtc	acaaattact	cagataatgg	agaaaaaagc	cttattcatg	540
tatcactcag	gtagatacat	caaggcaggt	ttcctgccat	aaaggataac	acagctaaaa	600
gagaaataaa	ggttttagta	ataagtgaca	attcatataa	cagagaaaga	aggcttctgg	660
ccataaggat	aactcatgta	ataaagaaaa	gttttagtca	taaataatag	agaaagaaag	720
gtttccgata	gaaaatggta	gagatagaaa	ggttctaggt	aacaaacggt	aactgaagtg	780
atagagcaag	gtcacaaata	ataactcagg	taatagagaa	agatttctag	tcataaataa	840
tacatctgct	acagaaataa	gggttttgat	tcataaagtt	atgtcataag	tgataagtgg	900
tagaaaagga	aaggttttag	ttataaatta	tgattcaagg	gatagaaaaa	caaaggtttc	960
aagttataaa	tatcatttca	atggtcaaga	aaggttttca	gtcatgaatg	aaaactgggt	1020
gaagttttcc	agtcacaggt	tataactcag	gcaatggaca	gagaaggaaa	gatttttgtc	1080
atcaatcaac	tcaggtggag	aaggaaaggt	ttttcaataa	gaaataactc	agttgagtga	1140
aagaaggctt	gaggtcatga	atgataatta	ggtgatagag	aaagaaatgt	tccagtcata	1200
agggttaaat	cagatgctag	agaaagaaag	gtttttagtc	ataaataaaa	ctcagctgct	1260
agaaagaata	gggctaccag	tcataattga	taactcaggt	gagagaaaga	ttgctggtca	1320
taaattgtaa	cccaggtgac	agaaaagaag	gtgtcactca	. cacatgataa	ttcgggttat	1380

gaggaaggtt tccagccaca gtggtaactc aggtgctagg gaaagaaggt ttgggcaata	1440
atgacaactc aggtaataca gaaaaacgat tacagtcata aatgacagag aaggaaaggc	1500
ttttattcat aaaggatatc	1520
<210> 10	
<211> 575	
<212> DNA <213> Homo sapiens 2.D.14	
<400> 10	60
geggeegegg etgtggetee tettggeege geagetgaea ggtaaggegg eggegeggg	120
gctacccaag ggtctgcgct cccggggcct gagcggggag gtgataagtg gctgtcctgg	
ccctggtcct ggcagggtgc agcgtcgagc ccgcggtggc ggggcgcccg ggaggcagct	180
tggcaggcac ggtccctaag ggtggaaata aaataccccc atatcgcatt accccggggg	240
accggagage ccctggactg aggccaccte ccctcaaaag cctggacgca ggagaagggg	300
aggcagtgaa aaggggagcg agtgagggaa ggaaagagag ggtcgctgga ggtcaccagg	360
ggaaggaaac aggtccctgc ccagggtccc cgcaggatgt gctcggagga aggttggcca	420
ggccatgggt cctgtggaca catttttatt acttccgggg aagtgtttgt agtacaatca	480
gacaaacatc gggcgttctc agttctcgga gggctagggc agggtgatcc ctctggctcc	540
cgttctccct gatgtcgctg gtgttgggtg tcatg	575
<210> 11	
<212> DNA <213> Homo sapiens 2.D.20	
<220>	
<221> n <222> (1)(741)	
<223> a or g or c or t	
<400> 11	
<pre>&lt;400&gt; 11 gcggccgcgt cgtcgctgag tacaccagct gcctcatcta tctggagccc ggcctccatc</pre>	60
togocaggot cagogocogo gtoogtgtog gtgcoggago cattggcogo gcctagcaac	120
acctegtgta tgcagegete egtagetgea ggegeegeea eegeageage etettateee	180
atgtcctacg gccagggcgg cagctacggc caaggctacc ctacgccctc ctcttcctac	240
tttggcggcg tggactgcag ctcataccta gcgcccatgc actcacatca ccacccgcac	300
cagetgagee ceatggeace etectecatg gegggeeace ateateacea eccaeatgeg	360
caccaccegt tgagecagte etcaggecae caccaccace atcaccacca ecaccaccaa	420
ggctacggtg gctctgggct tgccttcaac tctgccgact gcttggatta caaggagcct	480

ggcgccgctg	ctgcttcctc	cgcctggaaa	ctcaacttca	actcccccga	ctgtctggac	540
tataaggacc	aagcctcatg	gcggttccag	gtcttgtgag	cccaggaatg	aaagaggaga	600
agaaacgcaa	ctacctgcgc	cctccgtggt	cccgatcctg	ttgctgctgc	tgcaccgccc	660
gcctttgcct	cgtcttctcc	aaaaactgat	tntcaccccc	caaaagatgt	ccattgcctg	720
cactgccgcc	cncatttttg	t				741
<210> 12 <211> 458 <212> DNA <213> Hom		D.25				
	(458) r g or c or	t				
<400> 12 gcggccgcca	gtagcagagc	ccagcacatt	gegggtgeee	agttcatctt	cgtggggtta	60
aacctgcggg	aagagaggga	aagggccctt	agtttccatg	gagatcgggt	gcccaggggc	120
ggagggctca	aggctggaga	gcagagggac	ccccatcttt	tgtgggatca	gggtgccccc	180
agcatcttgg	aggcccactg	aggcctgggg	gggcgcggtt	taacttctag	catcagggac	240
ttaggcctgg	gggaggcgct	gggaagtggc	aggtggggca	ggagggttct	gcacctgaag	300
gttgtgcacc	tggattgggg	gtgtagaagc	ggngcaggag	cgccgcggtg	ggggcgtcca	360
ggcccgggcg	gnggagcaag	cctgggggag	ggagctctgc	acgcgttgct	gggatgtggg	420
gggcgngggg	aggcggcatg	gggggagggg	cgttgtgt			458
<210> 13 <211> 615 <212> DNA <213> Hom		.D.27				
<400> 13 geggeegee	ggcgtcccgc	tctggggggc	cgggaccgaa	gcgctcacgg	cccggggacg	60
cggggttggt	ccaggctgcg	gcctgtggcg	cgtgcaggcc	tgaaggaggc	gagatgccga	120
tgccgccaco	gctggtccgg	tggaccaggc	cccttggtcc	ageeteecet	cccgcagccg	180
cccgtctggg	g ggtgttcgca	gccccgggct	cccccggccc	geeegeeggg	gagtgggagg	240
gcgatggcg	c cccgcctccg	gctcttacgg	agagegegee	tcccctcaa	ctccggcggc	300
ggtgagccg	g ggtgcgatgc	gcggccgagg	cctcgcccgg	g accgccggtc	cccatcgcgt	360
ccctgggcg	a gggaggggg	gttggccgga	gatggcggag	gggcgtaccc	gccccgcctg	420

cccgccgtcc	ccagccctca	gcgcctqqqq	aagcccctqc	tgtggcagtg	ctcgggcgct	480
atccggagga						540
						600
aactaacttg		cagccgcccc	gegeeggetg	ceggetaget	caggeegaeg	
ccgaggggag (	cggcg					615
<210> 14 <211> 669 <212> DNA <213> Homo	sapiens 2.	D.34				
<400> 14	~~~~	anaanatat	taatttaaat	acaccttaga	cccctacata	60
geggeegege						
tctcccaggc						120
ggtgccggga	aagggcgctt	ctccccagtg	aggtggggaa	cttgggtgat	gggaccacgg	180
aggcgccggt	tcgtgcccgg	tggggacggg	tgaggcaggg	gagagtgaga	ttttattctc	240
ccccaaggaa	ggagtgtccc	cttctcctta	ttttgagggc	tattcaagct	tattgaaacc	300
agaaagcggt	gtttcttgtc	aatctctcag	ccccttcttc	caaccaagaa	caattgtcga	360
tgagtttcca	tcacaggcgc	ttgtgagaga	accggtaaac	ccagtacagc	aaaatccaag	420
cccttggttt	ccacatgcat	tttgctagca	gtttttggca	ttgaccctcg	ccctcccgtg	480
tttccactcg	acatcattta	gcgtttgagg	ttttttccc	tcctcaaaat	tgcaaatgag	540
aaaaaagag	gaaaccagga	aaagggggtg	gggggtagca	tttaaattgg	atgtgagttt	600
ctgctgagaa	ttctagcgaa	gtcccctgta	cactgaagcg	ccgagagatt	tttccgtttg	660
tgtatcttc						669
<210> 15 <211> 998 <212> DNA <213> Homo	sapiens 2	.D.40				
<220> <221> n <222> (1). <223> a or		t				
<400> 15 gatatccatt	ataatactat	ttgacctcaa	agtgaatttt	attgttccac	acaagcaaca	60
gattacacca	atttcacaac	tcccagaatc	caaacctaca	aagacccttc	ccaccaagca	120
ctttaccaaa	aacgggcttc	atctccatct	tcctttcttt	cacagttgaa	aaactgccct	180
tcctaattaa	gccaaccaac	ttcttacctc	aataaaatcc	ttgtttttca	gtagcatgta	240
cagtatttcc	agtgatgaac	agtgaactgt	ctttcgtctc	acacagtaac	ctccgtgaag	300

360 aagatccacc ttgttcttta ctgtatattc ctggcatgct aactgcatcc tcagacaatt 420 ttaagtgact gaaaactcag gcaaagaaag gcaagagggc aaatagaagg gcacaggaga caacgctttt caaatttttc tcactgcgac ctacagaaac acactgtaga acacctccta 480 gtacactcac acgtgtgtgt acacctgaag tgtcaagaaa caatacccta agtgcaacac 540 cctctgatat tttctatttc aagtggccgt gatctactaa actgatttcc aactcaccaa 600 taggattcag tttgaaaaac actgcaataa atcaaacctt acagttgcat tccacaagct 660 actaatgaac tottgaaaat ocagcataca goagagacgo tgaccaacta caagatccaa 720 acccccagg tgggcagtgt ccttctgttc agcagtggca gttccccacc accaccagcc 780 ctgagagtta attatctccc aaactcccag agtttcccaa gtagcctgag gtgtctgtca 840 900 tatgcccttt taacctcttt ataaattcag tcccgtccgt ctcttacggt ggcaaagttc atttatcgtc ggctgtggaa agcaatacnt tctttttgtc cccttcagga acccagaatt 960 998 aatgaccagg ttggtgcccg gtgtgccttt atgatcta

<210> 16 <211> 797 <212> DNA <213> Homo sapiens 2.D.48 <220> <221> n <222> (1)..(797) <223> a or g or c or t

<400> 16 60 gcccctctga gttacgggga gccctgcaga cacccagccc ctggggatcc tctccccgac ctgcccttcc cctccgacac ttgccagtac tccccggcct ggtattcctt tcgagacccc 120 ctcacctatt ccaggctgtc ctccactgag gcgaagctct atgaagtagc ccaatttcaa 180 tataattcac gttgtgtaaa agaactttga agacggacta catcgtgcaa ggacaccgtc 240 acccgaaaac cattggtgga acgttaaaac aaacaaaaaa caaaacggca aaaccttttt 300 gaaggcaatt ttgacattta tgaatttaca gttattattc ggtttgtccc tgaaatgtca 360 cttctgaaaa tttgcatagt tttcattatc actaaaataa tctagtaaat attcccgaat 420 gaatgcattc aagaatattc actaaattat tttagtgata aggaaaaagt ggaaatagct 480 gacagtcatc aatttataaa taaaatgatg gttaaataaa atgatgaaca ttcatataaa 540 ggaatactct atattcagac gagatctgtg tgctcacagg caaacaggtc taagcttact 600 ttaaatgaaa aaggataaat tgcaaaaaga atagtttgtg taatatgatt ccacatttgt 660 720 aaaaatggag aaagaaatng taagcanatg tctgcaagca atcagatatg attagtgact

<220>

taatttcatg gatagttata tag	gaaatat a	atgtatattt	tatatgcaca	tagatatgga	780
ggaatatact ttcactg					797
<210> 17 <211> 1024 <212> DNA <213> Homo sapiens 2.D.5	55				
<220> <221> n <222> (1)(1024) <223> a or g or c or t					
<400> 17 geggeegegg egetgeaegg ge	gtgacgtc	atggcgccgc	ggagccgcgt	cctccccgcc	60
ccgccccgg ccggggtcac cc					120
ctgcagcctc ctcagtcgtg cg	tgcgttca	ttccgctcat	agcttctgtc	actcagcaag	180
cgctcaacac agacgcatga ga					240
caacacgtgc ttagcgcgct gc	tgatctgt	gccaggcact	gggccagggc	cccgacacgc	300
gtcagggtag aagcaagcag aa	gcctggcc	ctgttggagc	ttacattggt	aaataaccaa	360
gataatttca ggtaaatatt ag	gtcctatt	aaaaatatgc	gtcttcgcca	ggcgcggtgg	420
atcacgcctg taatctcagc ac	tttgagag	gtcgagcacg	ggcggatctc	ctgaggtcaa	480
gagttcgaga ccaacctgng ta	aatggtga	aaccgcatct	ctacaaacat	acaaaaaaa	540
aaattagcag tgagctgtga gc	ttgcacca	ctgcactcca	gtctgggcaa	caggacgaga	600
tcttctaaca acaacaaaaa aa	aagtatgg	gccacctagt	ccagccaaaa	aaacaaagtg	660
ctttttttt gcttttttt tt	tttttt	tttttgagat	ggagtctcgc	tgtgtcgccc	720
aggctggagt gcaggggcgc ga	tctcagct	cactggaagc	tccacctncc	gggtttacgc	780
catteteetg geteageete ee	gagtagct	gggactacag	gcacatgcca	ccatgcctgg	840
ctaatnnttt gattttttgg tt	gggtgttt	agtagagacg	ggttcatcgt	gtagccagat	900
ggctaactct gactgtgatc to	gcacttgcc	tccagtgtgg	atacagggga	ccacttgcag	960
caaagctcta ttcctgtagg ag	gggtgttg	tgaatcagac	ccaatttgga	aatcaaattc	1020
tagt					1024
<210> 18 <211> 1854 <212> DNA <213> Homo sapiens 2.D	. 74				

<221> n <222> (1)..(1854) <223> a or g or c or t

<400> 18 60 gcggccgctg cagaccctgc tccaggcgcc gtagccttgc aggaagagca gacaaagaca ggagagaggc aaagcgccgc ttgcccagag atgcagtcgg ctcagtcaat agagggaaat 120 180 cgcctccaaa cccaggctgg gaatgaggga ggaggggcga ggcggctggg gactagaaaa 240 agcagcaggg aattaacgtg acagtcagag cccagccagt gcctcgccgg cgctgctctc tcgcctcgcg gttgcggngt ccggaatgga gagaggaggc gggggctgag ccgttggctg 300 ccggagacca gctgaggtag gagtattaac tccctctgct gctctcgcct gccttcctcg 360 420 cacccctta cacageteta ettgeageag getatggeee cattettet eetatttte taactactga gatcagagct gaattaagct ggtgaaagga gcaaaacgtg caagggattg 480 attgccctcc ttgggggaaa agcggaggct taaaatcaat tcgacaaatg agtgtttact 540 gggtgctgag tactgtgctc cgctattgtg agggagggtt atgaataagg taccccctc 600 ccgccccagg gtccgttgtc agatctcaga atcagtttcc cctgcagttc tggaagccca 660 720 aagtttcggg gttgagttgt ggtccctgat cccgatcctc aaccaatcta gctttctaaa tcagaagaag gtggaattca attttccttt ctccttcctg ggatgacttt aacctgcagc 780 cgaaatggag tctataggcc ccttaaaaaa gcgcgcgcac gccagtgtgt gtgtgtgcga 840 gcgcgctcgc gtgcgcgcgt gtgttttaag agtaagtcaa attaatggtt ttagtgatgt 900 tottatttca tgattttaat tatttaccat atctgcagta gacaccagtt tggggcagag 960 gaacccgcct ctccagactc tacaaatacc acctttttt ctaaagcttt tttccgctac 1020 cccagtcctc tgactcgagg cagaaatctt tcccctctct ttgccctctc agaattttat 1080 ttgccaatca cttgcggaac ttatatattt atagatttat ctcttcactc acatatgagt 1140 attccctgtg ctttttgttt gtttgttctc actgcaacat ccagcagtgt tttgtatcta 1200 atgggtactc aaggaaagct tatccagttg aaggtcattt tctccttctg tatgagctaa 1260 atctcagtgt ctctagaatt aaagagactc cagggatgga acttttgatt tagggtgtgg 1320 1380 tgaagggacc cacacataca gttagactca cagccccttt actggaaagg taataaagta tttaattcat tttggtctct agacaatcaa ccttctccca ctgaccaccc acctctgttt 1440 cctgaattcc caaaagcaaa agaaaaccaa actgctaagc aactgcctag agcaagacat 1500 gtatgttcag ctgccaacac ctagagcaaa cccattccaa gtggagaatg accaaaaaat 1560 cttgattatt tcttgacctg tgtcaagtat gttgaaagcc tgccaaagtt tcctcatttc 1620 tattgaagca ctcttattct ggatgcattt tagaacagtt tgaacagtgt tacattgctc 1680

agaggtgaag	aaaattgctt	tgtagtttaa	ggatatttaa	gatttgtttg	tttgtttgtt	1740
tgttttctgt	cccaccttct	acaaattgca	cgatagatac	ctcagatcag	gaatgctgca	1800
tgaaaaagta	tgtccataat	gcaggagatt	agactaaatg	actcttaaga	tatc	1854
<210> 19 <211> 674 <212> DNA <213> Homo	o sapiens 2.	E.20				
<400> 19 gcggccgcct	tcccttccca	ttcactggct	gcctcctttg	tgaactaatg	actgtaatta	60
ttacctccca	gagctctttt	gttatctcca	accccaagcc	ccggagaggg	ggaatgggct	120
ctttagtgaa	atgaaagtca	ttacaaagca	aattaccgtc	tagggaggga	cagccttcag	180
gaaagacaaa	tcagatctcc	atctgcatct	gaagtagggt	gtgtttaaat	aaaaaatgta	240
aatatcacca	ttagatccaa	agtactccag	agctgtggga	tttaatggag	tttaaacggt	300
agcacttgaa	gccattgctt	taccaaaaag	aaaaaaaaat	cagttaaatt	caggtgtttt	360
aatccgtttc	ttctttgggg	gttttgtgtg	atttaaacgc	ttgcttttaa	gaacctttat	420
gttttcaacc	actcatccat	agtagaaaag	ttctgcaacc	ctagactgct	ggcttgaagg	480
aaaacctttg	caggatttga	tatggatttc	aacaaagaac	cagcctctgc	gaggctggag	540
agagctgcgg	agctgccatg	cctgaagtgc	agatggctga	accacaagtc	tttaggtttc	600
cggagttgtt	attgtggtga	cctagagtgt	cagagccagg	agagcaagaa	agaggagcca	660
aactgagccc	tgag					674
<210> 20 <211> 676 <212> DNA <213> Hom		.E.24				
	(676) rg or c or	t				
<400> 20 gcggccgcag	ı acgegecagg	cccgccaggg	cgccgcacgc	: cgggcgcgcc	acgatgtcca	60
cgaagcccac	gatggacagc	aggaaggcgg	cgtcggtgtc	gggcacgccc	gcgtccttgg	120
cgtagttcac	cagcaggatg	gcggggacga	agagcccgag	g cgccatcagg	aacttggtga	180
cggcgtacac	ggcgaaggcg	cggtcggtgc	acactgccaa	gtccagcagg	cgccggcggg	240
accadacac	aaaaaataaa	tacacaact	acaaccccac	accatcaacc	tecqceteqe	300

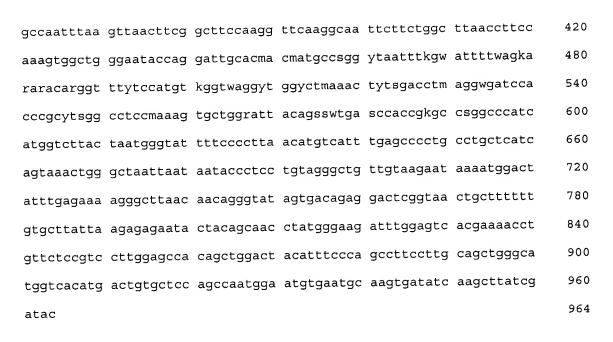
ccggagcgtc	cccggcgcgg	tcgccggcgc	tgtccctgcg	cggtcgcggg	cccggcccgg	360
geggeggeet	catgacagcc	ccgcaggcgc	agcagtgcag	caggagcccg	ccgagcagca	420
ggaagccgcc	gcgccagccg	aagcgctcca	gcagctgctg	gccgagcggc	gacagcgcgg	480
acaggaacac	ggngctgccc	gccgncgnca	gcccgttggc	cagaggccgc	cgncgctcga	540
agtacagccc	cagcatgatg	agcgacggct	ggaagttgag	ggccaggccc	aggcctgcgg	600
gcgaggcggt	gctgtgccgg	ggtccccgga	gagcccctcc	ttgggcccca	caggagggag	660
gggccaggcc	ccggaa					676
<210> 21 <211> 455 <212> DNA <213> Homo	o sapiens 2	.E.25				
<400> 21 gcggccgcgg	ctgggggcgg	ggagggggc	gcaggacccc	aagtgggggt	cccggagcca	60
gaggcaagtg	tcctggggtg	ctgggggcgc	cgtgccggcc	gggccgctgc	cctggcctag	120
gctggtccgg	gggctagcgc	gccgggggct	gcggccgatg	ggcggggcga	ggggccgcgg	180
gggtggcgag	ccgggggggc	acgggggtcg	ggggtgcccg	agggggcgcg	gccgggcggg	240
ggtggccagg	gatgggggtc	actgggggca	aaggggatcc	agtgggggg	tcccgatgga	300
ggcgtgcagg	gccaggggcg	cccgaggcgt	gcgggggtcg	ggtgccccag	actggtggcg	360
tcagacaggc	gtgggtcgtt	gggggcctgg	gtcgcggctt	gactgagggc	ccggccgggg	420
ctgtggggcg	tcaggagagc	gtggggtgtt	atggg			455
		.E.30				
<400> 22 gcggccgcgc	ttcgacgacg	acgacgacto	cttgcaggag	gccgccgtag	tggccgccgc	60
cagcctctcg	gccgcagccg	ccageetete	tgtggctgct	gcttcgggcg	gcgcggggac	120
tggtggggg	ggcgctgggg	gtggctgtgt	ggccgg			156
<220> <221> n <222> (1)						

<400> 23						
	cagtgcgtca	acaggcgctg	taatccgagc	gcataaacga	ggggtccggg	60
ggtgggggcc	cggggcggcc	gtggcagtgg	cccggggctg	gcagcccgct	ttgaaaatct	120
ggcgaagtcg	gggagcctgc	gtttgctttg	gcagctgcga	aggcgcacag	gtgcacgggg	180
gcggggggct	ggctggcggc	gccaccaccg	accgtcactg	acagagcctc	gccatgggcg	240
cccaaattcg	ttcacttgcg	aattgcgtaa	geggeeetee	ggtacccaac	ctctgggaat	300
tacgcgggct	tgtgcctgtg	gccaccttgc	taggccccac	cgctccagcc	tgaactccca	360
ccgctccctg	ccttgcgctt	gatgttccag	caacttcgaa	ctgtttttat	ctcctgtaaa	420
ccaagccgct	tctctccttg	acgctggcct	tcctgcctgg	cttgccctcc	cgccttcttt	480
tgccttttaa	gaccgggcag	ctatcccacc	ccgccagtat	atgcccctct	tctgggctcc	540
ttggcttcct	gtttatacct	acgtgactgt	gcttactttt	ttgcacatgg	tttttcttat	600
ccttctgtaa	gtttcttgaa	ggtaggagcc	atgtcttacc	ctgccaagca	cattgtctgg	660
cacgtagtag	ctgttcagta	gaggaagtgg	tccctttccc	taaagggctt	tncgtctcac	720
tggagagaaa	ggctagcctg	gtaccaggga	ctgccgagat	caagtgatgg	cagtacgtgc	780
gattcgatgg	tgccgaaagt	gacctagaga	ggcagctgng	agtgctctgg	tgctcgcgga	840
tagagctttg	gcgatattgt	catttacaat	gaggactgta	ctctgagacg	tggaccttct	900
aacagaccat	tataaccttt	gctctggagg	agtgagcnag	caacggactc	tgacancatg	960
ttttgacaat	gggtattg					978
<210> 24 <211> 321						
<212> DNA <213> Hom	o sapiens 2	.E.4				
<400> 24						
gcggccgcac	cggctcgggc	tctgccaagg	gacccggcct	gececaatge	cgccggcggg	60
cggtgcccgg	tegaceetge	acctgactgo	gaggcgcggg	aaatgaccgg	gtctgtcagc	120
ctcccatcgc	ggcttccgtc	tacaggtact	acctgtgctc	: tgtccagcct	cagccactgg	180
acgatccttc	ccgtagccgt	aggaagggg	ggcgcttcct	: tggaggggat	attagaggcc	240
cgaattcgcc	cgggaagcgg	cgggagggcg	ggggtgccgg	g gaaggaggga	ggggagaagg	300
agtgagggaa	gtgggtgtat	g				321
010: 05						

<sup>&</sup>lt;210> 25 <211> 1023 <212> DNA <213> Homo sapiens 2.E.40

<220>

<221> n <222> (1)..(1023) <223> a or g or c or t <400> 25 geggeegegg getgggggeg agegeaeaee eegegeeget ggagtteaet geegggegee 60 ggcatgggcc tgggggaggg gtgcacaggg cccggagggt gcgtgggtgt ggggtgcgcc 120 cggaggagag cgaggctgcc agagtgcgtg tgccgactga gccagtgtga gtgtgcaggg 180 gctggcggag agactgggag cgagtgtgtg tgcatctaac cgggaggttg tgagtttgtg 240 tgcgcgcacg cccgcagaga agttgtgagc ctgtgtgtgc acctaacaca gaggttctaa 300 gtgtgtgcac ttgtatgtgt gtgtgcacac gcggacagag tgattgtaag gatatgtgtg 360 cacctcacag agaggttgtg agattgtaag ggtttgcgca cctaacggag atgttgtgag 420 tgcttttttt cctgacaggc tgtgagtttg tgttgtgtgt attagaggtt tgtatggacc 480 540 tgactgaggg gttgtggaat gtgtgtgcgt gagcatgagc ctggagaggt tctatgcctg ttcactcctg acagagtttg tgagtgtgta tgattgtgtg actacaccac ccaactggcg 600 660 gattgaatgt gttgtataca tctactgnga gggcgtgtgt gtgtgtaaat tgtatacaat gaggetgtgt geateagtge acctaaceae gaacetgtgt gtacagatgt gtgtgeettt 720 ctgtgtatca gacatgaggc catgtgtctg ngtgtgttta gttggttgtg caagtgctgg 780 agtctggggg ggagagaggc agttcggagc cttcccgctt tctccttctn cactctntgc 840 ttgtctcggc caccagcatg ttggaggact acaaggctgc ccttcaggcc ctttagaccc 900 gettaaggea ettgtgatee tatatgeeag atgeeeteee aaagtgeeag getaeeacat 960 ggcttggctg attgattggc attgaccacc catttgttct ttgcttcctg ggcgggtcat 1020 1023 aaa <210> 26 964 <211> <212> DNA Homo sapiens 2.E.61 <213> <400> 26 agccacatgt gtacccatct teeteetetg tggaaggegg aaggaaacag atgeeeteea 60 aatatggaca gctgaaatga tgaagtgctg aagccctggc ccagaccctc agagagatgt 120 actcaaccac ctccccaccc ttggacaagc acaaaaccag agaaaacaaa ggccagcaac 180 tgtggctcag cccgcataaa tttcttctgg acactggcct gtctatttga atatctgtaa 240 tgtttggtgg agtcaggggt gagggtctca gcctttggct gctgcatctc cagacaccaa 300 tcatggggtt cttttctttt ttttaatttt ttttttttt ttggaaccgg attccaaggg 360



<sup>&</sup>lt;210> 27 <211> 748

<400> geggeegete egttgaetge agggeeeegg eggtetteet eegetgttee gaggeegttg 60 agggctgatg tgctccatcc tcccacttgt ggtttggcaa gccatccagc cgactacaaa 120 cccacgtttg tgagttacct gctggctgtg acgcttccgt caaatctgag taacagtttc 180 ctcatctcta agatgggtaa catagtatct acctcacagg atcgtgtggg cagtacatgc 240 atagaaagga tttaacacgc agtgtactca gctagtttta ttatttatcc gtaatgatca 300 tttgttcttt tcccctaact gtgcctcaca agcatgaaac agaatccacc aaacatttag 360 gtctgggtag tggttggatg gaaacccatc gcgggttaac gcttccaaca ccagtccctt 420 gacactetee egeegaggag getgatttgt aaaettgetg agaagagaat acceageaga 480 tctttcaggt ttcaaatcca cgttctttac aagttgtgtt aattgtttgt atatgctttc 540 gatatagagt ctctaggaag taatactagt acatgtttta aaattcaaat actgccaaac 600 agtgagatgt aagtctccct cctaacttct gtttcccaaa tcccatgtcg tttcttctga 660 tgcaatagac attgtatgtg tgtgtgtcta gatagataca tatgtgtatc tctcggcttt 720 748 ttttttttt tttaaagagt aaaccaag

<sup>&</sup>lt;212> DNA

<sup>&</sup>lt;213> Homo sapiens 2.E.64

<sup>&</sup>lt;210> 28

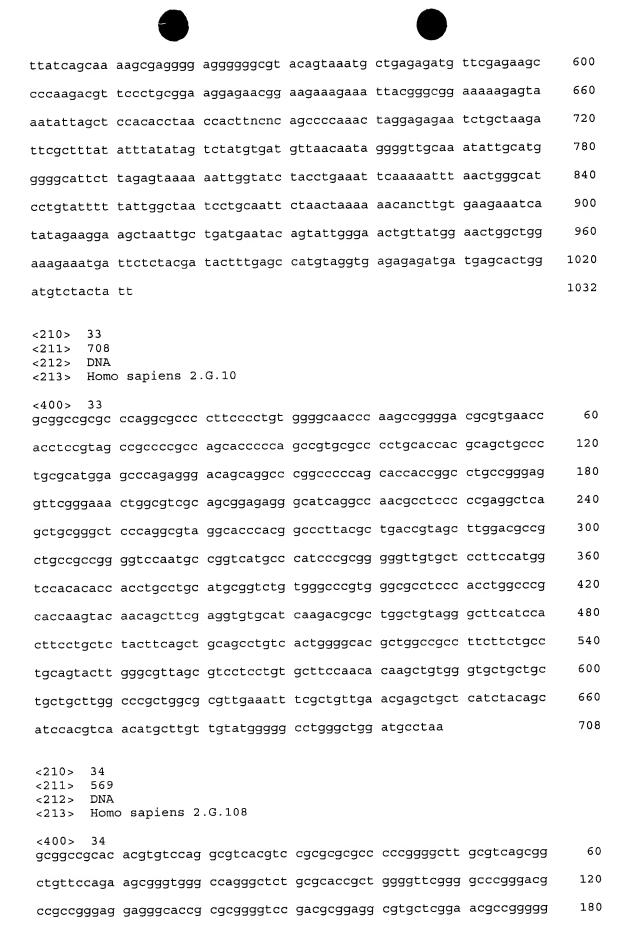
<sup>&</sup>lt;211> 250

<sup>&</sup>lt;212> DNA

<sup>&</sup>lt;213> Homo sapiens 2.F.2

<400> 28 gcggccgccg	gggaagggcc	ctggaagagc	aggaccaggc	agagcgggcg	ctggggtctg	60
cgctggagct	tgcgctgagg	ccggggtctg	gccaggagcc	gcagttgcag	ccgctgctgc	120
cgcagggtct	gaggatgagg	ctggagccgc	agcgggaacc	ggagccgcag	ccggtgctgg	180
cgttggcgct	ggaactgagg	ctggggccgc	cgccgggact	ggggttggcg	tggccggagg	240
agcacttact						250
<210> 29 <211> 657 <212> DNA <213> Home	o sapiens 2.	F.41				
<400> 29						60
	acggacagcc					60
	ctcgggccgg					120
	ageegeegee					180
tgtggggcta	gccctcgcgc	ctggcctgca	tcaggaccag	caacatggag	gcggccgttt	240
gcgaccccga	cacgcgagga	ccagggcggt	gcggagcccc	gcgaggacgc	gacgcccatg	300
gacgcctgtc	tgcggaaact	gggcttgtat	tggaaactgg	tcgacaagga	cgggtcgtgc	360
ctgtttctgg	cccgggcgga	gcaggtattg	cactctcagt	ttcgccatgt	ggaagtcaga	420
atggcctgta	ttcactcgct	tcgagagaac	agagagaaac	ttgaagcgat	tatagaacga	480
ccatttgaag	gaattttaaa	gcgcttcgga	aattcacagg	aatgggtatg	acaaatggaa	540
aaaagagccc	tttctcttat	gtacaggaaa	gattttattc	ctaaactgga	gccaaaggtt	600
ctttctcaca	agtaactgaa	aatattttcc	tgaaaggggt	tactggtgtt	tttaaat	657
<210> 30 <211> 318 <212> DNA <213> Hom		.F.50				
<220> <221> n <222> (1) <223> a o	(318) r g or c or	t				
<400> 30 geggeegeg	agcgattgca	tgcaggggcc	gcgtaccgng	aagtgcagaa	gctgatgcac	60
					ccagtggcta	120
					acacggcaag	180
					tttccacgcg	240

cgcctggtgc accagngggn ntgcccacgc ggtcgcagna tgggcgcagg gcaatnncaa	300
aacancactt gggcccng	318
<210> 31 <211> 525 <212> DNA <213> Homo sapiens 2.F.59	
<400> 31	
geggeegeet eeegeeagga agggtggegg geeeggaagg eeagagatge eeeagtgett	60
cccgcgccgc tacgcaccta gctgcccgcg ggtcccacat ggctgcggcc ggagggtccg	120
caccaggacc gccgccgct ggggaagcgc ttccctgtgg gcagggcgcg gcgggcagtg	180
cggaagcccg aaagctaccg gagcccgggg caggggggcg gcgatgcaga ggcggcgttc	240
gggggccccc agctgcctgc ggctcggcta cccagccgcg atcagagggg gcgggggacg	300
caggaacccc ggcgtccggg cggtgtgcag ccgcagacct attccaagtt tccacgtagt	360
tgcgagagcc caaaaactgt cacgtgcacg tcgctgctga gtgggaggag gtgtttgtca	420
tegegtteaa aaggggegtt teggtgtete eegteatgea agcaaatggt atggeteteg	480
gccgcctttg aataaacgag tgcttcgaac cctttaccag gaggg	525
<210> 32 <211> 1032 <212> DNA <213> Homo sapiens 2.F.70	
<221> n <222> (1)(1032) <223> a or g or c or t	
<400> 32 gcggccgcgg ccggggggct gagaagggcc tgggtgcctg tcgcccggga gccgaggttt	60
	120
cccggcctcc cccgaccccg ggcgccaaga gcagtcggtc cccccggcct cccgccggca	
aaggggccct ggggcccagg cgcgcggccc ctgcgtggcg gcaggcggcc caggccagcg	180
ccggcggcta gagaaggcct ccagtccagg cctcatggaa gggcctgcct cgagcggccc	240
ctcaacgccc cgcagtgtgg cactggaagg gacctaaaaa cccacctggc tttctccttt	
	300
ccccttcccc acgcttccca gggcccaatg cccgcatctc agtttcgctt tccggcaggg	300 360
coccttoccc acgettecca gggcccaatg cocgcatete agtttegett teeggcaggg teagggtga gagggaggaa tteteaggtg teaceteete accegeetgg aggeggagge	
	360



ctgcggagtg	catcagcgcg	gtccagccct	ccgcctgccg	ggcgccgagc	gtctccgccg	240
cccggacctg	ggctgggcgc	cgtggcgttg	cctcggagct	cgctgcccgc	ggggcgcgca	300
ccgccttgac	ccgggcggcc	ccgcggcagg	caggcgcccg	cagttccatg	gttggttcgg	360
agcgcgatga	gccgcccgtc	ctccaccggc	cccagcgcta	ataaaccctg	cagcaagcag	420
ccgccgccgc	agccccagca	cactccgtcc	ccggctgcgc	ccccggccgc	cgccaccatc	480
teggetgegg	gccccggctc	gtccgcggtg	cccgccgcgg	cggcggtgat	ctcgggcccc	540
ggcggcggcg	gcgggccggc	ccggtgtcc				569
	o sapiens 3.	.B.30				
<400> 35 geggeegege	tgagctcact	ccgggccctg	cggaaagaat	tcgtaccgtt	cctgttgaac	60
ttcctgaggg	agcagagcag	ccgcgtcctc	ccgcaggggc	ccccgacccc	cgccaagacc	120
ccgggcgcct	cggcagcctt	gccagggagg	ccgggaggcc	cgccgcgggg	tagccgcggg	180
gcgcgcagcc	agcttttccc	tccgaccgag	gccctgagca	ccgctgccga	ggcccctctg	240
gcccgccgcg	ggggcaggag	gcggggcccg	gggccggccc	gcgagcgtgg	aggccgcggc	300
ctggaggagg	gggtcagcgg	ggagagcctg	cccggagccg	ggggccggag	gcttaggggc	360
tctggcagcc	ctagccgccc	cagcctcacg	ctgtctgatc	cgccaaacct	cagcaacctg	420
gaggagttcc	ctcccgtagg	ctcggttccc	cccggcccta	cagggtgaga	ctcagctctc	480
atgcaggaga	tgggtaccac	gaaggctctg	gggagtcagt	cattcgagct	cggcgctccg	540
cagtggagcg	ccaggatggg	tagaaggctg	ggggtgatgg	tgagggtttt	tgtggggttt	600
cttcgcagcg	gccatgctct	gccccgtggg	ccgtcatttt	gtcgtttcgt	tttctctata	660
atgtaataac	taactaggca	aaaagtgtta	aaattaataa	ctactaaata	tccgatgtca	720
ttacaacatt	tataatatat	aacaatatta	aaacatataa	ttaataataa	aaaaaacctt	780
attttaatct	ttttctttt	gttaatttat	atcaccttat	ataccatttt	tctcaatacc	840
attcgataca	atcataaatt	tatttattgt	atattgtcaa	aataaaatat	tcctctatat	900
aaaaataact	ctccta					916

<sup>&</sup>lt;210> 36 <211> 998 <212> DNA <213> Homo sapiens 3.B.36

<sup>&</sup>lt;400> 36



60 gcggccgcgg cgctgttggg ccagcagggc agcaccgagc ccgacttggt gccgcagtac tgcgggggac tgcgggcgc ccagcccgac gggtcggcgt agtagccgag cgggcggcca 120 gtgcagcctg cagcctgcag cggcagcgcc ttcacgcccg ccgccgcgta agagagcagc 180 gtggccgcgt tgcccgcgaa gtccgtggcc gtgtcatagg ccgaggccgc gaagtccagc 240 300 cggttgttgg ccggcgtcac aaaccagcgt tgcggcgagg gcgcgcccgg gtcctcggcc tgctgcggcg acagcagccc gttggtgtgc ggcacgctgc ggtccgtacc cggcccgggg 360 420 cccqcqcccq cgcccgggtg gaagcgggcc ttggcgtagt tgctcacgaa ctggtcctgc aggaaagagc cggccatggc gtagcgggcc ccgggcacga tctgcgagcg cggcgagtcg 480 514 ttgggcgagg gggtcaggcg gtccatgtca cagc

<210> 38

<211> 608 <212> DNA Homo sapiens 3.C.01 <213> <400> 38 gcggccgcgg cgcagcggag gggctgcggg cccggaaccc aggccggtca gcgtgtaagc 60 gccccagccg gccgggctcc gtggggggtc agctccctga cccctacagc gcggtagcgc 120 ctctccgaga gctccgggac cagcggcccg gccgccccca aagccagcct ccctctccct 180 teccegeace gggateceag accagggagg gggegeaegt eegaeggetg aggaatagea 240 gggcgcgagc cggcccggca ggtgcccatc gtcgccctct gggaccccgg tggcgcgctc 300 tgtcctccgc gccacgctca gccaccaccc cggctgtttg ggacccggca cccagccgag 360 cgcgccgccc cctcggggac ccgctgggcg gggctgagcg aggcttggag tgcgggggaa 420 480 gggacgtggg gcgaacccgg ggcgctgcgc cacctcggct gtctccagcg gagaccggcg 540 ccctcgcccc ccgtctccgt tcattgtgct gtattcatcc agcagatttt gaaacaattc 600 togtgtaaaa aggcatttta otoogogogt ottoottaca gocatttagt tgggagtttg 608 cggtgggc <210> 39 <211> 1025 <212> DNA <213> Homo sapiens 3.C.16 <400> 39 gatatecteg etgggegeeg ggggetgeag etegetetge tgetgetget ggtagaagtt 60 ctcctcctcg tcgcagtaga aatmcgsctg caccgagtcg tagtcgaggt catagttcct 120 gttggtgaag ctaacgttga ggggcatcgt cgcgggaggc tgctggagcg gggcacacaa 180 240 agegggagge agtettgagt taaaggggte ttggtgegra aacetggege agegegeagt gcgcgccaca gtcccgaacc tctccccttg cagagctatc ccctaaagcg gctgggtggt 300 cttggtgggg gaataaaggg agcacccttt cacccccttt ggacagtccc ctgctatctc 360 ggagacgcac ttagtgaacc agcggcttgg tgcccgccga gcccccgctc ccccgggagc 420 ccggagcgca aagcccggga gtcggccccg cagcggcaga ggaatcgaaa tcggccctgg 480 cgcccttaag aagccgcggg aggtggcggt gaggaaaaca atttgccaaa atccaaggca 540 caaagttttg cgccacctga aggagaaggc gagaggcgcc tgggcgctag cggctgcgtg 600 aaccccgctc cgcgccgggg cccctccgct gcggctgttc ccactcgcgc cctagccgct 660 720 ctcctaccc cgccggcacc gcagccctc ccaaccttcc ytytccaccg sccccgtccc caccccagt accgccccg tccaacactc cttttgccag cttttcttct ttctctcgcc 780 ggctggagtg gcgagctcag ccgcgggctt taacacccct ccataaatac arggggggtg 840

<212>

DNA

<213> Homo sapiens 3.C.30

tcaaataata ataggggcac ctcccttcgc actcaatacg gagatgcaac tgcgccagag 900 according gatacetece eeggageeac eccaceaagg gtageagetg ttetggaace 960 gcccagagcc ccgctcctcg cagttcctyc gcatctcggg cgcgaggaca cccgagggcg 1020 1025 gccgc <210> 40 1010 <211> <212> DNA Homo sapiens 3.C.17 <220> <221> n (1)..(1010) <222> <223> a or g or c or t <400> 40 60 geggeegegg acegaettee ttegeeggee aceggaggga gggggegeee etaceceggg agggggctgg gcgagccggg agacggtcaa gttggggtcg ggggagcgcg ggcgctccgc 120 actotggggc acgcgggac gagcccggcc gcattgtctg cgcggcctcg gaacaagcac 180 ggccggcggt ggcaccggcg ggcgcgggga ggagttgccg tcccctttcg ccgccgccgc 240 ccaccgcgtt ctttgtgtgt ctctcgccgc cctccagccg cttcgccgct cgcctgacag 300 ctgatgggct caccgcgcg ggtcccgcgt cctctcggcc gcagccggcg gagcccggcc 360 cggcaggagg aggagggag aagaggagcg ttgacagatg ctgtcttgga gcgggcaccg 420 480 ccgggggaaa agtctggact gcctcggcga gaagcggccg gtaggcaacc ggccccagcc 540 tcgcattcgc ctcaaagacc ccaattggct aggagccctt ccctccgcag cggctcgcgc 600 agetecgete ttgegeeceg egeeeggete ageggaegga etagegegee eggteaagaa tcctggggaa cccgctccgc cccctggctc cagcgccctc caatggatgt cggcgtacag 660 aggggctgtt ccgcccaatc aggtgtcggg aagcccagcc agtccccggg gagtgtagcc 720 aatagaaggc gacttcggca cacacccgcc ctgatccact aggacaaacc gctcgagccg 780 gggtggtgga ccgatcctga ggcagatcag ccagtccgcc aaactgtgng caagtagatc 840 900 tgagacggtc cgtgttaatg actatatcta agagntggat gggaacgggg cgcccaattt tccctngtat acgcttttgg caagttgggt tgaaaactga caacctgagc tgttaatgag 960 1010 gcttctttaa ctgtttatgc tatacgccta gtggctcaga caacgttttt <210> 41 <211> 413

lele

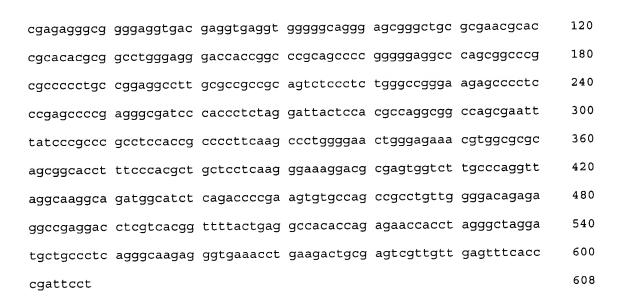
<400> 41 gcggccgccg	taaagcgcgg	atgcgcggcg	tggccacgcc	ccttcagtgc	ttgtgacgca	60
ggcgccctgg	gctttttggg	cgcgaaaaag	aagcagtcct	gggttgtacc	cggcgcagct	120
gggagcggct	gcttcctccg	gggtcgtatc	tccgcccggc	atggggctgc	tggacctttg	180
cgaggaagtg	ttcggcaccg	ccgaccttta	ccgggtgctg	ggcgtgcgac	gcgaggcctc	240
cgacggcgag	gtccgacgag	gctaccacaa	ggtgtccctg	caggtacacc	cggaccgggt	300
gggtgagggc	gacaaggagg	acgccacccg	ccgcttccag	gtatgcaggg	acccgccccg	360
aagacgaccg	gctgcgcggg	cctcccccta	gacttttggc	taccgggccc	cgc	413

<210> 42 <211> 927 <212> DNA <213> Homo sapiens 3.C.35 <220> <221> n <222> (1)..(927)

a or g or c or t

<400> 42 60 geggeegeeg eteettgeet gaeegettge teecegeeeg eeegeeegee gggttgtegg cgcggggcca ctggcgggtc gtgatgagca ctcgctcgcg cccccgcacg cacacgcgaa 120 acceggeeeg geeegeege eegeeeegee tetegeaete eeggageteg eecaeeggee 180 gcgctggctc acactetece teacageaeg eeggeegagg gaggaagggg geggteeggg 240 300 ctcccgaggc gtggggggg ctgtttattt tggggggagg aggggcgcga ggcaggaacg 360 agctgactgg ccgggatcct ccgacccgcc actgtggcag caccgggaag gcggggagag agaaagaggg agggagggag ggaccgggat gtagaactcc agcccgcgcg ggaggctacg 420 gcgagggggg cggtggcggc ccgcgggggg ggcggtgcca ggccccctcg gcaatctccg 480 tagtctcctc gctggctgcc cgagggaggc cgggaagcga tcggggaagc tcgggaatct 540 ccggcacggg cctgggattg tcctggaggc acagcgcggc tggagtgcgg ggcancgcgg 600 ggggggggg gtctgtctcc tttctgggcg gggccgtatc ctgaagcagg cggggcttga 660 gagacccgaa agccacggag tggctcctgc ttgcggtact agttggacag agtaaagtcc 720 tggagttacc tcgcctgagc accctggttt cccgagaggg aatgggcact ctgtgagagg 780 caagctattt gcctgctttc cctccgcaga agaaaaaagg ctcaattgga aggtggagga 840 tgaagccacc ctctatggtc accccaatct gagagcttta ctttatataa ctacattcta 900 927 aggagtagta aaatacccga ggtggaa

<210> 43 <211> 1365 <212> DNA <213> Homo sapiens 3.C.64 <400> 43 gcggccgcaa ggaccggctg agarmtgkgg gscsctgtgc tgggggcgsg arggagrcgg 60 ccytraggac tgcscscccc ccacaccggg gcccgggcgg gacacacgcc caacgggacc 120 180 cctgagcccc caggctgggg accggcaggg gctccgggga ggctggtgag gccaggacgg agccgccycc acgcgtagcc gtgaagcggg aggtacgcgg ccccctggag ctgccccgac 240 tgcagccgag ggcgcgacct gtggtgccaa ccgcctgacc ctgcttggcc gccgccgcct 300 gcgggtctcc agcaggtccc accccacgcg cccgcggggc ccgctccaga ggctcctcca 360 aggccgctgc agaggcgcgg ccaggctccc atttctgcgc atccctggcg ctcagacacg 420 gcctgagccg ggtacccggc gactcccttc ggcctccacc gcctcctggg gagggaccgc 480 gcgctgctcc cacgcgggcc cgggggtctc cgcagccctg gcctgggtgc gtccgtcggg 540 ctgctcggct cggagcaccc cccgccccgc cgccccacca gcgcctytyc ggagcgctca 600 660 ccccgccccc gactcgtgtt gttgttgcgt gggttttttc tctaattctc cggagttact cttttgttgc caattgtttc tatgcccgga ggccacgctg taaatgagat gttacatctg 720 caccgagcta agtaaacact ttataaatga ataaataagt gaataaataa cgaaatcgtc 780 atctcggggc ggcccggctg ccagggctcc ggccgccggc ctgcgggggt ctgtgtggtc 840 900 ccgggccctg ccctggggtc ggggaggcgc cgggaggggc cgtttcccag ccgtgtccct accetgacce catetteett ceteteccaa ateateetee agaetetggg egtttggtee 960 1020 ccagatgtcg tgtgggattc gtggcttcca cccaccgctt ctcaaacaaa aacgggttgt caccgcggct cttaaccctg ggcgagccac ggagcgtttc ttcccgggat cgggatcggg 1080 ccgcggctcg aaccggcatc tgcagaagga agacccggcc ctgtaggccg ccgccgcccc 1140 aggaccggac tggtggcctc tccacgtcgt gtccggaccc gacwcatcgc ctccaacgcs 1200 aacaaacgga agcagcggag cctccgcctc cmasscykgc cyctgyscgs yswgmcmggc 1260 1320 gcattsragt gcwcsakkym sgcycaatym mgagagckct gracktckca aytatcwcgg 1365 actarsrrsr rcawwtkmww argsactcay tgagtaactg atatc <210> 44 <211> 608 <212> DNA Homo sapiens 3.D.21 <213> <400> 44 geggeegeae tgeeetggee geeaegetee gegeetgege egegeaeete aggggeeege 60



<210> 45 <211> 1947 <212> DNA <213> Homo sapiens 3.D.24 <220> <221> n <222> (1)..(1947) <223> a or g or c or t

<400> 45 60 ctgttcccca caggcatcct cctcagtctt acacctttcc accccccaaa acaaatcatt 120 cagcatattt atttcatact gtaatatagg aaatagctat tttttagact ttttatatta 180 ttagcactga tcatacaaac atggaataga aattccttat gttttatctg gatttaaggt 240 300 gatacataat ggaatatatt tctatcaagc cgtacacatt agagataatg aaatcacttg tgttctagtt taaacattat gggaatttca gaactgcaac ataacaaata atcctcggat 360 gaaaactaaa tctctcctct ggtcaggcat ctatgtgcat cagwgatgag aagacaggga 420 ctgtggaagg gaaaacagcg agtcaggaag gactgtggcc acgtccattc cctggtccct 480 caagtaatta aatcctgacc tcctctaccc cagtctgtcc tggggaatgg ccaacactgg 540 cctttcacaa ctgtgtgtta ctagaaatgc aacagaaacc cagctgaatc ccctcctctg 600 cccttctcaa aggaaagatc tgtcccagga ccatttgttc caacattttc aattatgaga 660 actgggaaga taaagttatt tttacattta taaagaaaca catatttatt cacmctcatt 720 wcaagraagg tcaagaatct atmcaaanac caagaggaat ttttaaaaatc ccataatwcc 780 accatcaaaa gagccacact tagcatgttg gtccacaggc ttctttagca ccctcttyyg 840 ttggtgtatg cacaaaatgc acaatcacat tctgtctaca ttttataatt tgcctgtttg 900 ttgattamca ctatatattg aacaattttt aagacctgca acatatgttg acaacattac 960 ttccaaacaa tgtatttaca aataaatgca cacacacat atctgtctta tatacaacgt 1020 gtcttacttt ctaattctcc actcttgaag atttaggttt ttccaacttt ttcttaatat 1080 attcaccagg agtcagcaac ttttttccat aaaaggccaa agagtaggcc gggcgcagtg 1140 gctcacgcct gtaatcccag cattttggga ggccaaggcg ggcagatcac gaggtcagga 1200 1260 gatccagacc atgctggcta acacggtgaa accctgtctc tactaaaaac acaaaaaatt agctgggtgt ggtgagtgtg gcggcggaca cctgtagtcc cagctactcg ggaggctgag 1320 gcaggagaat ggcgtgaacc cgggaggcag aggttgcagt gagccaagat cgcaccactg 1380 1440 cactccagcc tgggcgacag agcaagactc tgtcacaaaa gmaaagaaaa aaaaaaggcc aaagagtaga tattttaaac tctgcaggcc ataggtttct gttgcaacac tcaactctgc 1500 1560 tgttgcaggg aaagaagcca tacacaattt gtaaatgaat gggcatgact gtgttcttcc 1620 cgacatggtt tgccagcccc tgatgtataa cactacagag gatgctgtta gaatgaaawt totttacata tototgatga totoottagg actaattact agacatgaca toatggtago 1680 tgtgggtcaa agggcatgca tgctctggga tgtacattcc cagattgctc atcatgagcc 1740 tttctcatgt caaaatgttt tgtgaccacc agaaaggctg gttctgcttt tawtacccat 1800 ggawtgagga atagaaatga catggcatgg cccttcccca cagcaccacg gcttctcttc 1860 ctcagcacgg cgacaggggc ttcccctttg ccgccgccgc ccgccaagct ccgccgccgc 1920 1947 cggccaagct ccgccgcgcc cgcggcc

```
<210> 46
<211> 1637
<212> DNA
<213> Homo sapiens 3.D.35
<220>
<221> n
<222> (1)..(1637)
```

a or g or c or t

<223>

<400> 46
gatatcttct gataaagaac caatctgcct gggagtttca aatctgaaaa agcaaatcat 60
agtttactgg agtaaactgc tgtttaaaaa taaaagagaa aggaaaaaaa aaagaatgtt 120
tcctagttcc agaactgaca actagagcct aaataaatac ctggacaagg gtaaatatga 180
cctcaaattt ataaccgccc tgaacgcaga acatcaaccg cgacagctgt ggcatcagcg 240
gcgacagtaa ttttctccct ggcattcaac cagagggcag ttggactgtg caccgactgc 300

actagtggtg	ggtagccaaa	gctagcctcc	aaagtgaacc	acggtctggg	gcctggtccc	360
gtttgaccga	aaatgctatc	cagaacmccc	wycgagactg	caggcccttc	ttcctgattg	420
agctagaggt	gagtgaagac	agggtctggg	gtagggaggg	gcgtccacgc	cagcttgccc	480
attacctgcc	ggccttggtg	atgatcatct	cagtgcctat	ctcatgaaag	cgcttccaga	540
gctcggctcc	ctgcagatcc	acccgcgggg	cctgcggcga	gggcagaggg	gtcccgggcc	600
gggccaggga	gncgcgccgg	agaccccttg	ggggaagcct	cccggtgacg	ccagagggga	660
agctccytgc	tggaagccgt	cctcacagcc	gcctggacag	caaaggacag	agaanaggra	720
actggtgagg	gaaaacagag	gggaagcmag	ccgcggagac	ggscccacct	ggtggctgag	780
aagargaaaa	tgaccgggag	aaaaggggaa	gctttggtgc	catcaggtcc	tcctaaagaa	840
caagccagtc	gatagacacc	cacattctgc	ctgtcgaagg	ggcgcattca	gagctccagt	900
gtggcctgct	tggtccccaa	gtcccaagcc	cggrakaggc	gygcggsmag	cgtccacmcc	960
accccgctgk	gcctccgcag	gkcsarggcm	cmasmaraaa	aggcttcacg	ccgnccgccg	1020
gggtctggga	cgcttgcccg	acggagtcag	agragctccc	sggtcmagag	tccacagtgc	1080
aaactycgac	gcaacctgcg	ccttgaarcg	caagcagcaa	aagcgcccsg	cactctgktc	1140
ccaagagcyt	gggcctcctt	aagccataag	cgtytgcggc	gcctcgcttt	kggccttctt	1200
ttgggccggg	ccggaggmat	cttctagaar	gctcttyaga	acmccgcttt	ygycaaactm	1260
ycggncgccc	tgcgcttcca	rcccarcaga	agaaaagtgt	gaaaagcaag	cccgcggtcg	1320
ccgtcggcct	tggcagagaa	atcaagagga	gaagggaagg	gaaccgctca	actacccttc	1380
gggaaaccaa	gtttccaaat	atgccgccct	cttcctggtt	tgcacaaacg	gtttagggca	1440
ttcgttccgg	tttcagggtg	gggtatgccg	tcgctcccct	cctccccgcc	ctgtgctttt	1500
aaaagttagg	aaacaaaaa	gagcacccat	tggctggaac	cccaagggag	gcagatgcag	1560
gaagcacaga	gctgcaccgc	taggcgcagc	aaacagccgc	ggccgaaggc	gcgggtcgcc	1620
gagtgggcgg	cggccgc					1637

```
<210> 47

<211> 900

<212> DNA

<213> Homo sapiens 3.D.40

<220>

<221> n

<222> (1)..(900)

<223> a or g or c or t
```

<400> 47 geggeegegg ceeggaceag cegeteecac eegeeeeage tactaeggeg eggegegace 6 gegggeteeg geeceageee aggeaegtge geecaggeeg eggggaggeg eeggegeete 120 ccggaacgcg ctcctggcct gcgagtgctg cccgctcagt ctccgggtgg gaagtgcgct 180 240 cqccccqqac cqagqggaaa gcccaacatc cccgggatgg aacagagagg cggccacccg 300 tgagtgggcg tgacccattg gttcccttgc gcagcatctg tggagaatta ggctttcccc 360 tcctctcttq ccaqccqttg ttcctaatct tgtctttttt aagggaggaa agcaggagaa ctcatgacac tttgtatcac aggaaatcaa gttggtggag agagggtttg ctgacctctc 420 ccgtcccttc tcagggtccc taggagaatt tttgaagaag taatcggcag caaggagatg 480 ggggcaatag agagteteag actegeaggg acceatgtte gteeceageg ceactaettt 540 caaaccgtta tccctcagag ctgtttcctc acctccacaa caactctccc gggttcgatg 600 660 acactatata tcccaccagt tcatcttggt acaggccaaa aggtaattca aaaagcgaaa cgaatctcat nttctgacct gtgccctcgg taaagtcccc angtttccac cccaagtaca 720 cttggaagcc aggcccctnc acacangctg ancaccacct tncacaaact gaaaacaaag 780 anaatccctt ggtttcaaag ttagaatagg gatacngcgt gagtggggtg aattgcnatt 840 gggtcaagga aaaaaaaaa gtaaatnaat taanttttnt tgacctcctg cgctgcccac 900 <210> 48 <211> 1511 <212> DNA Homo sapiens 3.D.44 <213> <400> 48 60 cgggcgcggc gagccccact ttctcccggc aggaaggggg gaggccgaga gcatttcctg ttgtgcagct gagccctgcg gagacgtcat tgcattcatg ctccctcggg tgtcagcgga 120 180 cggggggcca aagttcaagc cgcgtccagg gcaggcagcg cgcggcggcg cgggggcgcg gggcgggcgg ccagggctcc cctctcccgc tggcgctccc ggcgcctccg tccccggccg 240 gcccagcgct gctaccggag gccagccctg gggctccgcg gggaagagct gctcttcctc 300 ccggaggaaa ccgagctcgc aagcccagcg ctcccagccg cagactgcag agctccagta 360 aggtgaaagt aggcaagaag gccccctgag acgtttctaa aagcatattc tatatgtttt 420 cattatgaaa acacccactg cactcctttt atttattagg accttaagtt atcctatctc 480 aactaatact tttaacaatc agaatctctt aagaatcttt caatcttata cttatccact 540 ttaatagcca acaaaacctt tagccagagt gttttaaaat ggaaattacc tgttcatgtt 600 tottaaagat tittaaagto toottotaaa titooagoot tooatitagt tioaagooat 660 aaaccagatt ataacaatgt gtaattgtag agaagctgtg gcttacggtt aataacgatt 720 aaaaataagg ccataaggta ttttatgatc attttgaaat aaaaaattga aatagtttaa 780

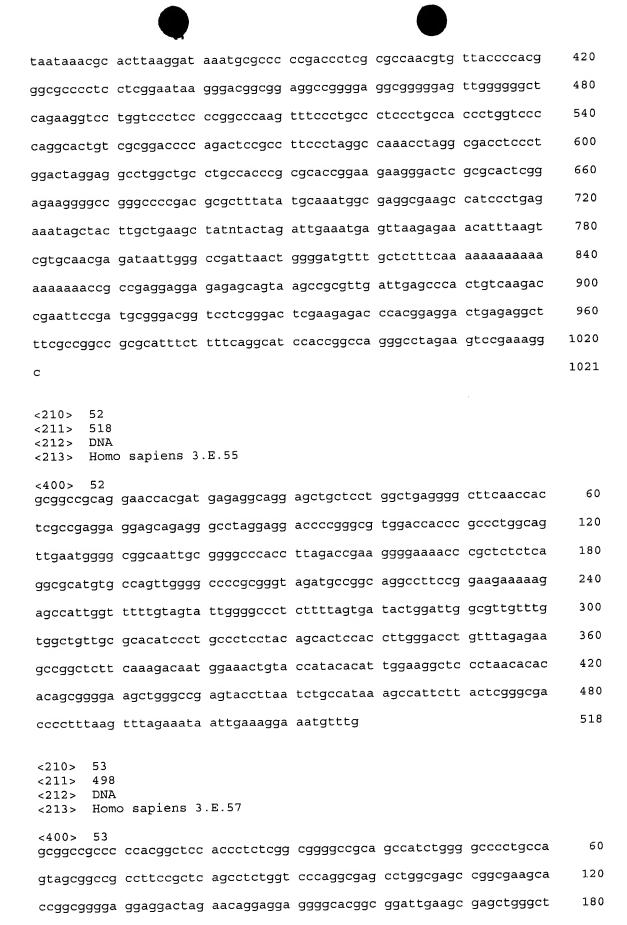
840 tttcaqcttq tqcaqtttqa gacagatcgt caactacaaa acaaattgta gattctgttc tcatggtgaa caaacattac agatgtttta ctgtgtcaac atctctaaca tttgaactaa 900 gcaatgtttc acatcagaac atgaattaaa acaatgtaaa ctatggacct ggggtgacca 960 tgatgtgtcg atgtaggttc ttggattata acaaatgtac cactctagcg caagacttcg 1020 atagtggagg aggctgtgtg tatgtgggga caggaagtac atgggaaatc tctgtacctt 1080 ccgctggatt ttgctgagaa gctaaaacta ccctaaaaat ataaactcta tttttaaaca 1140 1200 aattgacaag aaattaatct taagaattgg cacagaaatc atctcgatgt tttcatgaag 1260 ttcatcctcg gttctactgc ttcttgataa acaagtttca tgtttagaag gttactgaaa 1320 tttttttata tggtaaaggc acatcaaaga ctttaccatt taatatata tagttgtcct 1380 1440 atccagtcat gtactattta aggcaatatt aaaggtaact tagatttccc cacttacagt 1500 gatgcaaagc ccttcaataa tattctgttg tcttatttcc taaacatctg aataatacaa 1511 ctttatcaca t

<210> 49
<211> 835
<212> DNA
<213> Homo sapiens 3.D.60

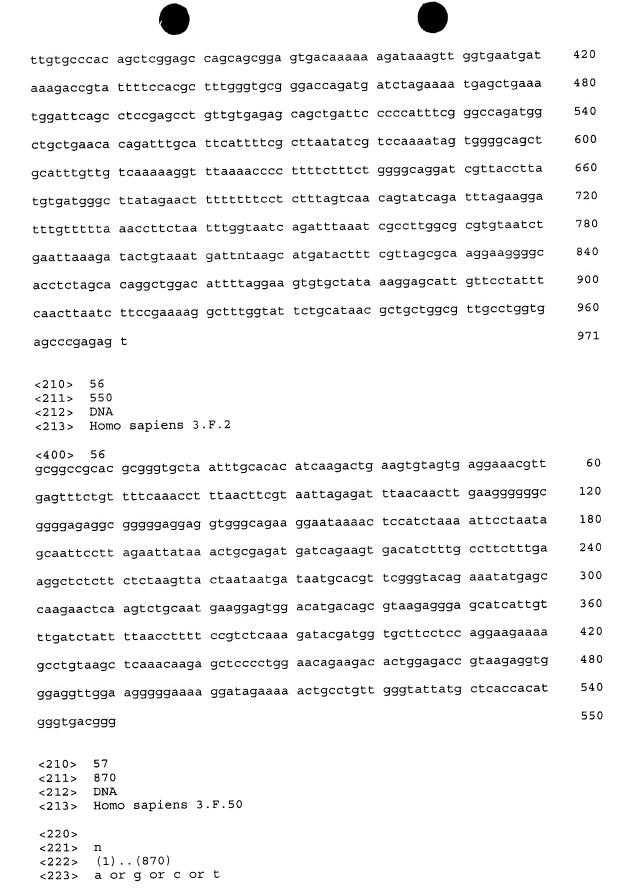
<220>
<221> n
<222> (1)..(835)
<223> a or g or c or t

<400> 49 geggeegee eggageegge gteegeageg getgegeate tegggeetge ageggggege 60 ttggcgggcg ggggccgggg gagagcctgt ttgcgcagta cccccggagg gcggaaggcc 120 gccgaggtaa gagccgggac tcggccaggt gggagtgggc accttgggcc gggcctgcag 180 ggcggtcccc gagcgtcccg gggtagggtg ggctccctgg ggacgatgcc cagggccccg 240 geogegetee ggtegegee caceeegget geagegege ettggggege tgetggeete 300 gccgcggggg tgggagcggt cgcggcctgg agcagctccg ggcgggcccc aggctctggg 360 gccagggcca gctgcgcgca ggggtgagtg agcagccccc gggccctcaa gtgagcccct 420 gtccgctccc caccttgcat ttctcctctc cgcagtgggc gtggcgcccc tttgctgtat 480 agggggcgcc ccaaattgaa gaaggctggg ggggagaacg cataaacagg tgtttagggg 540 gcccaggcct gtgcgccaag ggttgaagaa taaagagtaa ttctttttc cccctttta 600 agggggnccg gagtccccct ccccccggc cgtggtaagg gccccccctt gctccgtaag 660

gggccctcct	ttggnaaaac	aactcctttt	ttctttttt	attttgtccc	ccccnccca	. 720
ataatttaaa	nncctccctg	ntcgcccccg	ccccccgctt	tttttttt	tttttctnaa	780
acccccacc	ccccccccc	cccttnnttt	gtttccgctt	ttattccaag	aaaat	835
<210> 50 <211> 645 <212> DNA <213> Homo	o sapiens 3.	E.04				
<400> 50 gcggccgccg	gcttgacgtg	tacggcgctg	atcacctacg	cttgctgggg	gcagctgccg	60
ccgctgccct	gggcgtcgcc	aaccccgtcg	cgaccggtgg	gcgtgctgct	gtggtgggag	120
cccttcgggg	ggcgcgatag	cgccccgagg	ccgccccctg	actgccggct	gcgcttcaac	180
atcagcggct	gccgcctgct	caccgaccgc	gcgtcctacg	gagaggctca	ggccgtgctt	240
ttccaccacc	gcgacctcgt	gaaggggccc	cccgactggc	ccccgccctg	gggcatccag	300
gcgcacactg	ccgaggaggt	ggatctgcgc	gtgttggact	acgaggaggc	agcggcggcg	360
gcagaagccc	tggcgacctc	cagccccagg	cccccgggcc	agcgctgggt	ttggatgaac	420
ttcgagtcgc	cctcgcactc	cccggggctg	cgaagcctgc	aagtaacctc	ttcaactgga	480
cgctctccta	ccgggcggac	tcggacgtct	ttgtgcctta	tggctacctc	taccccagaa	540
gccaccccgg	cgacccgcct	cagcctggcc	ccgcactgtc	caggaaacaa	gggctggtgg	600
catgggtggt	gagccacttg	ggacgagcgc	caggcccggg	tccgt		645
<210> 51 <211> 102 <212> DNA <213> Hom		.E.50				
<220> <221> n <222> (1) <223> a o	(1021) rg or c or	t				
<400> 51 gcggccgcgg	gacggggaga	tgeggeeeeg	gtattgatgt	cgaaaatgat	ggataacgcg	60
ggaatggcaa	atatactatt	tgtctaatgg	ctcggcaatt	aaattcccct	gtaaatgacc	120
catgcctcat	ttcatcctaa	tctatggaat	tttgattgaa	ttcgtcagct	ctaattgaaa	180
aatactgcac	tttaatgtct	gcattgcagt	ttcaggacga	gagtggtttt	aatgagacag	240
tgccccatg	g acccgggaat	atttgagact	tttattcgga	atttaaagcc	aggagattgc	300
togactgago	cctgagattt	cctctcctqt	atccacqtcc	atccatctcc	agacgcgatt	360



gtgagcaagg	gacacccaca	gcctggagaa	acagccccgc	tctcttgcgc	gctgtctgct	240
ccagccgcta	ctgggggctc	taagcagcgc	gatgctgctt	cgcttcttct	aggcggcggc	300
cggcggaggc	tttccgcagc	cgcttggccg	gcgccggccc	ctattccgtt	ggcaagtccc	360
ttgtctatcc	cggagggcgc	acccggacgc	tcgagccgga	gcgagcgcga	agtccgaagt	420
ccgcccccag	agccgccaac	ttccctgtga	gcccctctcc	ccgccgcagc	ctgcgccaga	480
cctgggagcg	atgcgccc					498
<210> 54 <211> 471 <212> DNA <213> Home	o sapiens 3	E.59				
<400> 54 gcggccgccc	gggcccgcgg	gcggggggat	cggcgggggg	gacccgcggg	gtgaccggcg	60
gcaggagccg	ccaccatgga	gttccgccag	gaggagtttc	ggaagctagc	gggtcgtgct	120
ctcgggaagc	tgcaccggtg	agcctggcgg	gggtcccggg	agaagagtgg	gaggatctga	180
ggaggatgct	aattcccacc	tgggcgcaga	ctgacagatg	aacgggcgat	accccggcat	240
gggggtccac	ccatctgtcc	agttttctgc	cgtgggctcc	gacggcgctg	ttctccctgg	300
tcgagccttg	tccattatcc	tgttcctttt	tctgcacccc	accccacccg	gctccactct	360
ctctggtgct	gtaaatgcct	ctctcccggg	tctctggctc	ctcccccacc	acttctgggt	420
ctctgtcccc	gtctctttct	ggatgtctct	gccccttttc	tctctgggtc	t	471
<220> <221> n <222> (1)	o sapiens 3					
<223> a c	orgorcor	t				
<400> 55 gcggccgccg	tgggcctgca	aaacttccaa	agtagcagco	tgtttctcct	cgtctccctt	60
ctcctgggta	cccagcgccc	cgccttcccc	agaaagggcg	aggggtgggg	gcagggctcc	120
ctcgggaggt	ggccaagcgc	cgggacgcgc	tcccagcgtt	actcaggaca	cttgggattt	180
ggcctgcago	cccttcccc	atccctggcc	tggctgcggt	gtcccttgct	cccctctgct	240
gctgctcctg	g ccccatcaag	tcgaaaatct	gagggtggga	tggggtgggg	gaccaggggg	300
taccctccca	ggccgctccg	cagcaggccg	aggtggagac	cctgcccggt	aggcgagtcc	360



<400> 57						
ttagactctc	actgggcagg	tctgctgtcc	cctctgctcc	cgcaggactg	gagccaccga	60
gctcgcgcct	tcttctcggg	gtgcgatttc	tctcctcttt	tggactcaag	atcaatgctt	120
cccggccggc	gcagatcaca	cagcaggacc	ccaggggaga	ctgtggcctt	cttcccgcct	180
cccaattccc	caagaccgcc	tctagaggct	gctgtgtccg	gagaactccg	agcattttct	240
ggacacagat	tgcctaacag	aggaacaggg	gttagg <b>t</b> ggg	gagcggctgg	ccggcccaaa	300
cacagcagcc	ccaagctggc	tcccaagcct	gggctctcca	ccccgctcc	catcctctct	360
tgagcacagt	taggcccaac	acccctgtcc	cccaaaacac	ctcctaccct	ccctccccc	420
cagcccccat	cttcaggaac	atcacagggc	tcacactcac	taaccgcgga	gagcacatgc	480
aggccggagc	cctcagcccg	gcagctctcg	gaccctgccc	agctcgacgc	ggactcatgc	540
agaagaggac	attccgcagg	taggtacaat	cccagcgctg	gggcctgggg	cgtccggggg	600
gcggcctttg	agcttccccg	ataccgctcg	cctgctcccg	gagctgttcg	gccgacggct	660
gcccggntcg	tgcactttca	gtanggcccc	gctgactcta	ctgcccttgg	gctaggccta	720
ccggngatgc	ccagactcct	tgggacgctg	gacccgcngc	gcgggcggac	acgcanngac	780
teegetetne	gcccggaatc	gttgagacgg	aatctcagcg	gatcccgcgt	cgccgagcgc	840
cgggncaggg	agaaaggccg	tgtggcgctn				870
<210> 58 <211> 848 <212> DNA <213> Hom		.F.72				
<400> 58 geggeegeeg	cgtcgccgac	gcccggcagg	actgagcgca	cggagcggcg	gaactcctcg	60
ttcctccacg	tgtagagcag	cggattgagc	gcggacaggg	cgcagcacag	gagccagctg	120
gccgcctgca	ctccccaggg	caccggcagc	gagaagccgc	tggccaggct	cacccacacc	180
agtggctgcg	tggccagcag	gaagacgcag	cagagcagca	gcaccgacag	gccgctgaga	240
cgccgctgtg	cccgccgcgg	gtgcagcgcg	ggcggcaggg	gctgggcctg	cgccgggtgc	300
gcggcgccac	cggggcccgg	cgcgtgctgg	gcgcccggga	aggcggcggc	ggcggcggcg	360
caaccgggca	actggtgcaa	catgtggaag	ttgatcacgc	tgacccactt	gacacttaca	420
cacacgcago	gcacgatgcc	caaatagcat	tgcaacaaca	tatctgtctg	ctccaacaac	480
accacagaag	ccatcaacac	cagataatgg	attctcagtg	gcacagcacc	cagccccagt	540
gcccaaagcg	g agaacaacat	cactaggccc	aaggccagct	cccaaaacca	caccaacatc	600
ccccctagt	gcacctttta	tacaacaccc	tgtaaataga	caacccccca	ataataacca	660

acacaccacc ccctaaaaaa aaaataattt tcgccaatac cgtcccaatt tttaaaaaat 780 ttcccaaaaa cctctaatcc aaaaacccca accccgcctt cttctatatt tcaaaaaata 840 848 cccaaact <210> 59 <211> 2770 <212> DNA Homo sapiens 3.F.82 <220> <221> n (1)...(2770)<222> <223> a or g or c or t <400> 59 atccanatat tttnnaacct ctaacaatga agagtannac acanactcaa ttttanaagg 60 cacaggacct atgaanacat tttatggtaa aagaaataca aatggccatt tcccacgtna 120 agatgcatct aacctcaatg gtggtcacag naaaataaat tacaaaaaan aaagttttgt 180 gtgaccatca gttaggnnaa ttaaatgctt cctactaatc ttttcatgat aagtannaac 240 300 atactagcca ggcatggtgg ctcatgcctg tattctcagc atgttgggaa gctgaggcag aaggatacct taagctcagg agtttgaggc tacaatgagc tatgatcatg cactccagcc 360 tgggtaacag agagtgagac cctgtttcta aataaataaa taaatgagtg catgagtgaa 420 480 catacataca tacatataca cacacggttt tttacatgtt tatagagagt ataaatggcc aatgaccttt taaggcacaa ttagcaaata tgtattgagt ggaaagatgc atgttcttgc 540 600 atquaggatt ctacctcctg anatgcatct gataacactg cttgaaaatg tgtgtagaaa tgcccacact agcatgtttg tggtgggcat ataaataata gcaaaacaaa acaaaggaaa 660 aagaaaagta catatatgtg aggaaccctt ttggttatcc tgggtttttg agataatgtt 720 catagaagga aagcaaatca aatgaagagc aattgagcag gaaacggggg gaaataccct 780 cagagtaata agattatete attacaetta agttttgetg atgetteaag ttteetgagt 840 aagttatgcg aagcatcttt ctctgaaaat cttcttgctg cagaacaaac catgtttagt 900 960 gtctgtatat gtctcaactt cctgtcccca cctggcggat gggaaaaagg acacggtcct 1020 tgcttgtgtt ttggagtgaa agaagcatta aaggtcttgc agactttacc aaggattctc ctggtctcat ttcagatcca acttccaact ccaggcagcc tctgtgtttt tctttaatgt 1080 ataatcagga tgtacttcaa tttggactct attgctgttt ggcctgtata tgcagtttca 1140 agatagcccc atacacctgc ctgcaatgat ccttcaggaa tagaatgggc ttctgagttg 1200 aggaatttgg gagtatactg agccctttgt gtatttttat taagtttctc tattcatgcc 1260

1320 aggagaaggc tgtggacaaa aagtaaagga ggagacactg gaattgtgat gtccaaagat 1380 tccaatgttc aaggattatt tgaaccettc acgcetettt agccaccgcc gccgacagcg 1440 aagacgcgga gaaaaaagtt ctcgccacca aagtccttgg cactgtcaaa tgggtcaacg 1500 tcagaaatgg atatggattt ataaatcgaa atgacaccaa agaagatcta tttatacatc 1560 agactgccat caagaagaat aacccacaga aatatctgcg cagtgtagga gatggagaaa 1620 ctgtagagtt tgatgtggtt taaggagaga agggtgcaga agcagccagt gtgactggcc 1680 ggggtggagt tcctgtggag ggcagtcgtt acgcgctgat tggcgccgtt acagacgtgg 1740 ctactatgga aagcgccatg gccctccccg ggattacgct gggaggagga ggaagaaggg ageggeagea gtgaaggatt tgaccccct accaetgata ggeagttete tggggeeegg 1800 aatcggctgc gccgccccca gtatcgcccc cagtacaggc agcagcggtt cccgccttac 1860 cacgtgggac agacgtttga ccgtcgctca ccggtcttac cccatcccaa cagaatacag 1920 1980 gctgttgaga ttggagagct gaaggatgga gtcccagaag gagcacaact tcagggacca tttcatcgaa atccaactta ccgcccaagg taccatagca ggggacctcc tcgcccacga 2040 2100 cctgccccag cagttggaga ggctgaagat aaagaaaatc agcaagcctc cagtggtcca aaccagccgc ctgttcgccg tggataccgg cgtccctaca attaccggcg tcgcccacgt 2160 tctcctaacg ctccttcaca agatggcaaa gaggccacgg caggtgaagc accaactgag 2220 aaccctgctc catccaccga gcagagcagt gctgagtaac accaggctcc ccaggcacct 2280 tcaccatcgg cagggtgacc taaagaatta atgaccgttc agaaacaaag caaaaagcag 2340 2400 qccacaqcct taccaacacc aaagaaacat ccaagcaata aagtggaaga cgaaccaaga 2460 tttggacatt ggaatgtttg ctgttattct ttaagaaaca actacaaaaa gaaaatgtca 2520 acaaattttt ccagcaaact gagaacctgg gaattcctgc acagaagaca agagagcagc ctccccagtt tcagcaagcg ctaggtttat atttttttcc tggtttttac tgtttgggta 2580 atagatattg aaacaagtaa tattaatacc gcatggggag aaccccaacc aaagaaatct 2640 gaaatataaa ataaatgctt ttttttccgt ttttgttcat tttggatgct ggcgctaagc 2700 ctccaagtgt catgattaaa aaaaaaatta tgtccttatt tatttctagg atgaggggag 2760 2770 gataacattt

60

<sup>&</sup>lt;210> 60

<sup>&</sup>lt;211> 563

<sup>&</sup>lt;212> DNA

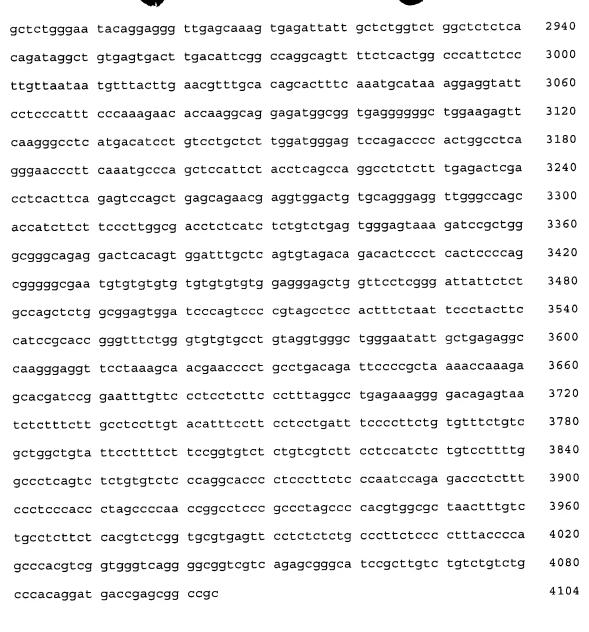
<sup>&</sup>lt;213> Homo sapiens 3.G.46

<sup>&</sup>lt;400> 60

geggeegeeg eetteegeag taatggttgt teagegaaca agateeggge ggaaacagta

gataggcggg tgcagcgggg cagaacatag gttgccttag agaggttccc cggtgtcccg 120 acggcggctc aagtcagagt tgctgggttt tgctcagatt ggtgtgggaa gagcctgcct 180 240 gtggggagcg gccactccat actgctgagg cctcaggact gctgctcagc ttgcccgtta cctgaagagg cggcggagcc gggcccctga ccggtcacca tgtgggcctt ctcggaattg 300 cccatgccgc tgctgatcaa tttgatcgtc tcgctgctgg gatttgtggc cacagtcacc 360 ctcatcccgg ccttccgggg ccacttcatt gctgcgcgcc tctgtggtca ggacctcaac 420 480 aaaaccagcc gacagcagat gtgagcagcg gcacacgggt ccgggcaggg ggcaagggct aaggaaggag tggctagggc aggggcggga accggggtgc ttgaccacac gtgaagactc 540 563 agaactaacc caggcagcct gga <210> 61 <211> 4104 <212> DNA Homo sapiens 3.G.78 <213> <400> 61 gatatetete tecaageece etteccaact ceatttetgt aggaaagtae ageecetgga 60 attgggttct ggtttcgctt tgggctggag gtgggtggat gggggtcaga gagagaatga 120 ggtgggggg acttcaaggt tctgtcccac cgaccagagt ctgaagacta ttcgcctttc 180 ccaacacgga cctccgccca tccaggcccg ggactatece ttcgcggtgt ageggcagec 240 ggagacctgg ctgaggaggc aaccgcgtag acacctccct gcttagaaaa caaacactga 300 accagaccga tcccagttgg agggttcgaa aatgttccag acagcctgtc gggaggggtt 360 gttgttgctg ttggactaaa tagctattcc tgattggtca tgtatagggt tttttaaggc 420 480 gggtgggggg aggaggggt agaggaaagg ctccaaacac ctgcaggttg ggggcggaaa 540 gctgtttgcg attccctgga ctggttggtc ggggacagga ggtaattccc agccattgac ccccatttct ctctctccct ccctcttgcc ctgcctcttt ctctccaccc ctatctttcc 600 660 tggaaactcg ctttgggcgc ggcagatcgc ccaggaccac accgcagcgt aactgcaggc ctctcagcga aaaaggggga aagcaaagac ccgggtgtgc atcctcttcc tcggcttccg 720 780 cccctttccg gcggagtgga gatcctattc agaggggccg gtctctctaa atatgcccca 840 ggtgagtttt caggggaatg gtgccggtgg aaacggtgtc taggaaggcc ttgtgttccg gcctggggtg aggaaggctc aggacagagg agagcccatt ctcagattgg gggtggggg 900 960 aggggaggac cagccagagc ttggaatcgg gatctgactg ctgtagctgc ctctgtggca 1020 ttcagegget tttteeettt tecacecagg gtaaaaecag etagttggae ttagtegtee aggeetttee cattggteee ggttetgtgg aegttteeea aggeeggtaa etttggggeg 1080

1140 gctgtatccg ggtggtacag actgtgcctg gagctcccgc aggaggaagg cggcagcctt cctggctagt gcagtcccag ctcgagtggg ccctgatccc aggcctgagg cctagggtgg 1200 ggaggcagga acacccctct tctccggtag aggcgaggat ggtggtgctg ttccctggtg 1260 1320 ggtttggtac ttgtgcaggc ttggggcttc tccagggtgt tgtgctggtg tgggcccaga agagagacca gaggctgggt ctaagggcct gaggctgttt tcatctaaga aattctctgt 1380 1440 atgggggatt gggtctgctt gagacctgtc cccaggaaga atctcctggg gtcttctgtc 1500 ttgttctggc acaggtggaa atattctggc tgtctggcaa ctgcagatga ggatttcctg ttgggggcta taagcagggt ctccgtagta caaagagaga ggagctgtag tcgtcaaata 1560 1620 ctctagaacg attcagtcta aaatctccct cctccttcat tctccccaaa taaaaacaaa 1680 caaaatctct cgggcgttcc tttctgtaat ccaaatcaag tgatgcagct tagtcgccaa 1740 caaccatcag tgtttgtgag tggcttcttt ggggcatgga cctctggctg gtaatcctaa 1800 accggcagga ttttcctaaa atgtggggag gagccgggag aggtcctcca cagatcctgg gatccaatca tatatttctt acaaggaacc ttggcgatgg gatatttata ggtgtctgga 1860 gaggacattt gtggccaggg tcaattcatc tggaatatgt actcccattg cctctcagga 1920 atccaccgct agagcaggag cctaagaatt aattggaggg taaaaatgtg tcataacaga 1980 gcttgagctc agtctgcaac tgcagtgcac actgtcactc ggttagaagc tggggcttaa 2040 gcatggatca ctgggctcac accggtgtgt caggacggag agcagtgagg tagggaacca 2100 2160 ataccttgaa gcttgtatgt ttcccagggg ttggtatatt tctggcacat ttcgctgctg 2220 ctgggagcaa gaggacctgg ctgatatact tctggtgcat ttccagtggc cttggtgtct tggtggttgc attctatgga tagagaccta ttgtctccac caaaatcata aactcacttc 2280 2340 caatgaagtg tcagggacct actgccttta cagcttgtat acaccaggac ttagggaatt ttgtggtttc tgtgccagac ctggggggct ggcattccca aagaaggtgt acagcagtct 2400 gaatettgae tetetgteat eetgggtgte tagtggeaat tgageeaage teeagaggag 2460 2520 gctgcagatg atccattctc ccttctgggg tgggagggat ggttcctagg atgactcctg tccagagcat tgcagtggca gtatgggagc tcaatggctg ctatgtatga tttagatgga 2580 2640 ctctgcatgg gggtaaattg tttttttgta tttgttttct tcttttaaat acccaattat ataattcaga gagcagaaag cttattttaa acaacttatg tggtgttgat catatatgta 2700 2760 caactcacaa ctcacaaact ctggcccttg agtctcctga tttttctgtt ttggttcttg 2820 ctggtgccca gctctatctg gatgaagcca ggtgatggaa gagccccagc acacctgtgg gaagtagagt ggctgtggtc atctcggagt atgcttgtgg ggtcacaagg tggtttcact 2880



<sup>&</sup>lt;210> 62

<sup>&</sup>lt;211> 570

<sup>&</sup>lt;212> DNA

<sup>&</sup>lt;213> Homo sapiens 4.B.44

<sup>&</sup>lt;400> 62
gcggccgcct gtctgggcgc cgcgctcctg ctcctatgcg ccgcgccccg ctccctgcgc 60

ccgggtgagt gcccgccggc cgagccgcgc acccccaacc aaacctggct cctcgcgctt 120

tccaccgcgg cctgacccct cgacagcgcg ggggacacct gttgtctcct tcctggctgg 180

ggctaggggt ggcgggcagg ggcgctggtg cggcacagaa aggctctaga cgcccccgcg 240

agcaaaggct cttgctcctc ctccggagtt acctccccac tcccagagcg gtgactgtt 300

tgagtcccac agccggtgcc tggagaccgg ggtcagttgt gggggtaga ggacaattgg 360

ccaatccggg	aaggccatct	cccttacctt	cacccccttc	ccctgcgcac	cccacggccc	420
ctggacatga	gcgctgctgg	gcgcatgcgc	ataggagggg	aagcttgggc	cactcggtcc	480
ggtcccttgg	ttgtcctact	gtgcagtggg	tgccactccc	tgctccaccc	tgaaatccac	540
actgggtagg	gcttgggact	cctgtgcacc				570
<210> 63 <211> 535 <212> DNA <213> Hom	o sapiens 4.	В.56				
<400> 63 gcggccgcgc	tttctccatg	gccccggcct	cggcgcgctc	ggctccggct	cgggggtccg	60
gcacggcagt	ctcagtgcgc	ggtcgccagg	cgcgccgtcc	caccccggct	cggcttgggg	120
gtggccccgc	gcctccgccg	ccgacgcagc	tagctggttt	ttaaattgct	aatctcatta	180
acggcgcgcc	cgtccgagag	gcgaggctgg	taaatggatg	acggcgagcc	ccaccccgcc	240
cgatcgtcgc	ggccgggaag	gcacccgaga	ttgcagagga	cagggcggag	tcccctgggg	300
tcctccggct	cggcggggcc	tttcttcagg	ctgcggaact	cctcgaagtg	ggcgccttcc	360
ctcggccact	cacctgtcat	ttatcgagcg	cctactgtgt	gccaggcatt	gtctggggac	420
acggctgtga	accacttccc	agctccgtct	tggagctgac	attctggtag	agggaaacac	480
ttgaattgga	ctgcatgaaa	tgccccattt	tcaaccattt	tttaatttat	agaaa	535
<210> 64 <211> 737 <212> DNA <213> Hom		.C.05				
<400> 64 geggeegee	ggcggggtta	aggcctctca	gccaaggccg	cggccagctc	actgccaggt	60
cgggtcagcg	cctgcgcgcc	aggtccggcc	ttggataccc	tctgccgcca	cgcgtcggtc	120
cggcctctac	gecegeetgg	ccctctgcgc	gcgccgccga	cgccgcaggt	ccgggcctcg	180
gtgactgccg	gaggggcgcg	gcgccccgcc	tcctgtcacc	atggccaccg	caaccccttc	240
caccgcctca	cggccggccg	gcatccaatc	acaggcgagc	gttaccgatg	ccggggcggg	300
gcaagacagg	gagaggaagt	cccggaaggg	agtgcggagg	gatgcggcgc	ttcggcgagc	360
acccgttgtg	g tgggaactcc	gtctcaagtc	gcccccattg	tacggatgaa	ggaatcgaag	420
ccacgagcca	gaatttcctc	actcgcaact	cgagaataaa	ttgcgcctcc	ctgagtgtgg	480
aggattaaat	aagtagttta	aggcgtgttt	aaagagcgct	tgtaagttgc	caagtcgctg	540
gagagccagt	cccttatccc	ttgaaccagg	tgatgctgac	gtctgatttc	aagacagttc	600
chacccchc	r togaaggaaa	gececatege	aagaaqtcqa	tgtcctgtaa	tttacgttat	660

aatcttcgca	tcataaagat	tactcggcag	taattggttt	cttgactaat	tataccagat	720
gagaattgaa	gactatt					737
<210> 65 <211> 684 <212> DNA <213> Homo	o sapiens 4.	C.25				
<400> 65 gcggccgcca	taggaaacac	ctggcagtta	gttcctcaaa	aggttaagcc	cagaactccc	60
gtaagaaccc	gcaattccac	tccttagtat	agacccgaga	gaaaacatgc	gtccgtccac	120
gcaaaaatct	gcacacgaat	gttcacagaa	gcatcaggca	taacagtcga	aatgtagaga	180
caacccaaat	gtccatatgg	atgaactaac	tgtggtccat	ccatgaccgt	aatggaacac	240
gaccataacc	aggtgtgaag	ttcagctgtg	acagggatga	ccctcgaaca	cggcacgctt	300
ggtaaaacaa	gcccgatgca	gaacagcacg	attctattta	tgcgcctgcc	cacaagaggc	360
acaccccggg	aaagaaagca	gatcagcact	tcccaggaac	cgggacgcag	ggacgcaggg	420
agggagggac	tgctgaagat	gcacggcgtt	tcttttggga	tgaagaacag	gttctaaaat	480
cgactgtggt	gatggctgcg	taaatcagtg	aatacactaa	aaaccttact	gaactgtata	540
ttatttattt	atttattgaa	acagagtctc	gctttctcgc	ccaggctgga	gggcaatcgc	600
accatctcgg	ctcactgcaa	ccttcgcctc	ccgggttcaa	gggattctcc	tgcctctgcc	660
tcccgagtag	ctgggactac	aagc				684
<220> <221> n <222> (1)						
<400> 66 gcggccgcgg	cggcagcggc	tgcggggagc	tccagcagcg	gcggcggcgg	cggcggcggc	60
agcggcagcg	gcagcagcag	cagcgacacg	tccagcaccg	gcgaggagga	aaggatgcgg	120
cgcctcttcc	agacgtgcga	cggcgacggg	gacggataca	tcagcaggta	cgcggggagg	180
tacgaggaaa	ccgacaggag	cgagatcagt	ccctccgcgc	gcccttgacc	cctgctctgc	240
cccctcgccc	caacttgcgg	caagttgctc	agaagctcgc	gggaaaagtt	ggccgcgact	300
ccaaaaacac	atageegget	cggccacgaa	ggccgaggg	actgctctgt	tcgccttgcg	360

ggggtgccag	ttggtccaac	ttttcccagc	gctgtctttg	tctaggcgtt	gggagacatc	420
tccttaggat	gcgcactctt	ccgggggctc	ggagtgttct	tccctgtggg	aaaaggagtt	480
ctggccgctt	gtcccaggta	ggaggggctg	ccccacagcc	tcggggtcct	gggcatcaag	540
atgccgcagc	acggggcagc	gatctgcccg	gcggcttggt	ggacacccca	gggccgcacc	600
gggaggagat	gagctaagcg	acagcctcgg	acagggaaat	aacctgtgaa	gaaactttct	660
tgtgccgcag	aacccatgaa	ttccaaactt	cagagcccaa	gaatgggtat	cgtttgccac	720
ccagtattga	tttaaacgca	gtagcctgag	aggaacgaag	cgctcaggag	caaactaggg	780
ctagacccga	ctnctacccg	gctctgtgcg	ctgaccaggt	gagcttcggc	gtggttccgg	840
gcgcctcgng	cctcactaca	acaacttttg	ggtgttgctt	cgatccccga	cttctacaga	900
gcngattaag	cttctgctcc	ngctgncaat	atactctgcc	aattggacta	acttgngtga	960
gaagatccac	ttctgatgct	ttgatgtgca	cgctgaatgg	ttccngatga	tg	1012
	o sapiens 4	.C.9				
<400> 67 gcggccgcct	tgaaggcgct	ggacgggatg	gtgctgaagt	cggtgaagga	gccccggcag	60
gtgagctcgc	ggcccgccag	cccgctgccc	acgcagtagt	ggaagaggcc	gaagtagcca	120
ggcttggggg	tgctcacgct	gtcgcccacc	cagtagggct	ggatgaagac	caccacgttg	180
atgatggcga	agcagatggt	gaagatggcc	cacagcacgc	cgatggcccg	cgagttccgc	240
						2.00

atgtagtgct cgtggtagag cttggaggcc tcctgcgagg gcagcatggt gcccggaggc

ggggccggcg gcggcggcgg ctggcggggg ccgccggccc gggacggagc gccgggctgc

cgggcgggag ctggggacgc acgcgagaag cggccctgag tcaaggaacc cgcgagggcg

gggcctgggg cagagctggg ggcgtctggg agctgctaag ggagagagga aggggtcatg

agagtgttga ggccgtgtct agggggactg gcaaaggtct cctactgggg ggcctaggaa

ggggccatga gaaagttggg gggcgcctag gatggggata tgagacctga agtgc

300

360

420

480

540

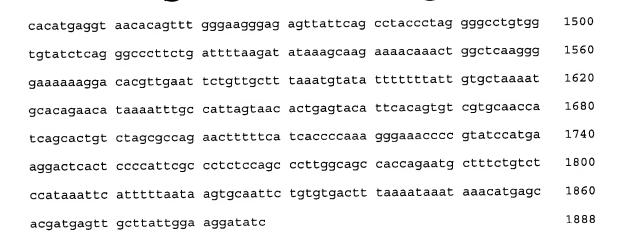
595

```
<210> 68
<211> 1955
<212> DNA
<213> Homo sapiens 4.D.07
<220>
<221> n
<222> (1)..(1955)
```

<223> a or g or c or t

<400> 68 60 atatctatcc atatctatac ctacatctac ctgtatgtgt gtagtgtata tatatacata ttatatgtgt gtatatatgt acatatatac atttaaacaa aaatttctcc ttcgtcctcg 120 180 aagcaaacaa accagcaccc tcgagtgtcc gccaggaggc gcagggggca gcgtgggacc tgcggtacct ccacggttgt agaggtgtag agggatgccg cagcgacgga accgggcttc 240 300 ttttttaaag aatcaatgtg agggaagggt gcagagccgc gttatttcag ggagacattg tcgcactccc cctcccacgt gtaggtagca tctggggtgc gtgcgccctg ttcgcagacc 360 420 ccatggagag acgctggcgg cggcagatgg ggctcctttc acggttgcag ccggcagtaa cccgaccccg ccggcgcaga gactgaagaa gcgcakggga cagcggcgag ctgcgaacaa 480 540 aagcccttgg cgcggggccg aagcccakga cgcggtgtga gtaaaccggc tcgggtaccg ggagctgcgg gaacctgggc ggccaggttc tttgcactcc aggagcccac ccactgggat 600 gctgtggggg aactntcgga gggcacccga rggcgggtat ctgaaccccg actggggtgg 660 atggtatctt tagcacattc agacttggag gagawycggk gcggtctgag artatccagg 720 780 caccttctcc atccccagca aaacamccgg tgggggtggw ggtgggggcg gaggcggcgt 840 gcagagccct cagtaagccc tgccagagct gctggagcaa gaatccatca cccctcccgg agaggccttt ggggacttct cccagccctt taatcacccg ggggccttgc gaccgagtct 900 cctttggcag gggaaatcaa ccataaactt cttyccytag gcaaatgggg tcccttggga 960 tgaacaggcc tcttgctttt ttgttcctgc aaagctgcat ccccagtagc ccgcctaagc 1020 tacaaacaaa tacgctaatc ctcccgggaa tcctccagcg cctccctctc tagctcctgc 1080 1140 ctgcacctgg atcttttcat cttaacttgc agcagaaagg ggatgcatct agcgggctag 1200 gcgcccagag gagcctcgcc acaggcctcc accccgcatt ccgggggctg agggagaccc aggetgetet etgaacacga gtgteegeee cacceemate cesgtyytgg egeteageet 1260 gggctttccg acatcggttt tatgatttac gtyccaccaa agcctctgag cctaatccga 1320 aagcggatta agttgggatg gggtgactat ggatgaggag gggggaagag ctctcagacg 1380 tattcctcga tgtccctcct tgtgatctgc agagattcca acaaaggacg gggctcagcc 1440 atggtggacc cagtgcctga agaagagaag gcaggagcgg aacccggcga ctctggaggg 1500 1560 gacgaggccg tggcgtccgt gcccctgat tcccagggcg cacaggagcc cgcagcctcc tcggcctcgg cctcggcctc cgcggcggtg ccccgcaagg cagaagtccc atgtgcagcc 1620 gcagaaggcg ggcggcggga gcagtccccg ctgctgcacc tcgacctctt caacttcgac 1680 tgcccagagg cggagggcag ccgctacgtg ctgaccagcc cccgctcgct agaggcctgc 1740 gcccgctgtg cggtcaagcc ggtggagctg ctgccacggg ccctggccga cctggtgcga 1800

gaggeteegg geegeteeat gegggtggee aceggeetgt atgaggeeta egaggeg	gag 1860
cggcgcgcca agctgcagca atgccgggcc gagcgcgacc gcatcatgcg cgaggag	aag 1920
cggcgtcttt tcacgccttt gagccccgcg gccgc	1955
<210> 69 <211> 1888 <212> DNA <213> Homo sapiens 4.D.08	
<pre>&lt;400&gt; 69 gcggccgcca gctcacaaag gatagggagg gatattgctc ttggcatttg atgggaa</pre>	igca 60
tctgctgcat cccattgggg tgttgcccag gatggattgg aaaagagttg gcaggaa	iggc 120
tgagctctgt gctcacaacc tggcttggtg gtggccgagg agcttggcag gagcaga	igtg 180
caggacctgg gaactggggg ttggtgcatg tgtgcacgca cgtgtgtgtg tgtgtg	gtg 240
cgtgctgggt gggtagggag gaagctgtga aaccacatcc cctcctctc gctgctg	gtgt 300
tgctgtgtgt ttcagcagca cgtgggtgtc accacacttc ctagcaggtg tcaacct	cca 360
agactgttct gggctcttct cccagttggc tgagttggag gtgggagtcc caactgt	ccc 420
ctgtggcttc cagagtggga ccttgctgtg ggataggctg gccaatggtg ctccct	ccc 480
tgtgaccctt ctgttgggtg ggtcacgagg aaggactgtg ggtgttgccc acagaca	aggt 540
ggacatgtgg caaggacacc ttgggacctt ctttctgacg ccccttgaag ggggcac	cttt 600
ctcagctttg agatgagtct ctgtggatgt gggaagttca ctatctcaag agcagca	agcc 660
ttggaaaatc caacacagaa ccccgagtag gggcgggaag gggtcctgtc ccgctc	actg 720
gctgcctggc agagttctgc acaaggaagc gcctgtgttg ctgtgggcgg aggaat	ggac 780
tgagggctac attcgcttcc tgttgccgct gtaactgctt atcacaaact cagtgg	ctta 840
aagcaacaga ggctccttcc tttacagtgc taagggtcag aagccgatca gtctca	ccgg 900
actaaagtca aggtgttggc agaatccatt cctgcctctt ccagctttgg gtggga	ggct 960
ctgctggagt tecttggett geggetgeat eceteeagee tetgeeteea teetee	taca 1020
gcctcctcct tctctgcagt cagatctccc tctgccttcc tcttttttt ttttga	gacg 1080
gagtcaccca ggctggagtg cagtggcaca atcttggctc actgcagcct ccgcct	cctg 1140
ggttcaagcg attctcctgc ctcagcttcc cgagtagctg ggattacagg catgtg	ctac 1200
tacacctggc taatttttgt atttttagta gagacagggt tttgccatgt tggcca	ggct 1260
ggtcttgaac tcctgacctc aggtgatctg cctgcctcag cctcccaaag tgctgg	gatt 1320
gcagccatga gccatcacac ctggcctgcc tccctcttaa aggacgcttg tgattt	gggg 1380
cccacctggg taatctcttc atctcaacat cttcagttac atctacagag tccctg	ttgc 1440



<210> 70 <211> 994 <212> DNA <213> Homo sapiens 4.D.12 <220> <221> n <222> (1)..(994) <223> a or g or c or t

<400> geggeegeta ggaaaagget eageteegge egeteegatt ageegtggee ttgetetgeg 60 agcagataaa cgtgacctcc gtggcctgtg gccagcctcg gccctctgga ggcggggctg 120 tgtgcggccc tcccctcccc agcagggctg agctcagaag cagcaggcag ccggaagggc 180 tgggcagtcc ccgcacctgt ccctgtgcca gtctggtggg tgttgtgtgt gcagggtggg 240 cgtgccggga ccctctggcg tggggctgtc tggcaaaggg cgaggggga gggggctgtg 300 360 cttcagcata gaagggaagg gcgtgtccag aagagggaac agaagagggt ccagaggccg 420 aaccagaaca cgtcccttca ctgatggaaa cttcccaccg cgctcgaatc aattcccaat tgctcgactc ctcgcacctc ccgggaggtc ctgtagaggc agcgctccct cccagcctca 480 540 cccgccggcc tgttcctgcc acagggctct gcccttcctg agctctccgc ccggactctc atcccggact ctcctcccca tctccttcca aagccagttc tttctcatta ctcagggctc 600 tgctccaatg ccacctcctc ggaggggcca cctcatcctc tgaacggcgc ccatccctcc 660 ctcctttctc ggngccagct ccattntccc cttctccttt ntcaccacgc ccacaactta 720 gaggcgcgtg tcccgtccct agaactgctg cggncacagg actnctggcc cttngcatag 780 gctggcacgt ggcacgttcg ccccagcctc gtacgcattt tgatggagag ttggaccaga 840 gagggcgcgg agcatgaatc tctgaagagc tgaggagccc aaatcagaag ctggtgagtg 900 agtttaatct gacttggagc atggagttat acgggagctg cttccagaag cccagctctg 960

cactgctacc ata	tatggca cggacgc	ttt agct			994
<210> 71 <211> 677 <212> DNA <213> Homo sa	piens 4.D.13				
<220> <221> n <222> (1)(6 <223> a or g	77) or c or t				
<400> 71	cattgag acaggaa	agc tattttaa	ıqa tggtgtggtg	aaaaaggata	60
	caagete tagetta				120
	ctgcaag ctcctat				180
	cttaaac tccaaco				240
	tggagat ttaaaat				300
					360
	caagcac tagaaaa				420
	ccctata aaactga				
	getgaet eeettge				480
gtttggtgat ctc	tgttaat ctctate	catg ggagatc	ata agaatccagg	gcaacagtaa	540
cagcttctga gtt	tttaaat taaaaa	aac agtaata	taa tccttaaatt	tttaaaatgt	600
aggacactaa aca	aagtaaaa totaaa	cca gagtaca	tct gacctcaaag	ttcatgggct	660
tctcacttcc ctg	ggcca				677
<210> 72 <211> 435 <212> DNA <213> Homo sa	apiens 4.D.47				
<220> <221> n <222> (1)(4 <223> a or g					
<400> 72 geggeegegt nee	ectetege eegnaa	agag gactgga	gaa ggggctgggg	tggaggtnnt	60
ctctgtgtgt gt	ctanggtt gngggc	agga gaggtta	att ctattaagan	ntcatcaatc	120
anccngtgtg ca	cttttcgc tcgaca	negg ntectne	tac ttnanagcaa	gtctggncca	180
gctgggatcc ga	ccagaaac cgcaag	cgna ggagacg	cat gancgnaggo	tgagcgctaa	240
ctgaaggcnc ga	cctgagcc ctgcag	cctg ctgggga	gct gcgcaaccac	ggacagcagt	300

tcggcaatac acggcctggr	ctgcatggcc	cccgtcacca	cctcacgtgg	gaagccagca	360
ctgctgccgc cagccctgcc	gctgccctca	gactnncaag	gcgnccaggg	tcctcccaac	420
gegeetgeee cacae					435
<210> 73 <211> 2343 <212> DNA <213> Homo sapiens 4	ł.E.53				
<400> 73 tggccaggtg aggtcaggct	ctgtttcttc	cgagctacca	tcctctacct	gattcctcac	60
acctttttct tgttaggcgc	agctaagaga	cagagagaga	gagagagaga	gagagaga	120
gagagaagcg actgaaacag	g agagtaaatt	ctagtttctc	ctttttagtc	tcttttcttc	180
tgccctttgc tctgctagt	tatctgcgtc	ttttctcttc	tcgcgctgca	agagtggaaa	240
actcgtgctc agttctagg	c aaacattaac	cccgggcgac	gtttccaagc	gggagacaaa	300
ctctagagag tgagaagcg	a gatgcgaggg	caccaagggc	aagaaggggg	ctcggggtac	360
gccacgttgg cgggacgcc	g ccgccgcctc	cctctgctgc	gcggcctgcg	ccgggagcct	420
ggtgggggg gcaagacga	c agaccccgcg	cccgggcctc	ccaccagtga	ccacctccct	480
cgcagcttgg gctgatcct	c cagacagcat	gcaacggtgg	ggagggaagt	cccctgactg	540
ggcgggggac ctagcggct	g ctctgaaact	ccgaacacct	gaagaggagg	cgcggaaggt	600
ccagccgccc aagactcgc	a ctttcccctc	ctccgcagcc	cgggcaggtt	accgtcctgg	660
gcctgggtga gcgcggagg	g gatccgggcg	ggagctgagc	tcggttcccc	aggcctgaca	720
agtggccgcg tggcacgac	c aaccccgggc	acagggctgg	ggctgctccc	caaggtgggg	780
aatttaattc tcacatttt	c gcactaccct	gacggagctg	gacgcgggaa	gcgggaaaga	840
cccgttcctg tttgcagtg	c ccgagggca	ggacacctac	cagaagggct	ctatcacagt	900
ggtgttaggc cgggcgcag	t ggctcacacc	tgtaatccca	gcactttagg	aggccgaggc	960
gggaggatcg cttgaaccc	a ggaggcagag	gttgcagtga	gccaagatcg	ccccactgca	1020
ctccatcccg ggcgacaga	g ctgtcttgaa	aaaacacaca	aaaaacaaa	aacagtggtg	1080
ttagagggat gggattata	g gtgacatgac	tttcgttttg	aactttcctt	aaccttgcag	1140
gggcagccgt gccctgaaa	a cgcctgtgat	ttggagtaga	gggtccaggc	gcagtgtggt	1200
gagtgaccct aggcaggtc	a ctagttcttt	ttcagccttc	actgaatcct	ctcttacacg	1260
gggatgttac ccccaggtc	t ccgtgtcttt	: cagggagaaa	ttagttcatg	agttagatgg	1320
tgcactatca atcatcctt	t tattagaca <u>c</u>	g aaacaataag	ı tttgaggaag	aggacgtcta	1380
ccttacaggg ggtttaatt	t tcagcttctt	: tgagataaaa	ttcattgaac	ggtgttttac	1440

gtgcgcgcct	tttccaacag	accccacgcc	tattcccagc	gccagaggcg	gacaaccgcc	1500
ttactgagat	acagagacag	gtacttcctg	aggcacttca	gtccagttcc	actgggttta	1560
ctacaactaa	taatgactgt	ttctgtttac	taggtattag	gcgatgtgtt	ttaagtaaat	1620
gaattgtctc	taatcctcac	aactctaaag	caagttaggc	gtcacccgca	ttttacaaat	1680
catagcgccc	tgctcaccat	atctggaatc	ttgcctcgcc	ccgagggttc	taattttcac	1740
tttagagagc	tgagcaagat	gattgcccag	cgctaactcc	gtgaaatccc	tgggactgaa	1800
aatcacaggt	aactcgccag	agtttttcaa	ttttaggcct	aggagattat	gcaaagattt	1860
ccttcaagta	aacgctgttc	tctggggcct	ctgggatcta	cagtcggaga	aggggaataa	1920
gtcccgggcc	ggtgggggat	gggtgggtgc	agtttcctaa	atagaggaaa	gccactttca	1980
ttcaaagggc	tgtggaactc	tggctagagg	tgggtttctt	tgcagttaat	catctgcaag	2040
gctctttgga	tgcctgattc	cagaaaccca	gaactcacac	ttagggtcac	aaaatccagg	2100
gcatttattt	gccgagcccc	atggatgtta	tccctatgga	tgcaccccgc	ccctgtccgt	2160
tctcctttgg	agcagaacga	aacccattcc	agagcttttg	caggaagtct	tcaggccctt	2220
gegteeggee	cctttagaca	tcaaagcccc	ccctgagagc	aaaggacttt	gaaagatagg	2280
aaaagctcag	gatccttatc	gcgtctctgc	tccctcccga	cctagtcgta	aattccgagc	2340
ctc						2343
<210> 74 <211> 507 <212> DNA <213> Hom		.F.15				
<400> 74 tacgactcac	tatagggcga	attggagctc	cacgcggtgg	cggccgcggg	cagtgcggac	60
caggcggggg	ccctgtggct	gccggccaca	tcccggagca	acagcagaaa	caacggcagc	120
agcagcagca	gcagctgggg	cccgggtccc	gggctggtcc	gagcggggac	atgagccatg	180
gcgtggtgag	ggcggcaaag	ggtcgaagtc	caggaggagg	aaggcgagcg	ctggcgcacc	240
ggaggctgcg	gactgaccto	gcggcagtag	ggcgcgcggg	gagagcccgg	gcagcagggc	300
gctggatacc	gaggtccgcg	g cggggcgagg	ggcttagcgg	g agcaggcacc	cgggcgcgcg	360
gtccgtgggt	accggtggc	: cgagcccccg	gccagcggtc	: acagccgtcc	ggagcagcgc	420

<210> 75 <211> 446

tagcccactg cgcgcccggc ccggctg

agagecgage egagecegag teggegeget geettggegg actegegetg egaaagtttg

480

507

<212> DNA <213> Homo sapiens 4.F.17	
<400> 75 geggeegeac acaegaggge cegtegegee eccegeeetg eccegeeteg ecctecaegt	60
ccctgcaccc ccgagtcgca ctaagaaccc agtccccgat cggtttcctc tacgccgtct	120
gagcagaaga gagtgggaac cggggtgacg gataaggggg gggcgcccac gcgacgtcgg	180
ggtgcatggg agcgcgggg aggcgctagt gggtgcacgg ggcgtgaggg ggacacagcg	240
cgggcgtggg gatggccact gcgcggggag ggttctgcct ggagaaggag ggatgggagg	300
aggttggggg agcagggcgc gtggaggagg gaggttggac gtgtgtacag cgcctgggga	360
cctcgctggc cccttggtgc ccccaggact ctgaggcttc tcctttcggc ttgaaatgtt	420
tttcccttcc tgcttttcaa atctgt	446
<210> 76 <211> 424 <212> DNA <213> Homo sapiens 4.F.22	
<400> 76 gcggccgcct tgaaggcgct ggacgggatg gtgctgaagt cggtgaagga gccccggcag	60
gtgagctcgc ggcccgccag cccgctgccc acgcagtagt ggaagaggcc gaagtagcca	120
ggcttggggg tgctcacgct gtcgcccacc cagtagggct ggatgaagac caccacgttg	180
atgatggcga agcagatggt gaagatggcc cacagcacgc cgatggcccg cgagttccgc	240
atgtagtgct cgtggtagag cttggaggcc tcctgcgagg gcagcatggt gcccggaggc	300
ggggccggcg gcggcggcgg ctggcggggg ccgccggccc gggacggagc gccgggctgc	360
cgggcgggag ctggggacgc acgcgagaag cggccctgag tcaaggaacc cgcgagggcg	420
gggc	424
<210> 77 <211> 558 <212> DNA <213> Homo sapiens 4.F.6  <220> <221> n <222> (1)(558)	
<223> a or g or c or t	
<400> 77 geggeegeag eteaceactg geetagagat geeetttgeg aggeggeage aactgacaag	60
atggtcgcgg gtcgccgcgt ccggagccgc ccaccaggtt gccaggagga ggcgggagcg	120
gggatcaagc ttatcgatac cgtcgacctc gagggggggc ccggtaccag cttttgttcc	180

ctttagtgag	ggttaatttc	gagcttggcg	taatcatggt	catagctgtt	tcctgtgtga	240
aattgttatc	cgctcacaat	tccacacaac	atacgagccg	gaagcataaa	gtgtaaagcc	300
tggggtgcct	aatgagtgag	ctaactcaca	ttaattgcgt	tgcgctcact	gcccgctttc	360
cagtcgggaa	acctgtcgtg	ccagctgcat	taatgaatcg	gccaacgcgc	ggngagaggc	420
ggtttgcgta	ttgggcgctc	ttccgcttcc	tcgctcactg	actcgctgcg	ctcggtcgtt	480
cggctgcggc	gagcggtatc	agctcactca	aaggcggtaa	tacggttatc	cacagaatca	540
ggggataacg	caggaaag					558
<210> 78 <211> 865 <212> DNA <213> Homo	o sapiens 4	.F.69				
<400> 78 gcggccgcag	cgagttttct	ggcagcgcta	gcgccgcggg	gcctgggttc	ccgggttccg	60
gtctccgccg	gctccgggct	cgcccccgcg	agttggccgc	accgttcccc	cgcccgcggg	120
gcagccgctc	ctccgggagg	ctccggcagg	gaccttcgcc	ccggcccccg	agcggcagtg	180
cggctccagc	tggaggcctg	gcccgggaag	caaagtgaaa	ggacagaggc	ctccttcctc	240
gccagccgcc	cgccgcgcct	ttcccagctc	aggccggcgg	cccgcggcgc	ggagggagcg	300
aaagagtcgg	ggcctgcccc	ctccaccgcc	cgcatctcgg	ccgccgcacc	cgggtccgcc	360
ccgggaggcc	ccgcgggagg	gaacccccgg	cccgctgggc	gcttccgcac	tgacgccttg	420
gggccgcgcg	ccccgccc	ttactaccgc	tacacccgct	gggcccccga	ccccgctccc	480
gggctgctgc	cagcgccgtc	ttcccccgta	gaaacttcgg	agacacccgg	gaagctgctc	540
tttggagttg	gggaaactta	ggaagaatgg	gaaaagccga	ggaagtcggg	gaggaccccg	600
cagttgcctt	gccctcggcc	gaaattcctg	tgcaattgga	cgggaagcct	gccacgccca	660
gagagccacc	cggtggcacc	ccgttgggga	cctgcggctg	ccctaggctt	gagctggcga	720
ccaacggcgc	ataccccggg	cacccctagg	ggaccgtgcc	cggcccggct	tgggggctcc	780
taacgccagg	cttgtgagct	atagggtgga	gagtgggccg	gctcttaagg	ggaaaaattt	840
gcggcctttt	accaggcaca	gccag				865
<210> 79 <211> 983 <212> DNA <213> Hom		.D.9				
<400> 79 gcggccgcag	ccagegeege	ceeteeegge	cgggcgggcc	ccaaaagccc	: tttctgtcac	60

cgcaccaggg	cgcgaccggg	tgatgcattt	ccacaccagc	ccgcccaaac	ctccatggtt	120
ttggagctcc	cgggcaggcg	gtggaaactt	ggcgcaccgt	gcccactctc	cggcgccgct	180
ccgacagccc	gacgggtccc	gcggccagga	agccactcgg	cgcccctcgc	cgtcactcga	240
cccccggccc	ctttcggact	ccgatcctcc	cgtccccagg	ccacacggcg	cggaaagggg	300
atgccgagcg	ggacgcgcac	gaccagggcg	cccaggacga	gggcgctgga	ggagactccg	360
ggcagggacc	ggggtcccag	gggcccgggc	cggggctcaa	cacccacccg	atggggtgcg	420
ggcccgacgg	ggcccggggg	tgggagtagg	ggcggcgggg	gcccgcggag	gaggagtggg	480
gataggccgc	gcagggggtg	cccgggaccc	cgggcgcaag	ctgggaaaga	ggcacgcggg	540
ggcggcgcgc	cggggccggg	acaggcgccc	gtcctcacct	gccgggcagg	tgtcccgccg	600
gcgagtcgcg	cgcgttgctt	tccgaggtgg	aactgtcgtg	gtccacggcg	catggcgcgc	660
tgaaggcagc	ggccagcagc	ttcataaggt	cggcggcggg	gcaggtgccg	gggccgggtc	720
ggaggccacg	ccggggccct	gggctggggt	cggggcgact	agcgggctgc	gagcgggttc	780
cacgcgcgcg	gttcaacggg	ctgcacccgc	gccgcaccgt	gccaacactt	cgggcgggcc	840
ccgctgaggc	tccggttgcc	cgcactagga	ggcgagggcc	cccgcgtgca	agccgccggc	900
ggcgggcccc	ggttgccacc	ggccccagcc	atgggtgggc	tccgggttgc	tttcccccc	960
tgccccctag	ggaattgagc	cga				983
		.E.2				
<400> 80 gcggccgctg	gtgacctccg	cccgcggtca	ctcgacgccc	agccttggcg	cgtttgcgca	60
actgcttttg	tecegageet	tcattctggg	cgcagtcccc	tctcccagtc	cccctgccgc	120
ggcgcctgga	actctcctgg	tggctgtaag	attttcctac	cgttaggtcg	tctgtggcga	180
ccgccaggcc	tgccccacat	cgctagccgc	cctgtctacc	cctcagcctc	ccagccacta	240
aactcgctgg	, acaaccttac	gctagtaaca	gtttttgagt	ctcagactca	tctgtgaaag	300
ggcagtcata	ı tttgaggact	ccaaatgggc	tgcagtgcgt	aaaccaccat	gcgatatttg	360
gttgctattg	g cccacctcag	cctgtggcca	atgtgtctct	gtaggaacag	cactagattc	420
tttggggttt	: tt					432

<sup>&</sup>lt;210> 81 <211> 746 <212> DNA <213> Homo sapiens 5.E.25

<220>

<221> (1)..(746) <222> <223> a or g or c or t <400> 81 geggeegegg gggegteagg teettgegee teeteeteeg getetteece eageetetge 60 ggggcgtcct ctcccacctc cggggcccac tcctcccccg gagagccccg gggcgcatcc 120 tcaaaagcat cctcctcacc ctcctcatcc gtgtccccag cccctcgcac gggggctccg 180 gccgcttcct cccccggccc ggcctcggga aatgggaaag ccgtggagga gggcgagtct 240 ttggccgcgg gttgcgctgc cgggagactg ggcgcctcgg agaccgggag gccgccgggg 300 gacggcggtt gctggggctc ccggggctcg gcggccaggc tctcgggcag gtcggagagc 360 geggacageg cetgeteggt gteeggactg ceeggggeet ceeeageece geegetegge 420 cccagcagga accggtccag gcccaggaag gccccgggct gaggggagac ggcagtgggg 480 ggcgctgcag gctcctcggc gccctggagc tgctgctgct gctgctgttg ctggagctgg 540 agctggagct gctgctgctg ctgctgctgc aggcggatcg cctgctggat gtctgaaagc 600 aaatcctctt gctccgtagc cgaatggaag ctatagatgt ccgtgtccga gcccgagctg 660 gtcctttgtc catcctgcgc ccctgctgca gtttncacat cctcggcgat cggccggccc 720 746 ccgaccctag cctcggcagg cccagg <210> 82 <211> 617 <212> DNA Homo sapiens 5.E.4 <400> geggeegeg geeggtgttt eaggeagete ttgggegeeg gegggetegg ggegggegee 60 gtggagggct cggtcccaat tctctcgggc tcggtccccg ctcctctctc gggctccgtc 120 180 teegettete tetegggete aggegeegge eettgggggee eetteteete ateegggage acgggcggcg tcggctccgc ttccttcggg acactgcgtt ctggcccgtc gcgagcagag 240 ggcgcctctg aggtggcggc ggggtcagtc tcggggggag tcgtgtcccc ctcagggatg 300 gcggtgggaa acgggctcgc gacgtcttcg ggagcacaga ccacctcctc cgccttgtcc 360 gtggccgggg cacacgggcc tgcggggggc gcctccccat cctgctttcc gccgtcggga 420 ccgggattcg gggggccctc cggcggggac gggggctcca cgcggagagt gggggccgac 480 tegggetegg egageteegg ggtggeeggg eggettgagg ggteeteece ggggaegeee 540 ccctcctcca cgctggccgt gagcgcggag gagtgctgca ggcgggcgcg tctggcacgg 600 617



gcccctccgg gtggcgg

<210> 83 <211> 1840 <212> DNA <213> Homo sapiens A.2.F.45

<400> 83 ggcgcgccga ggcgcaggcg cggagaggcg cggcgctctt ggggagacgc ggcgcagggc 60 120 atagacgtac gccggcgcct ccccggaggg gaggggtcgc tgggcgggcg ggagtgaggc gcggcgccgg cgcagagacg cacgtcgctg ggctgaggt ggcggggagt gttgcagtcg 180 tacattcgcg cgccgccggg cggggagcgc gggggtggcg cggtgcaggc gcagagacac 240 acgtacccgg cggcgcagag acgagtggaa cctgagtaat ctgaaaagcc cgtttcgggc 300 gcccgctgct tgcagccggg cactacagga ccagcttgcc cacggtgctc tgccattgcg 360 ccccctactg gcgactagga caactacagg gccctcttgc ttacagtgct gtccagcgcc 420 ccctgctggc gccggggcac ggcagggctc tcttgctcgc agtatagtgg tggcatgccg 480 540 cctgctggca gctaggaaca ttgcagggcc ctcttcctca cattgtagtg gcagcacacc 600 cgcctgctgg cagctgggca cactgccggg ccctcttgct cgcattgtcg tggctgcacg ccacatgcag gcacatgggg actacgcagg gccctcttgc tcccggtgtg acggctggcg 660 toccatattg gocacotoot goaccactta aagtoagago gocagttatt aatooccato 720 780 agttctgtaa attaaaactg aaaaggagct attactgcgg agagctgatg tcccagttat taacttggaa gacagctttt caccaagagg cagtacaaag atggaagata acttcattga 840 aaagaaatac agtgtaaaga gcttattgta caaaaatagg gaggagtagg ctgatactgc 900 atgaaaacag cctaagagtc ctgtgcaggg atttttattt tggacttctt cacattccta 960 1020 cctctgtctc aagtctccgc ctgttttctt tggttttcct gctactgcct taggtccccg acttgcccca cttagccttg tgggacctcc tcacttgatt gaggtacatg tgtggtgatc 1080 1140 aatccgaatc cactctggca ccagcctcct tcccaccata ccaggcaggc tgacagcggt cacgtttgta tctactgcag ctgcctcttt tgaatgtctt tctctgcctt aatctgtact 1200 tatggtgcca ggtttctctt aagaatgtcc cctttgtcct tcttatcagc atgtagctag 1260 caatattctg acatttttat tgcagaatga atgatgattg gggcttcttt tttttttt 1320 tttttgagac ggagtctcac tctgtcaccc aggccagact gcggactgca gtggcgcaat 1380 ctcggctcac tgcaagctcc gcttcccggg ttcacgccat tctcctgcct cagcctcccg 1440 agtagctggg actacaggcg cccgccaccg cgccagctaa ttttttgtat ttttagtaga 1500 gacggggttt caccttgtta gccaggatgg tctcgatctc ctgacctcat gatccacccg 1560 cctcggcctc ccacagtgct gggattacag gcgtgagcca ccgcgcccat ccgattgggg 1620





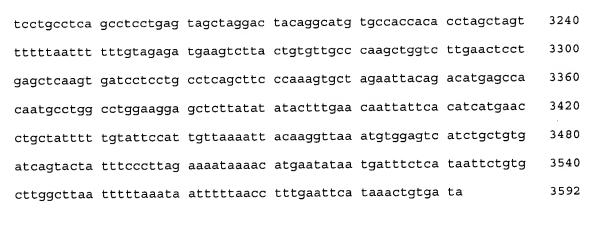
catcttaaga	gaagttctag	ggtgtttctg	cgtaggtacc	tcttctccct	cctaaccaca	1680
attgacaagt	gcccatccac	tccagcacta	gagatgctac	taatatgtgc	atttttggtg	1740
gtccctccag	gtgagccttc	acagactttc	ccttttccag	gageteeece	tcctgttcat	1800
gtctagctag	ctatctactc	taacagagcc	cactatcctg			1840

<210> 84 <211> 3592 <212> DNA

<213> Homo sapiens A.2.F.50

<400> 84 gccgaggagg cggctccgac ccaggtcgtc gcagcagcac aggaagctgt aacacaggta 60 agtgcaggag agcgagagcg tgaaggcgaa gagcagcctg cgcgccctcc gcggctgagg 120 180 tggcccgcg cggcccagga ccctataggc catggctcca tgggcccgcg ccgggggtca tggtttccga gggggcaccg gcggctgagc tgctgtggcc ctgcggtcgc ctagagggct 240 300 cgcgtggcgc tgccacggcc acgcgggtcg ggcgttgggg gcgccgtctt ctccgggggc tgctgaccag ggtgcgcaca gtgccagggg gtcccggggg cagcggctcc tcggggaaca 360 420 ggcggttgca tttccagcat ctcccggtcc taggcgatgg ggctccgggc agccgggcgg ctcgggcgct cccaggctct tacgtgcgcc gggttcggag cgcgcccagc gcccgaagcc 480 ccattcctga tcctcggagc gccgctcacg aaacgctcgg cggcggcgcg gctgtgcggg 540 ctggcgggtg gaccggacgg tggcgctggc gccggccgcg atctggctct tcgggaaatg 600 660 ccgagcggag cgcgctgccg gctctattta aggagtggcc tgacgtcagc cgcgcgggtc 720 ccccgagccc gcgccgcgcc cagggacctg gcccgccccc tgcgccccca ctctcttacc 780 cctcccagaa acacagcacg cgggccctcc ccatgcaggc cactccctac ggagccccag gccagctttg gggcggtgaa acgaaggtgt caaggcatag tactcctccg ggaggctgga 840 900 cacccccacc acgctggcct ctcgacatcc agggacacga atccaggtcg agatcgcgcc 960 gacatgcaga ccagacagac ccagacgcag acgcaggcac cctgccctga tgcgcggtcc caccaccetg accegeacae geacgeacag geacagaage acaegegeee tageeeggae 1020 acaccccac acccacgcgg gggtggggag gagaagtccc ctaacctggg cccagataca 1080 ccgacaagga cactccccc gctctcgaca tctcgccaaa tggacacaca cagcccggaa 1140 tcggacaccg agcgcacgca cgccctggac tgggacacgc gctgtagacg ggatgggtgg 1200 aggageegag egtgagtgag atteegtgae tatteaceea gettettage eeccagegeg 1260 ctgactcaca ccccggcggc tcgctctgtc tcgcacctat gaggcacgcg cgcaccccaa 1320 1380 cccattgtca ccccacctct ccccgggcct gccggagagc gagccccgga gcggcagact

ccgcgtcagg agggttcctc tcttagcagc cgccgcctag cggtagactg ctccccgggg 1440 agctgtccag ggtaccagag ggtcgccgag ggctgagtga ggagggcttc ttcacacaga 1500 gacactagga ggaggaaaca gagtacaagg agaacgtatc caggagcaat tccacttcga 1560 atgattecta agtgaatgee tacaggacag tteteggtga ceatgtecag aacaggeata 1620 agtgacgatc cccagtactt ccctgaggga ccacactggt accttggatc agaaccctgc 1680 atcagaacag gcctaaatgg ccatggctaa gaacacggct gagttgtcct tcaacagcaa 1740 tgccaatgcc aattcaccat gtccgagtgt tcacaaggtg agtgccctcc accaccaccc 1800 agccatagaa tgtctagatg accaccatga ccccaccct gatcagggta taactgactt 1860 1920 ccttcctcag gctgtaaact gatcattagg ttctgtggat cttagcccaa accagaaaat attttgtccc caaactagtc ccatccctag aaaccttaaa ccaattctac ggcagataat 1980 2040 aataatagct gccaactttg tatcaagcac ctggcatggg ttaactgatt aaatattcac 2100 aacctatgaa gttgttacca ttaccctggc atcactttgc tgtcttaatt ctaatagtag ctagcattta ttgagtgctt gttttatggg agttatgcgc taatcacttg acatgcacta 2160 2220 cctcatttat ctttggagat aggtattatt gtaatttcta atctacaggc agtgataaga 2280 agatttaaca aacatataca cagtaactgg cagagctggg attaaacccg ggcagtcttg actocaagat toaagotott agttacagoa otttgcagot tootaactto otttgaccat 2340 2400 tattcatata attccatcct aggeteetet eetggatgta agetaatttg tetatgtete 2460 ttctaaaatc tcacacctgg gactgcgcga ggaatttcag atatggattg aaaagttcaa caggactete acctetett tgtaagttet atttetagta atgecaceta agacteeatt 2520 2580 atctttttct tgtggctata tcacactgct gacatctcaa acttgcagcc aagtaacatc 2640 tctaaatgtt tcttacaagt gctgctgatt aaggcacagc taccccatac tgtgcttgta 2700 cagtgggcct ttttggaccc aatgtgtagg tccttataga tttgacttga ttgcatttca 2760 tcttgtctca tcagttcgct gccctagttt tttttaaatg tctatttgaa gtcaaaccac 2820 gaggtagctt tcatttattc aaaaagaaaa agtagaaaga ttgtatccca gctttaccct 2880 ttattccagg tgtactttgg gcaagtggac cccctttaag cctcaggttc ctcagctgta 2940 aaatgggacg ctatgattca ccttaaaagt ctctcaaagt ttagatgttg catgattcta 3000 tgattccatt acccaaagca tgaaccactc acttggcatc atgtaatttc cacagttgat 3060 cacaatttaa ttaattcctc attctaattg ttaataaaaa tgtcaaaaca aatatactta 3120 aaggagttot tottottott ttgggtgagg ggaagtgtot cactotgttg accatgotgg catgcagtag tgcaatcata gctcatgctg cagcctccac ttcctgggct caagggatcc 3180



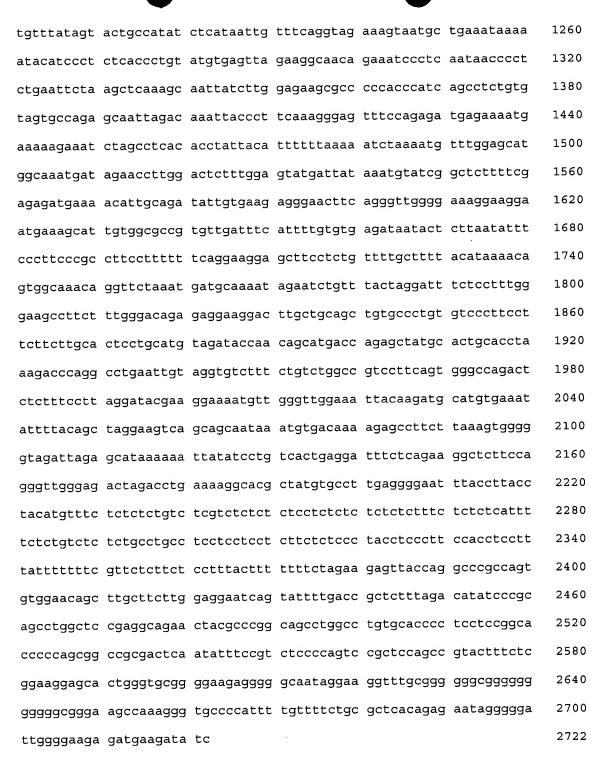
<210> 85

<211> 2722

<212> DNA

<213> Homo sapiens A.2.F.67

<400> 85 cgccgccgag gacactcggg cgcacacccg ccgcgctggc gtcccccacc cccagcccaa 60 acaaaagaca agccttgggg tcgtggcctc gctgggccgg ggcgccccga gccggccagg 120 gcgccctctg gggccagagc tccatggttt gcctaaggca tagcttcttg gcggtaggcc 180 gcaagcggcg gggagacgcc aggcagggct gggccgccca gaggtccgaa gatgcctcca 240 gtcgccgccc cggggaaggc gcgggcgacc tctgagtgtc ccggtaacgt gtgcctttgt 300 tccccaactc aggtgaaaat ctggtttcag aacaaaagat ccaagatcaa gaagatcatg 360 aaaaacgggg agatgccccc ggagcacagt cccagctcca gcgacccaat ggcgtgtaac 420 togoogcagt ctocagoggt gtgggagooc cagggotogt cocgotogot cagocacoac 480 540 cctcatqccc accctccgac ctccaaccag tccccagcgt ccagctacct ggagaactct gcatcctggt acacaagtgc agccagctca atcaattccc acctgccgcc gccgggctcc 600 ttacagcacc cgctggcgct ggcctccggg acactctatt agatgggctg ctctctctta 660 720 ctctcttttt tgggactact gtgttttgct gttctagaaa atcataaaga aaggaattca tatggggaag ttcggaaaac tgaaaaagat tcatgtgtaa agctttttt tgcatgtaag 780 ttattgcatt tcaaaagacc ccccttttt ttacagagga ctttttttgc gcaactgtgg 840 acactttcaa tggtgccttg aaatctatga cctcaacttt tcaaaagact ttttcaatg 900 ttattttagc catgtaaata agtgtagata gaggaattaa actgtatatt ctggataaat 960 aaaattattt cgaccatgaa aagcggaatg tttctgaaaa atacttcatt ctgcccctct 1020 gataactggc tagtgaagtt ttattgaagg caactaaaga aggacaagct ctgcagagat 1080 ccaacaaggc aaaaaagaaa acagaagtcg gggctctatg catgcagact gtatatgtat 1140 atatgttcaa tgctatactt tgtgtgtgtg tgtgcatata tatatataat atatatggca 1200



<sup>&</sup>lt;210> 86

60

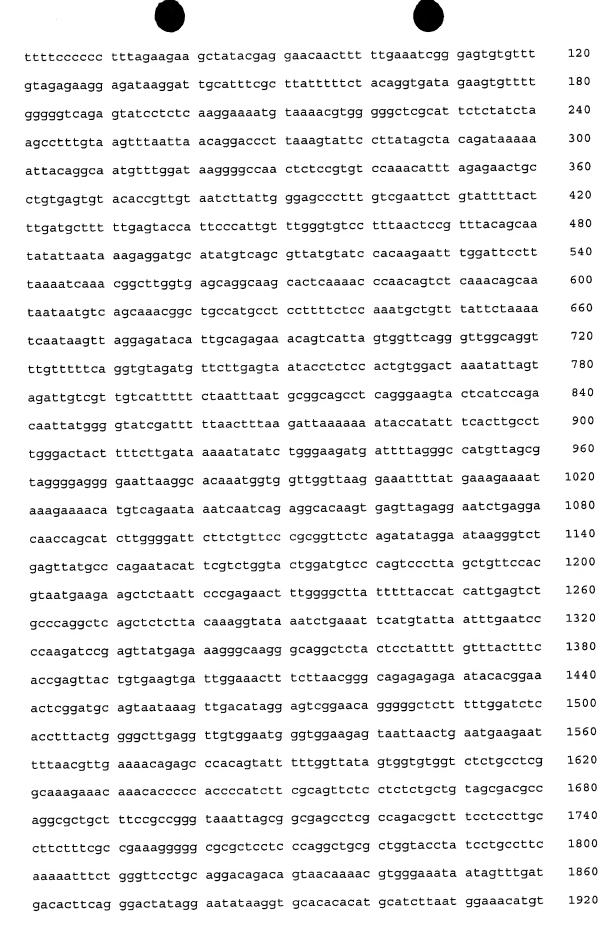
<sup>&</sup>lt;211> 3366

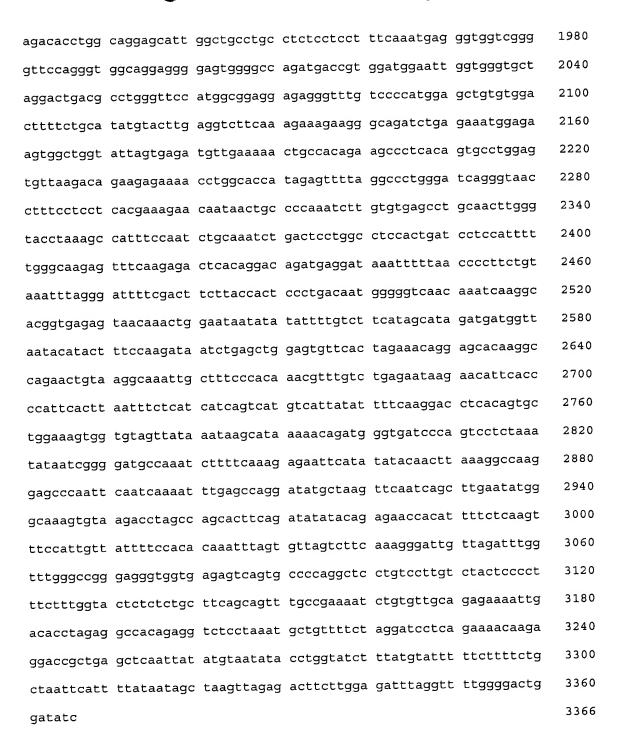
<sup>&</sup>lt;212> DNA

<sup>&</sup>lt;213> Homo sapiens A.3.F.38

<sup>&</sup>lt;400> 86

ggcgcgcctc cagttccaag gccgagctca ctttcaacag ctctggaaat atgaatgtat





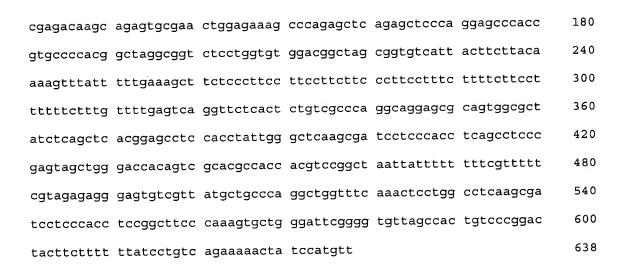
<sup>&</sup>lt;210> 87

<sup>&</sup>lt;211> 638

<sup>&</sup>lt;212> DNA

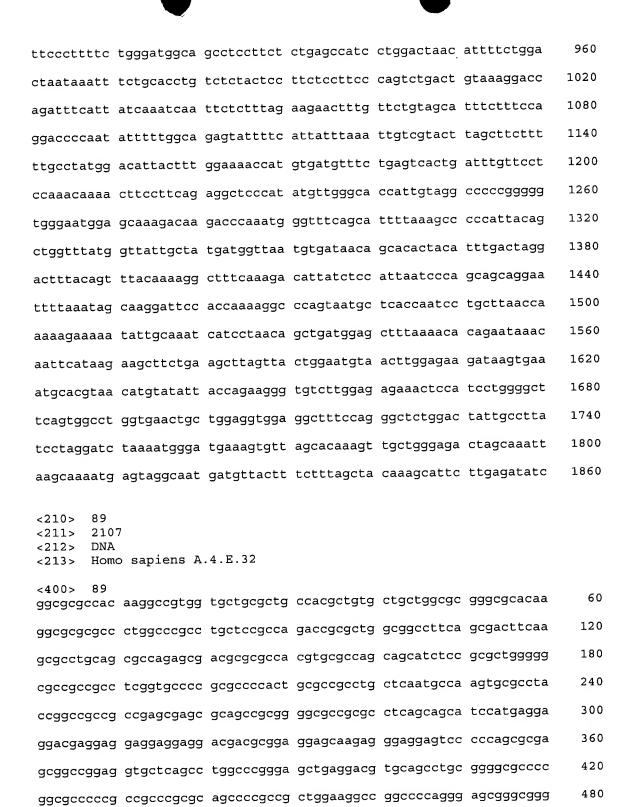
<sup>&</sup>lt;213> Homo sapiens A.4.D.30

<sup>&</sup>lt;400> 87
ggcgcgcctc gcccgagatg cccctgcgtc cgcctggcca ggcctggggg ttacccgacc 60
cgggttctcc cttcgctggc tttgcgcccc ttcacacctc tgcggtgggg acggagctgc 120



<210> 88
<211> 1860
<212> DNA
<213> Homo sapiens A.4.D.36
<220>
<221> n
<222> (1)..(1860)
<223> a or g or c or t

<400> 88 ggcgcgcctg tccccaccta atgccacgat concecetce eccaccetne egcactgect 60 120 cccttgcgcg tgtaggggag atccctgacc ttgtctgccc agctgcaggc cacttgccca ggcggcccct cccttgttgc cacctcccgc ccagctcacc aggagcgtgt gccctgttgc 180 tactggcaac tgcctgtgcc taaagctcag cccccaaact ggcttaatgc tgattgatgg 240 tcagaaatag gatattttct ggaacagagc ggagcgctgg tgcaaggccc tctctgctgc 300 tgagtcctag ggacctcccg ggtggcaggc cttcctcctc ctctccttt ggccccaccc 360 accetacact accectcaga gaccaacggg ctetteggac atceteatet caggitaagt 420 gctgagccag caagccagtg ttcgctttct tgctgagtaa caggcagcca ccccggaatt 480 tctcttctta tccttgaggc ttctgagttt tatgaatgag gcccgtgttg ctggacgcta 540 600 ccacttccct ttttattttc atccccacta acttgttcac tcgttcactc ctccttatac ataggtacct aaaatagact acccctctag taaccagaac tattcctgca aacgcttaca 660 agagcatttt ccagaaataa atcatttcat atcagtatcc cttcctcagt catttcccgg 720 cttcatgcca cctccctcct aagacacaga attggtcatt tccaccactt taaagacaca 780 gtctagataa aaagcctgca tttataatgt tctttgcagg agtagctttt gcctattttg 840 900 tgggggtttt gtttgttttt tgttttctgt ttgatactcc ctctcaaact gcagcctccc



ccaggcgcgc tgagagccga aggacaggac tcgcagcccc aggcccgacc cgccagactc

acageeteca acceeggeee tgeeegette ggetgeeeeg geeeeeggee egtgteteee

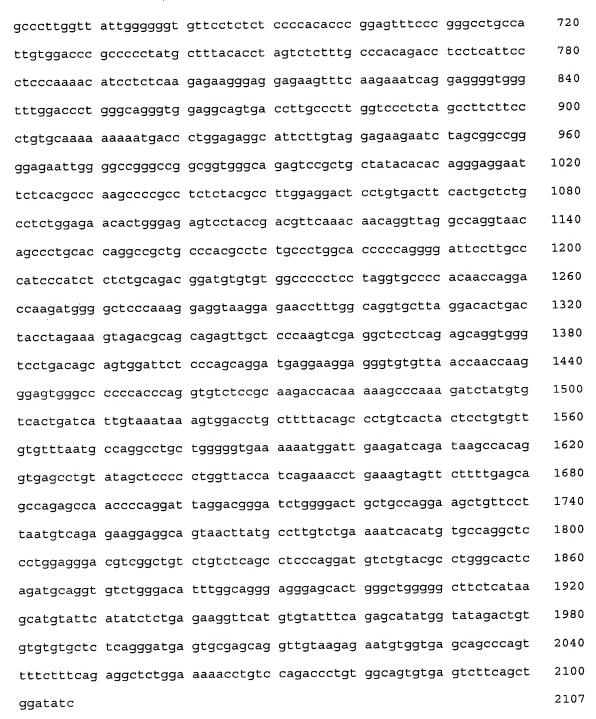
ccgtggtctc cgtgttgtcc gccccgccgc ctcattttgg ctcaaggtga tgcctgatac

540

600

660





<sup>&</sup>lt;210> 90

<sup>&</sup>lt;211> 498

<sup>&</sup>lt;212> DNA

<sup>&</sup>lt;213> Homo sapiens A.5.E.28

<sup>&</sup>lt;400> 90
ggcgcgccgg agttcgggct gccggctcct tagccgcggg gcgggggaga cgctcgggga 60
aggggagagg cgcgggcggg tgggaacggg cgggagacga gcggggacgg ggagacgcgc 120





cggaggcccg	gagcccgcgc	atgctcagtg	cgcggccgga	ggaggcgagc	gctggggacg	180
cagcacctgc	cccgcgcggc	cgagaggcgg	cagccccagg	tccccagcgc	gcgaaattag	240
taaagggcgc	ctggcccgat	tctcaggcaa	gaggagatta	tcagccggat	tcccgtgcgg	300
ggacgtaggg	gttgcgttgt	tcagcggcca	gggatgcgcc	gaggcgatgt	ctcctccctt	360
tacaacccga	gtatcggggc	acgaggaggc	gcgaccttcc	tgggtaccca	aacctctggc	420
ctccgggaga	cgcggaattc	gggggatcgt	taaggcgccc	tggccaggga	aacagatgct	480
tetgegtetg	ggctgaaa					498